

SYNTHESIS, STRUCTURAL CHARACTERIZATION AND ENZYME INHIBITION STUDIES ON 5-(2-NITROSTYRYL)-1,3,4-OXADIAZOLE-2-THIOL DERIVATIVES

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ABSTRACT

In the present work, *S*-substituted derivatives of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**) were synthesized by successive conversions of 3-(2-nitrophenyl) acrylic acid (**1**) into its respective ester, hydrazide and 1,3,4-oxadiazole. Finally the target compounds were obtained by reaction of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**) with a series of various electrophiles, (**5a-1**), in *N,N*-dimethyl formamide (DMF) in the presence of sodium hydride (NaH). The structural characterization of these newly synthesized compounds was done by IR, ¹H-NMR, HR-MS and EI-MS spectral data. All these compounds were evaluated for their enzyme inhibitory potentials and found to exhibit broad range spectrum against acetylcholinesterase, butyrylcholinesterase and lipoxigenase enzymes.

Keywords: 3-(2-nitrophenyl)acrylic acid, 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol, ¹H-NMR, EI-MS, enzymatic activities.

1. INTRODUCTION

1,3,4-oxadiazole is a versatile group of heterocyclic compounds that have attracted attention for last two decades because of their broad range biological effectiveness.¹ It is derived from furan when the replacement of two methine (-CH=) is done by two pyridine like nitrogen (-N=) at position 3 and 4.² Aromaticity of oxadiazole ring is reduced because of this replacement due to which oxadiazole ring bears properties of conjugated diene. The electrophilic substitutions reactions in oxadiazole ring are very difficult at carbon atom due to low electron density at carbon atom. Conversely the attack of electrophiles can occur at nitrogen atom. Nucleophilic attack can occur if substitution of ring is done with electron releasing groups. Oxadiazole normally do not undergo nucleophilic attack. However halogen substituted oxadiazole undergo nucleophilic substitution reactions. Nucleophilic substitution reactions in oxadiazole are same as occurred at *sp*² carbon atom of an aliphatic compound. Extra heteroatom has inductive effect due to which oxadiazole acts as very weak base.³ 1,3,4-oxadiazole can be used as a skeleton in medicinal chemistry to synthesize large number of bioactive agents.⁴ 2,5-disubstituted-1,3,4-Oxadiazole derivatives have possessed broad range of biological activities such as antifungal,⁵ antimicrobial,^{6,7} anti-inflammatory,^{2,8} and antiparasitic.⁹ HIV replication is inhibited by the use of 2,5-disubstituted derivatives of 1,3,4-oxadiazole.¹⁰ Several 2,5-disubstituted-1,3,4-oxadiazole derivatives have powerful effect against 60 malignant tumor cell lines. Biological effects demonstrate a very significant anti-tumor activity against leukemia, breast cancer and colon.^{11,12} It is extensively used in the treatment of arthritis (jaundice, rheumatoid and osteoarthritis).^{13,14} Oxadiazole also exhibit herbicidal,¹⁵ pesticidal, analgesic¹⁶ and plant growth regulatory activities.¹⁷

Literature survey showed that slight modifications in the structure of 1,3,4-oxadiazole can result in quantitative as well as qualitative variations in the biological activity. So in continuation of our work^{18,19} to search for the enzyme inhibitors bearing 1,3,4-oxadiazole nucleus, here we report a series of *S*-substituted derivatives of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**) derived from 3-(2-nitrophenyl)acrylic acid (**1**) and screened results of enzyme inhibition activities for all the synthesized compounds.

2. EXPERIMENTAL

2.1. Chemistry

Griffin and George melting point apparatus was used to record the melting points of the synthesized compounds by open capillary tube and were uncorrected. Purity was checked on thin layer chromatography (TLC) on pre-coated silica gel G-25-UV254 plates using different percentage of ethyl acetate and *n*-hexane giving single spot. Jasco-320-A spectrophotometer was used for the IR spectra (wave number in cm⁻¹). ¹H-NMR spectra were recorded in CD₃OD or CDCl₃ on a Bruker spectrometers operating at 300 and 400 MHz. Chemical shifts were recorded in ppm. Mass spectra (HE-MS & EIMS) were taken on a JMS-HX-110 spectrometer, with complete data system.

2.2. Procedure for the preparation of Ethyl-3-(2-nitrophenyl) acrylate

3-(2-nitrophenyl)acrylic acid (1g; (**1**)) was taken into a 100 mL round bottom flask fitted with a reflux condenser, then absolute ethanol (4 mL) and conc. sulphuric acid (1/2 mL) were added into flask and the reaction mixture was refluxed for about 1 hour. The reaction was monitored by thin layer chromatography (TLC). When the reaction was completed, the reaction mixture was neutralized with concentrated solution of sodium carbonate (Na₂CO₃). From this mixture, the desired ester was extracted with ether. This ethereal layer was evaporated to afford yellowish brown crystalline substance, ethyl-3-(2-nitrophenyl)acrylate (**2**).

2.3. Procedure for the preparation of 3-(2-nitrophenyl) acrylohydrazide

The methanolic solution of ethyl-3-(2-nitrophenyl)acrylate (0.01mol, 2.21g, (**2**)) was taken in a round bottom flask. Hydrazine hydrate (0.03 mol, 1.47 mL) was added drop wise in this solution with stirring for 3 h at room temperature. Completion of reaction was checked by TLC. After completion, distilled water was added and precipitates of 3-(2-nitrophenyl)acrylohydrazide (**3**) formed, were filtered and re-washed with water.

2.4. Procedure for the preparation of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**)

3-(2-Nitrophenyl)acrylohydrazide (2.07 g, 0.01mol, (**3**)) was solubilized in absolute ethanol (12 mL) in 250 mL round bottom flask and potassium hydroxide (1.68 g, 0.03mol) was added in the reaction mixture. Upon complete dissolution of potassium hydroxide, carbon disulfide (1.81 mL, 0.03mol) was added to the solution. This reaction mixture was well stirred and refluxed for 6 h. Initially the color of solution was yellow which turned to brownish yellow with the progress of reaction. Evolution of hydrogen sulfide gas occurred during this reaction. After the completion of this reaction, it was diluted with water and acidified by concentrated hydrochloric acid to pH 2-3. The precipitates formed, were filtered and washed with water. Re-crystallization was done with ethanol to afford purified sample of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**).

1.1.1 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**)

Yellowish brown solid; m.p. 190-192 °C; yield 79%; IR (KBr, ν_{max} cm⁻¹): 3200 (S-H), 3056 (Ar-H), 1680 (C=N), 1550 (Ar C=C), 1530 (NO₂), 840 (C-N); ¹H-NMR (300 MHz, CD₃OD): δ 8.06 (dd, *J* = 8.1, 1.2 Hz, 1H, H-3'), 7.89 (d, *J* = 16.2 Hz, 1H, H-8'), 7.76 (dd, *J* = 7.8, 1.8 Hz, 1H, H-6'), 7.74 (br t, *J* = 7.8, Hz, 1H, H-4'), 7.61 (ddd, *J* = 8.4, 8.4, 1.2 Hz, 1H, H-5'), 6.95 (d, *J* = 16.2 Hz, 1H, H-7'); ¹³C-NMR (75 MHz, CD₃OD): δ 164.5 (C-2), 163.7 (C-5), 149.3 (C-2'), 138.3 (C-7'), 135.5 (C-5'), 134.3 (C-1'), 132.6 (C-4'), 129.6 (C-6'), 126.9 (C-3'), 110.8 (C-8'); HR-MS: [M]⁺ 249.2685 (Calcd. for C₁₀H₇N₃O₃; 249.2458); EI-MS: *m/z* 249 [M]⁺, 203 [M-NO₂]⁺, 174 [M-CHNOS]⁺, 148 [M-C₂HN₂OS]⁺, 135 [M-C₃H₂N₂OS]⁺, 122 [M-C₄H₃N₂OS]⁺.

2.5. General procedure for the preparation of *S*-substituted derivatives (**6a-n**) of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**)

The calculated amount of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (0.2g, 0.0008mol, (**4**)) was taken in a 50 mL round bottom flask and 10 mL of *N,N*-dimethyl formamide (DMF) was added in order to dissolve it completely. Then

Lithium hydride (0.0064g, 0.0008mol) was added to the mixture as base. The reaction mixture was stirred for 30 minutes at room temperature and then respective alkyl halide (0.0008mol, **5a-n**) was added to the reaction mixture and it was further stirred for 3 h. The progress of reaction was monitored by TLC test till single spot. Then few drops of 5% solution of sodium hydroxide were added to convert the unreacted oxadiazole-2-thiol into its salt. Finally, the addition of distilled water into this solution resulted in the formation of precipitates of respective S-substituted derivative (**6a-n**) of parent molecule. Precipitates were filtered and washed with water and dried to carry out further studies.

2.5.1. 2-(Ethylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6a)

Brownish yellow amorphous solid; m.p. 84-86 °C; yield 84%; IR (KBr, ν_{max} cm^{-1}): 3049 (Ar-H), 1683 (C=N), 1555 (Ar C=C), 1527 (NO₂), 844 (C-N); ¹H-NMR (300 MHz, CD₃OD): δ 8.06 (dd, $J = 8.4, 1.2$ Hz, 1H, H-3'), 7.95 (d, $J = 16.5$ Hz, 1H, H-8'), 7.93 (br d, $J = 7.5$ Hz, 1H, H-6'), 7.75 (br t, $J = 7.2$ Hz, 1H, H-4'), 7.62 (br t, $J = 7.5$ Hz, 1H, H-5'), 7.14 (d, $J = 16.2$ Hz, 1H, H-7'), 3.33 (q, $J = 7.2$ Hz, 2H, CH₂-1''), 1.49 (t, $J = 7.2$ Hz, 3H, CH₃-2''); ¹³C-NMR (75 MHz, CD₃OD): δ 164.1 (C-2), 163.3 (C-5), 149.2 (C-2''), 138.1 (C-7'), 135.3 (C-5'), 134.1 (C-1'), 132.2 (C-4'), 129.1 (C-6'), 126.7 (C-3'), 110.3 (C-8'), 23.4 (C-1''), 16.5 (C-2''); HR-MS: [M]⁺ 277.3067 (Calcd. for C₁₅H₁₁N₃O₃S; 277.2991); EI-MS: m/z 277 [M]⁺, 248 [M-C₂H₅]⁺, 231 [M-NO₂]⁺, 203 [M-CNOS]⁺, 177 [M-C₂N₂OS]⁺, 164 [M-C₂H₅N₂OS]⁺, 151 [M-C₄H₉N₂OS]⁺.

2.5.2. 2-(Isopropylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6b)

Brownish yellow amorphous solid; m.p. 68-70 °C; yield 80%; IR (KBr, ν_{max} cm^{-1}): 3059 (Ar-H), 1686 (C=N), 1557 (Ar C=C), 1533 (NO₂), 847 (C-N); ¹H-NMR (300 MHz, CD₃OD): δ 8.06 (dd, $J = 8.1, 1.2$ Hz, 1H, H-3'), 7.96 (d, $J = 16.8$ Hz, 1H, H-8'), 7.93 (br d, $J = 7.2$ Hz, 1H, H-6'), 7.76 (br t, $J = 7.2$ Hz, 1H, H-4'), 7.62 (ddd, $J = 8.4, 8.1, 0.9$ Hz, 1H, H-5'), 7.14 (d, $J = 16.2$ Hz, 1H, H-7'), 3.94 (septet, $J = 6.9$ Hz, 1H, H-1''), 1.51 (d, $J = 6.6$ Hz, 6H, CH₃-2'', CH₃-3''); ¹³C-NMR (75 MHz, CD₃OD): δ 164.3 (C-2), 163.5 (C-5), 148.9 (C-2''), 138.3 (C-7'), 135.6 (C-5'), 133.7 (C-1'), 132.5 (C-4'), 129.5 (C-6'), 126.3 (C-3'), 109.8 (C-8'), 37.2 (C-1''), 23.9 (C-2'' & C-3''); HR-MS: [M]⁺ 291.3367 (Calcd. for C₁₅H₁₃N₃O₃S; 291.3256); EI-MS: m/z 291 [M]⁺, 245 [M-NO₂]⁺, 248 [M-C₃H₇]⁺, 217 [M-CNOS]⁺, 191 [M-C₂N₂OS]⁺, 178 [M-C₃H₇N₂OS]⁺, 165 [M-C₄H₉N₂OS]⁺.

2.5.3. 5-(2-Nitrostyryl)-2-(vinylthio)-1,3,4-oxadiazole (6c)

Off-white amorphous solid; m.p. 92-94 °C; yield 78%; IR (KBr, ν_{max} cm^{-1}): 3056 (Ar-H), 1684 (C=N), 1553 (Ar C=C), 1526 (NO₂), 848 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, $J = 8.0$ Hz, 1H, H-3'), 7.94 (d, $J = 16.4$ Hz, 1H, H-8'), 7.68 (m, 2H, H-4', H-6'), 7.54 (br t, $J = 7.4$ Hz, 1H, H-5'), 6.98 (d, $J = 16.4$ Hz, 1H, H-7'), 6.88 (dd, $J = 16.8, 9.6$ Hz, 1H, H-1''), 5.76 (br d, $J = 17.2$ Hz, 1H, H-2''), 5.72 (br d, $J = 9.6$ Hz, 1H, H-2''); ¹³C-NMR (75 MHz, CD₃OD): δ 164.2 (C-2), 163.7 (C-5), 148.6 (C-2'), 138.8 (C-7'), 135.5 (C-1'), 132.9 (C-4'), 129.6 (C-6'), 126.6 (C-3'), 116.9 (C-2''), 109.6 (C-8'); HR-MS: [M]⁺ 275.2997 (Calcd. for C₁₅H₉N₃O₃S; 275.2831); EI-MS: m/z 275 [M]⁺, 248 [M-C₂H₅]⁺, 229 [M-NO₂]⁺, 201 [M-CNOS]⁺, 175 [M-C₂N₂OS]⁺, 162 [M-C₃H₅N₂OS]⁺, 149 [M-C₄H₇N₂OS]⁺.

2.5.4. 2-(Allylthio)-5-(2-Nitrostyryl)-1,3,4-oxadiazole (6d)

Yellow amorphous solid; m.p. 75-77 °C; yield 81%; IR (KBr, ν_{max} cm^{-1}): 3056 (Ar-H), 1688 (C=N), 1554 (Ar C=C), 1539 (NO₂), 846 (C-N); ¹H-NMR (300 MHz, CD₃OD): δ 8.06 (dd, $J = 8.1, 1.2$ Hz, 1H, H-3'), 7.96 (d, $J = 16.2$ Hz, 1H, H-8'), 7.92 (br d, $J = 7.5$ Hz, 1H, H-6'), 7.75 (br t, $J = 7.8$ Hz, 1H, H-4'), 7.62 (ddd, $J = 8.7, 8.1, 1.5$ Hz, 1H, H-5'), 7.14 (d, $J = 16.2$ Hz, 1H, H-7'), 6.04 (m, 1H, H-2''), 5.39 (dd, $J = 17.1, 1.2$ Hz, 1H, H₃-3''), 5.20 (dd, $J = 9.9, 0.6$ Hz, 1H, H₂-3''), 3.93 (d, $J = 7.2$ Hz, 2H, CH₂-1''); ¹³C-NMR (75 MHz, CD₃OD): δ 164.6 (C-2), 163.9 (C-5), 148.4 (C-2'), 138.7 (C-7'), 135.2 (C-5'), 133.4 (C-1'), 132.1 (C-4'), 129.8 (C-6'), 127.2 (C-2''), 126.7 (C-3'), 119.8 (C-3''), 110.8 (C-8'), 36.9 (C-2''); HR-MS: [M]⁺ 289.3286 (Calcd. for C₁₅H₁₁N₃O₃S; 289.3097); EI-MS: m/z 289 [M]⁺, 248 [M-C₃H₇]⁺, 243 [M-NO₂]⁺, 215 [M-CNOS]⁺, 189 [M-C₂N₂OS]⁺, 176 [M-C₃H₇N₂OS]⁺, 163 [M-C₄H₉N₂OS]⁺.

2.5.5. 5-(2-Nitrostyryl)-2-(phenylethylthio)-1,3,4-oxadiazole (6e)

Off-white amorphous powder; m.p. 85-87 °C; yield 70%; IR (KBr, ν_{max} cm^{-1}): 3056 (Ar-H), 1680 (C=N), 1550 (Ar C=C), 1532 (NO₂), 844 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, $J = 8.0$ Hz, 1H, H-3'), 7.91 (d, $J = 16.0$ Hz, 1H, H-8'), 7.69 (m, 2H, H-4', H-6'), 7.54 (br t, $J = 8.4$ Hz, 1H, H-5'), 7.30 (br d, $J = 7.2$ Hz, 2H, H-2'', H-6''), 7.24 (m, 3H, H-3'' to H-5''), 6.97 (d, $J = 16.4$ Hz, 1H, H-7'), 3.52 (t, $J = 7.2$ Hz, 2H, CH₂-8''), 3.14 (t, $J = 7.12$ Hz, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.6 (C-2), 163.5 (C-5), 148.9 (C-2'), 140.5 (C-1''), 138.5 (C-7'), 135.6 (C-5'), 133.7 (C-1'), 132.6 (C-4'), 131.7 (C-3'' & C-5''), 129.9 (C-6'), 129.5 (C-2'' & C-6''), 127.4 (C-4''), 126.5 (C-3'), 110.8 (C-8'), 35.3 (C-7''), 28.5 (C-8''); HR-MS: [M]⁺ 353.4072 (Calcd. for C₁₈H₁₅N₃O₃S; 353.3951); EI-MS: m/z 353 [M]⁺, 307 [M-NO₂]⁺, 262 [M-C₆H₅]⁺, 248 [M-C₈H₉]⁺, 279 [M-CNOS]⁺, 253 [M-C₂N₂OS]⁺, 240 [M-C₃H₇N₂OS]⁺, 227

[M-C₄H₉N₂OS]⁺.

2.5.6. 5-(2-Nitrostyryl)-2-(3-phenylpropylthio)-1,3,4-oxadiazole (6f)

Yellow amorphous solid; m.p. 78-80 °C; yield 80%; IR (KBr, ν_{max} cm^{-1}): 3051 (Ar-H), 1684 (C=N), 1558 (Ar C=C), 1531 (NO₂), 843 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, $J = 8.0$ Hz, 1H, H-3'), 7.90 (d, $J = 16.0$ Hz, 1H, H-8'), 7.67 (m, 2H, H-4', H-6'), 7.55 (br t, $J = 8.0$ Hz, 1H, H-5'), 7.26 (t, $J = 6.8$ Hz, 3H, H-3'', H-4'', H-5''), 7.19 (br d, $J = 6.8$ Hz, 2H, H-2'', H-6''), 6.97 (d, $J = 16.0$ Hz, 1H, H-7''), 3.28 (t, $J = 7.2$ Hz, 2H, CH₂-9''), 2.79 (t, $J = 7.2$ Hz, 2H, CH₂-7''), 2.17 (quint., $J = 7.6$ Hz, 2H, CH₂-8''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.3 (C-2), 163.1 (C-5), 148.6 (C-2'), 142.4 (C-1''), 138.8 (C-7'), 135.9 (C-5'), 133.7 (C-1'), 132.7 (C-4'), 131.5 (C-2'' & C-6''), 129.9 (C-6'), 129.5 (C-3'' & C-5''), 126.7 (C-4''), 126.3 (C-3'), 110.8 (C-8'), 33.9 (C-7''), 30.4 (C-8''), 27.8 (C-9''); HR-MS: [M]⁺ 367.4401 (Calcd. for C₁₉H₁₇N₃O₃S; 367.4216); EI-MS: m/z 367 [M]⁺, 321 [M-NO₂]⁺, 276 [M-C₆H₅]⁺, 248 [M-C₈H₉]⁺, 293 [M-CNOS]⁺, 267 [M-C₂N₂OS]⁺, 254 [M-C₃H₇N₂OS]⁺, 241 [M-C₄H₉N₂OS]⁺.

2.5.7. 2-(Benzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6g)

Brownish yellow amorphous solid; m.p. 128-130 °C; yield 73%; IR (KBr, ν_{max} cm^{-1}): 3051 (Ar-H), 1685 (C=N), 1559 (Ar C=C), 1532 (NO₂), 847 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.04 (d, $J = 8.0$ Hz, 1H, H-3'), 7.89 (d, $J = 16.0$ Hz, 1H, H-8'), 7.67 (br d, $J = 7.6$ Hz, 2H, H-4', H-6'), 7.53 (ddd, $J = 8.4, 7.8, 1.6$ Hz, 1H, H-5'), 7.44 (br d, $J = 7.2$ Hz, 2H, H-2'', H-6''), 7.28-7.34 (m, 3H, H-3'' to H-5''), 6.96 (d, $J = 16.0$ Hz, 1H, H-7''), 4.50 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.7 (C-2), 163.5 (C-5), 148.2 (C-2'), 138.4 (C-7'), 137.5 (C-1''), 135.6 (C-5'), 133.5 (C-1'), 132.4 (C-4'), 129.7 (C-6'), 129.4 (C-3'' & C-5''), 128.5 (C-2'' & C-6''), 128.1 (C-4''), 126.5 (C-3'), 110.5 (C-8'), 32.5 (C-7''); HR-MS: [M]⁺ 339.3827 (Calcd. for C₁₇H₁₃N₃O₃S; 339.3684); EI-MS: m/z 339 [M]⁺, 293 [M-NO₂]⁺, 248 [M-C₆H₅]⁺, 265 [M-CNOS]⁺, 239 [M-C₂N₂OS]⁺, 226 [M-C₃H₇N₂OS]⁺, 213 [M-C₄H₉N₂OS]⁺.

2-(2-Methylbenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6h)

Off-white amorphous solid; m.p. 148-150 °C; yield 78%; IR (KBr, ν_{max} cm^{-1}): 3050 (Ar-H), 1686 (C=N), 1552 (Ar C=C), 1534 (NO₂), 846 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.04 (d, $J = 8.0$ Hz, 1H, H-3'), 7.91 (d, $J = 16.4$ Hz, 1H, H-8'), 7.69 (m, 2H, H-4', H-6'), 7.53 (br t, $J = 8.4$ Hz, 1H, H-5'), 7.40 (br d, $J = 7.2$ Hz, 1H, H-4''), 7.20 (m, 3H, H-3'' to H-6''), 6.97 (d, $J = 16.4$ Hz, 1H, H-7''), 4.45 (s, 2H, CH₂-7''), 2.44 (s, 3H, CH₃-8''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.1 (C-2), 163.5 (C-5), 148.9 (C-2'), 141.4 (C-1''), 138.6 (C-7'), 136.3 (C-2''), 135.6 (C-5'), 134.2 (C-3''), 133.6 (C-1'), 132.5 (C-4'), 129.9 (C-4''), 129.7 (C-6''), 129.6 (C-6'), 127.5 (C-5''), 126.5 (C-3'), 110.5 (C-8'), 28.5 (C-7''), 19.5 (C-8''); HR-MS: [M]⁺ 353.4121 (Calcd. for C₁₈H₁₅N₃O₃S; 353.3951); EI-MS: m/z 353 [M]⁺, 338 [M-CH₃]⁺, 307 [M-NO₂]⁺, 248 [M-C₈H₉]⁺, 279 [M-CNOS]⁺, 253 [M-C₂N₂OS]⁺, 240 [M-C₃H₇N₂OS]⁺, 227 [M-C₄H₉N₂OS]⁺.

2.5.8. 2-(4-Fluorobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6i)

Off-white amorphous solid; m.p. 141-143 °C; yield 79%; IR (KBr, ν_{max} cm^{-1}): 3059 (Ar-H), 1686 (C=N), 1553 (Ar C=C), 1536 (NO₂), 842 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, $J = 8.0$ Hz, 1H, H-3'), 7.90 (d, $J = 16.0$ Hz, 1H, H-8'), 7.67 (m, 2H, H-4', H-6'), 7.55 (br t, $J = 8.0$ Hz, 1H, H-5'), 7.42 (d, $J = 8.4$ Hz, 2H, H-2'', H-6''), 7.02 (d, $J = 8.4$ Hz, 2H, H-3'', H-5''), 6.96 (d, $J = 16.0$ Hz, 1H, H-7''), 4.45 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.1 (C-2), 163.3 (C-5), 162.5 (C-4''), 148.8 (C-2'), 138.5 (C-7'), 135.7 (C-5'), 133.5 (C-1'), 132.4 (C-4'), 132.6 (C-2'' & C-6''), 131.7 (C-1''), 129.7 (C-6'), 126.7 (C-3'), 115.6 (C-3'' & C-5''), 110.5 (C-8'), 32.5 (C-7''); HR-MS: [M]⁺ 357.3692 (Calcd. for C₁₇H₁₂FN₃O₃S; 357.3588); EI-MS: m/z 357 [M]⁺, 311 [M-NO₂]⁺, 248 [M-C₆H₄F]⁺, 283 [M-CNOS]⁺, 257 [M-C₂N₂OS]⁺, 244 [M-C₃H₇N₂OS]⁺, 231 [M-C₄H₉N₂OS]⁺.

2.5.9. 2-(2-Chlorobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6j)

Off-white amorphous solid; m.p. 144-146 °C; yield 72%; IR (KBr, ν_{max} cm^{-1}): 3051 (Ar-H), 1683 (C=N), 1556 (Ar C=C), 1530 (NO₂), 848 (C-N), 560 (C-Cl); ¹H-NMR (400 MHz, CDCl₃): δ 8.04 (d, $J = 8.4$ Hz, 1H, H-3'), 7.90 (d, $J = 16.0$ Hz, 1H, H-8'), 7.67 (m, 2H, H-4', H-6'), 7.59 (dd, $J = 8.8, 2.4$ Hz, 1H, H-3''), 7.52 (ddd, $J = 8.4, 8.4, 2.0$ Hz, 1H, H-5''), 7.40 (dd, $J = 9.2, 2.4$ Hz, 1H, H-6''), 7.24 (m, 2H, H-4'', H-5''), 6.95 (d, $J = 16.0$ Hz, 1H, H-7''), 4.61 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.3 (C-2), 163.4 (C-5), 148.7 (C-2'), 138.3 (C-7'), 135.9 (C-5'), 135.8 (C-1''), 133.9 (C-1'), 132.4 (C-4'), 130.8 (C-5''), 130.6 (C-3''), 129.3 (C-6'), 128.9 (C-4''), 128.3 (C-6''), 127.4 (C-2''), 126.7 (C-3'), 110.7 (C-8'), 31.4 (C-7''); HR-MS: [M]⁺ 373.8314 (Calcd. for C₁₇H₁₂ClN₃O₃S; 373.8134); EI-MS: m/z 374 [M]⁺, 328 [M-NO₂]⁺, 249 [M-C₆H₄Cl]⁺, 299 [M-CNOS]⁺, 273 [M-C₂N₂OS]⁺, 260 [M-C₃H₇N₂OS]⁺, 247 [M-C₄H₉N₂OS]⁺.

2.5.10. 2-(3-Chlorobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6k)

Light brownish yellow solid; m.p. 120-122 °C; yield 75%; IR (KBr, ν_{max} cm^{-1}): 3056 (Ar-H), 1687 (C=N), 1552 (Ar C=C), 1539 (NO₂), 845 (C-N), 563

(C-Cl); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, *J* = 8.0 Hz, 1H, H-3'), 7.90 (d, *J* = 16.0 Hz, 1H, H-8'), 7.67 (m, 2H, H-4', H-6'), 7.54 (ddd, *J* = 8.4, 7.4, 1.6 Hz, 1H, H-5'), 7.44 (br s, 1H, H-2''), 7.34 (m, 1H, H-4''), 7.27 (m, 2H, H-5'', H-6''), 6.95 (d, *J* = 16.0 Hz, 1H, H-7'), 4.56 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.5 (C-2), 163.6 (C-5), 148.6 (C-2'), 139.6 (C-1''), 138.5 (C-7'), 136.4 (C-3''), 135.6 (C-5'), 133.6 (C-1'), 132.2 (C-4'), 130.8 (C-4''), 130.3 (C-5''), 129.5 (C-6'), 128.4 (C-2''), 126.5 (C-3'), 124.5 (C-6''), 110.3 (C-8'), 33.5 (C-7''); HR-MS: [M]⁺ 373.8314 (Calcd. for C₁₇H₁₂ClN₃O₃S; 373.8134); EI-MS: *m/z* 374 [M]⁺, 328 [M-NO₂]⁺, 249 [M-C₂H₆Cl]⁺, 299 [M-CNOS]⁺, 273 [M-C₂N₂OS]⁺, 260 [M-C₂H₂N₂OS]⁺, 247 [M-C₄H₂N₂OS].

2.5.11. 2-(4-Chlorobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6l)

Cream yellow solid; m.p. 128-130 °C; yield 76%; IR (KBr, ν_{\max} cm⁻¹): 3053 (Ar-H), 1681 (C=N), 1558 (Ar C=C), 1534 (NO₂), 847 (C-N), 566 (C-Cl); ¹H-NMR ((400 MHz, CDCl₃): δ 8.05 (d, *J* = 8.0 Hz, 1H, H-3'), 7.90 (d, *J* = 16.4 Hz, 1H, H-8'), 7.68 (m, 2H, H-4', H-6'), 7.54 (ddd, *J* = 8.4, 7.4, 1.6 Hz, 1H, H-5''), 7.39 (d, *J* = 8.0 Hz, 2H, H-2'', H-6''), 7.30 (d, *J* = 8.4 Hz, 2H, H-3'', H-5''), 6.95 (d, *J* = 16.0 Hz, 1H, H-7'), 4.45 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.4 (C-2), 163.7 (C-5), 148.6 (C-2'), 138.3 (C-7'), 136.1 (C-4''), 135.6 (C-5'), 135.2 (C-1'), 133.7 (C-1''), 132.5 (C-4'), 131.3 (C-3'' & C-5''), 129.6 (C-6'), 126.9 (C-2'' & C-6''), 126.4 (C-3'), 110.8 (C-8'), 32.7 (C-7''); HR-MS: [M]⁺ 373.8314 (Calcd. for C₁₇H₁₂ClN₃O₃S; 373.8134); EI-MS: *m/z* 374 [M]⁺, 328 [M-NO₂]⁺, 249 [M-C₂H₆Cl]⁺, 300 [M-CNOS]⁺, 274 [M-C₂N₂OS]⁺, 261 [M-C₂H₂N₂OS]⁺, 248 [M-C₄H₂N₂OS].

2.5.12. 2-(4-Bromobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6m)

Pale yellow amorphous solid; m.p. 110-112 °C; yield 78%; IR (KBr, ν_{\max} cm⁻¹): 3058 (Ar-H), 1688 (C=N), 1553 (Ar C=C), 1537 (NO₂), 844 (C-N), 510 (C-Br); ¹H-NMR (400 MHz, CDCl₃): δ 8.05 (d, *J* = 8.0 Hz, 1H, H-3'), 7.90 (d, *J* = 16.0 Hz, 1H, H-8'), 7.68 (m, 1H, H-6', H-4'), 7.54 (br t, *J* = 8.4 Hz, 1H, H-5'), 7.45 (d, *J* = 8.4 Hz, 2H, H-2'', H-6''), 7.33 (d, *J* = 8.0 Hz, 2H, H-3'', H-5''), 6.95 (d, *J* = 16.4 Hz, 1H, H-7'), 4.44 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.2 (C-2), 163.4 (C-5), 148.1 (C-2'), 138.7 (C-7'), 136.2 (C-1''), 135.4 (C-5'), 133.8 (C-1'), 133.4 (C-3'' & C-5''), 132.9 (C-4'), 131.4 (C-2'' & C-6''), 129.1 (C-6'), 126.7 (C-3'), 124.2 (C-4''), 110.2 (C-8'), 32.4 (C-7''); HR-MS: [M]⁺ 418.2823 (Calcd. for C₁₇H₁₂BrN₃O₃S; 418.2644); EI-MS: *m/z* 418 [M]⁺, 372 [M-NO₂]⁺, 248 [M-C₂H₆Br]⁺, 344 [M-CNOS]⁺, 318 [M-C₂N₂OS]⁺, 305 [M-C₂H₂N₂OS]⁺, 292 [M-C₄H₂N₂OS].

2.5.13. 2-(3-Nitrobenzylthio)-5-(2-nitrostyryl)-1,3,4-oxadiazole (6n)

Off-white amorphous solid; m.p. 82-84 °C; yield 78%; IR (KBr, ν_{\max} cm⁻¹): 3050 (Ar-H), 1685 (C=N), 1552 (Ar C=C), 1538 (NO₂), 843 (C-N); ¹H-NMR (400 MHz, CDCl₃): δ 8.34 (br s, 1H, H-2''), 8.15 (br d, *J* = 8.0 Hz, 1H, H-4''), 8.05 (d, *J* = 8.0 Hz, 1H, H-3'), 7.91 (d, *J* = 16.4 Hz, 1H, H-8'), 7.84 (br d, *J* = 7.6 Hz, 1H, H-6'), 7.68 (m, 2H, H-4', H-6'), 7.51-7.56 (m, 2H, H-3'', H-5''), 6.94 (d, *J* = 16.4 Hz, 1H, H-7'), 4.56 (s, 2H, CH₂-7''); ¹³C-NMR (75 MHz, CD₃OD): δ 165.1 (C-2), 163.9 (C-5), 149.6 (C-3''), 148.5 (C-2'), 138.9 (C-7'), 138.2 (C-1''), 135.1 (C-5'), 133.2 (C-1'), 132.6 (C-6''), 132.4 (C-4'), 131.3 (C-5''), 129.2 (C-6'), 126.8 (C-4''), 126.3 (C-3'), 124.4 (C-2''), 110.7 (C-8'), 34.5 (C-7''); HR-MS: [M]⁺ 384.3847 (Calcd. for C₁₇H₁₂N₃O₅S; 384.3659); EI-MS: *m/z* 384 [M]⁺, 338 [M-NO₂]⁺, 292 [M-N₂O]⁺, 248 [M-C₂H₂NO₂]⁺, 310 [M-CNOS]⁺, 284 [M-C₂N₂OS]⁺, 271 [M-C₂H₂N₂OS]⁺, 258 [M-C₄H₂N₂OS].

3. Enzyme Inhibition Assays

3.1. Acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) Assays

The AChE and BChE inhibition activities were performed according to the reported method²⁰ with slight modifications. Total volume of the reaction mixture was 100 μL. It contained 60 μL Na₂HPO₄ buffer with concentration of 50 mM and pH 7.7. 10 μL test compound (0.5 mM well⁻¹) was added, followed by the addition of 10 μL (0.005 unit well⁻¹) enzyme. The contents were mixed and pre-read at 405 nm. Then contents were pre-incubated for 10 min at 37 °C. The reaction was initiated by the addition of 10 μL of 0.5 mM well⁻¹ substrate (acetylthiocholine iodide), followed by the addition of 10 μL DTNB (0.5 mM well⁻¹). After 15 min of incubation at 37 °C absorbance was measured at 405 nm using 96-well plate reader Synergy HT, Biotek, USA. All experiments were carried out with their respective controls in triplicate. Eserine (0.5 mM well⁻¹) was used as a positive control. The percent inhibition was calculated by the help of following equation

IC_{50} values were calculated using EZ-Fit Enzyme kinetics software (Perrella Scientific Inc. Amherst, USA).

3.2. Lipoxigenase (LOX) Assay

Lipoxigenase activity was assayed according to the reported methods²¹⁻²³ with slight modifications. A total volume of 200 μL lipoxigenase assay mixture contained 150 μL sodium phosphate buffer (100 mM, pH 8.0), 10 μL test compound and 15 μL purified lipoxigenase enzyme (600 units well⁻¹, Sigma Inc.). The contents were mixed and pre-read at 234 nm and preincubated for

10 minutes at 25 °C. The reaction was initiated by addition of 25 μL substrate solution. The change in absorbance was observed after 6 min at 234 nm using 96-well plate reader Synergy HT, Biotek, USA. All reactions were performed in triplicates. The positive and negative controls were included in the assay. Baicalin (0.5 mM well⁻¹) was used as a positive control. The percentage inhibition (%) was calculated by formula given below.

Where Control = Total enzyme activity without inhibitor

Test = Activity in the presence of test compound

IC_{50} values was calculated using EZ-Fit Enzyme Kinetics software (Perrella Scientific Inc. Amherst, USA).

3. RESULTS AND DISCUSSION

3.1. Chemistry

3-(2-nitrophenyl)acrylic acid (**1**) was converted to corresponding ester **2**, hydrazide **3** and 1,3,4-oxadiazole **4**, successively. Finally **6a-n** derivatives of **4** were synthesized by reacting it with different electrophiles **5a-n** (scheme 1). Structures of the synthesized compounds were elucidated and confirmed by IR, ¹H-NMR, ¹³C-NMR, HR-MS and EI-MS techniques. For compound **4**, the IR absorption band of aromatic C-H str. appeared at 3056 and a band at 1530 cm⁻¹ indicated the presence of a nitro group while the peak at 1680 cm⁻¹ (C=N) was typical for an oxadiazole ring. Its molecular formula was confirmed through HR-MS showing molecular ion peak at *m/z* 249.2685, corresponding to C₁₀H₇N₃O₃ (Calcd. for 249.2458). The EI-MS spectral data was also in complete agreement for the proposed structure of 5-(2-nitrostyryl)-1,3,4-oxadiazole-2-thiol (**4**) which was finally confirmed through its ¹H-NMR spectrum. The peaks at δ 8.06 (dd, *J* = 8.1, 1.2 Hz, 1H, H-3'), 7.76 (dd, *J* = 7.8, 1.8 Hz, 1H, H-6'), 7.74 (br t, *J* = 7.8, Hz, 1H, H-4'), and 7.61 (ddd, *J* = 8.4, 8.4, 1.2 Hz, 1H, H-5') for aromatic protons and their splitting pattern confirmed that the ring was *ortho*-substituted, while the larger coupling constants for two olefinic protons at δ 7.89 (d, *J* = 16.2 Hz, 1H, H-8'), and 6.95 (d, *J* = 16.2 Hz, 1H, H-7') disclosed their *trans* disposition. The structure was also thorough supported by its ¹³C-NMR spectrum. Similarly all other synthesized derivatives were characterized by aforesaid spectral techniques and screened for enzyme inhibitory activities to check their possible therapeutic potential for the treatment of various ailments associated with the over expression of studied enzymes.

3.2. Enzyme Inhibition Activity:

3.2.1. AChE inhibitory potential

All the synthesized compounds were screened against acetylcholinesterase and these molecules exhibited weak to moderate inhibitory potential against this enzyme (table 1). Among these, the parent molecule **4** exhibited relatively greater inhibitory potential having IC_{50} value of 68.91±0.22 μM relative to eserine, a reference standard, having a value of 0.04±0.001 μM. Moreover, it was also inferred that the substitution of various alkyl/aralkyl groups at the sulfur atom of the parent molecule **4** was not an effective tool to enhance the biological activity of such molecules against this enzyme.

3.2.2. BChE inhibitory activity

Butyrylcholinesterase inhibition potential was also analyzed for all these derivatives. Results (table 1) revealed that a number of derivatives possess moderate inhibitory potential against this enzyme. The molecule **6n** showed better activity with IC_{50} of 61.21±0.12 μM which could be attributed to the substitution of 3-nitrobenzyl group in this molecule. The compounds **6m**, **6c**, **6i** and **6k** demonstrated an endurable potential with IC_{50} values 73.21±0.17, 74.51±0.11, 80.31±0.21 and 99.61±0.15 μM, respectively which depicted the impact of substitution of various moieties on the parent molecule.

3.2.3. LOX inhibition potential

A few of the synthesized compounds indicated very temperate potential against lipoxigenase (table 1). The molecule **6m** exhibited better potential with IC_{50} value of 28.11±0.11 μM. The compounds **6b**, **6d** and **6g** also bared a moderate inhibitory potential with IC_{50} values of 99.61±0.18, 85.75±0.11 and 76.75±0.14 respectively, relative to baicalin having the IC_{50} value of 22.4±1.3 μM.

3.2.4. Structure-activity relationship

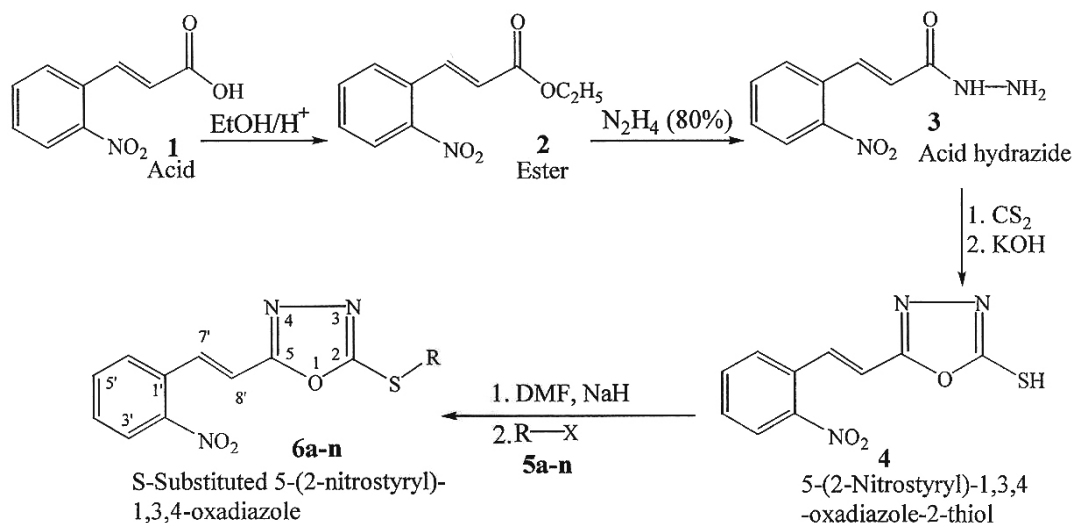
Overall, the heterocycle **6m** was found to exhibit inhibitory activity against all the three enzymes namely, acetylcholinesterase, butyrylcholinesterase and lipoxigenase, but inhibition activity against lipoxigenase is appreciable, while some compounds were active against butyrylcholinesterase while some others revealed inhibitory potential against lipoxigenase. The **6m** derivative possesses a *p*-bromobenzyl substitution on parent oxadiazole ring and mesomeric and inductive effect on parent moiety collectively could influence

enzyme inhibition activity against all the three enzymes. However, the better inhibition potential by parent **4** against acetylcholinesterase enzyme revealed that the substitution of such groups was not a useful means to synthesize potent inhibitors of this enzyme. In compound **6n**, the substitution of *m*-nitrobenzyl group makes it a better entity as butyrylcholinesterase inhibitor. However, the compounds bearing olefinic group and *m*- and *p*-substituted halogenated aromatic ring also rendered moderate inhibition against this enzyme.

4. CONCLUSION

The anticipated structures of the synthesized derivatives of

1,3,4-oxadiazole are well supported by spectroscopic analysis. The enzyme inhibition data help to conclude that the compounds reveal moderate activity against butyrylcholinesterase enzyme, and sound inhibitory potential against acetylcholinesterase and lipoxygenase enzymes, as it was evident from their IC_{50} values relative to the standard used. Hence, on the basis of aforesaid results, these synthesized derivatives provide an overall indispensable basis to introduce potent drugs. It is concluded from the described research work that halogenated aromatic substituted derivatives of 1,3,4-oxadiazoles seem valuable drug candidates for the treatment of Alzheimer and inflammatory diseases.



Compound	R	Compound	R	Compound	R
6a	$-\text{H}_2\text{C}-\text{CH}_3$	6f		6k	
6b		6g		6l	
6c		6h		6m	
6d		6i		6n	
6e		6j			

Scheme-1: Outline for the synthesis of *S*-substituted derivatives, **6a-n**, of 5-(2-nitrophenyl)-1,3,4-oxadiazole-2-thiol (**4**)

Table-1: Enzyme inhibition activity of *S*-substituted derivatives, **6a-n**, of 5-(2-nitrophenyl)-1,3,4-oxadiazole-2-thiol (**4**).

S. Code	AChE		BChE		LOX	
	Inhibition (%) Conc./well (0.5 mM)	IC ₅₀ μM	Inhibition (%) Conc./well (0.5 mM)	IC ₅₀ μM	Inhibition (%) Conc./well (0.5 mM)	IC ₅₀ μM
4	89.51±0.56	68.91±0.22	71.31±0.62	115.41±0.22	52.91±0.51	> 300
6a	72.34±0.37	135.21±0.31	69.39±0.14	132.61±0.12	63.23±0.33	201.31±0.23
6b	66.56± 0.68	254.61±0.18	68.11± 0.34	138.71±0.18	69.33± 0.61	99.61±0.18
6c	61.24±0.55	301.21±0.19	86.51±0.31	74.51±0.11	27.91±0.14	-
6d	57.41±0.68	> 400	22.41±0.88	-	73.65±0.32	85.75±0.11
6e	-	-	-	-	-	-
6f	62.13±0.22	289.61±0.19	70.97±0.91	114.21±0.14	61.92±0.87	201.91±0.14
6g	54.88±0.52	> 400	33.83±0.45	-	71.51±0.58	76.75±0.14
6h	53.71±0.85	> 400	47.89±0.82	-	63.95±0.18	196.11±0.21
6i	53.41±0.61	> 400	87.54±0.61	80.31±0.21	59.01±0.15	> 400
6j	47.34±0.88	-	65.94±0.66	152.31±0.10	31.69±0.71	-
6k	32.54±0.64	-	76.23±0.11	99.61±0.15	45.93±0.44	-
6l	56.96±0.11	> 400	71.77±0.21	109.61±0.23	37.79±0.21	-
6m	75.89±0.23	101.21±0.17	86.29±0.22	73.21±0.17	98.69±0.22	28.11±0.11
6n	52.51±0.21	> 400	90.78±0.24	61.21±0.12	54.51±0.27	> 400
Control	Eserine 91.29±1.17	0.04±0.001	Eserine 82.82±1.09	0.85±0.0001	Baicalein 93.79±1.27	22.4±1.3

Note: IC₅₀ values (concentration at which there is 50% enzyme inhibition) of compounds were calculated using EZ-Fit Enzyme kinetics software (Perella Scientific Inc. Amherst, USA).

AChE = Acetyl cholinesterase.

BChE = Butyrylcholinesterase.

LOX = Lipoxigenase.

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