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Electronic Supplementary Information

Hierarchically controlled protonation/aggregation of a porphyrin– spermine derivative.

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General experimental methods.

All stock solutions of H₂TCPPSpm4 were prepared dissolving the solid in ultrapure water obtained from Elga Purelab Flex system by Veolia, at pH 1 for HCl, in order to achieve concentrations ranging from $2x10^{-4}$ M to $3x10^{-4}$ M. The concentration of these stock solutions was checked by spectrophotometric experiments using $\varepsilon_{\lambda 435nm} = 338000 \text{ M}^{-1}\text{cm}^{-1}$ at pH=1. To prepare independent solutions we dilute in several cuvettes at different pH, the calculated amount of H₂TCPPSpm4 stock solution to obtain in each cuvettes 2 μ M work solutions. As soon as we prepared the solution, we recorded the spectrum. For the continuous titration we dilute the calculated amount of stock solution to obtain 2 μ M work solution, at pH 1. Then, every 15 minutes, we added small amounts of NaOH to increase the pH of the solution and performed the spectrum. A JASCO V-560 UV-vis spectrophotometer equipped with a 1 cm path-length cell was used for the UV-vis measurements. For RLS and fluorescence data, a fluorolog FL-11 Jobin-Yvon Horiba was used.

SEM analysis of H₂TCPPSpm4 drop casted on silicon substrates at different pHs, has been performed by using a Field Emission Scanning Electron Microscope (Zeiss Supra 55VP). The EDS analysis has been performed using an Oxford x-Act 10 mm2 SSD detector.

The NMR experiments were carried out at 27° C on a Varian UNITY Inova 500 MHz spectrometer (¹H at 499.88 MHz, ¹³C-NMR at 125.7 MHz) equipped with pulse field gradient module (Z axis) and a tuneable 5 mm Varian inverse detection probe (ID-PFG). ESI mass spectra were acquired on an ES-MS Thermo-Finnigan LCQ-DECA using MeOH (positive ion mode). Positive MALDI-TOF mass spectra were acquired by a Voyager DE (PerSeptive Biosystem) equipped with a nitrogen laser (emission at 337 nm for 3 ns) and a flash AD converter (time base 2 ns).

All of the chemicals were purchased from Sigma-Aldrich and used without further purification, H_2 TCPP was purchased from Frontier Scientific and used as received. Ultra-pure water (18.2 M) was used in all experiments.



Synthesis of tri-BOC-Spm (2)

Compound **2** was obtained by a modified literature method [1] with a two-step procedure as shown in Scheme 1.

Synthesis of 2-Tfa

A solution of Spermine (2.123 g, 10.5 mmol) in MeOH (120 mL) was stirred under nitrogen at -78 °C. After 10 min, ethyl trifluoroacetate (1.24 mL, 10.5 mmol) was added dropwise over 1 hour and the temperature was slowly increased to 0°C. Then a solution of Boc₂O (11.46 g, 52.5 mmol) in MeOH (10 mL) was added to reaction mixture during which temperature was brought back to 25° C. After 24 h, the solvent was removed under reduced pressure and the residue was extracted with dichloromethane. The organic phase was washed with H₂O and then dried over anhydrous Na₂SO₄. The product was purified by column chromatography (EtOAc) to give the desired compound (**2-Tfa**) as a yellow oil (3.1 g, 49% yield). ¹H NMR (500 MHz, acetone- d_6): δ 8.54 (br s, 1H); 5.95 (br s, 1H); 3.34-3.24 (m, 10H); 3.07 (m, 2H); 1.82 (br s, 2H); 1.69 (br s, 2H); 1.54-1.41 (m, 31H) ppm. ¹³C NMR (126 MHz, acetone- d_6): δ 155.7, 155.2, 146.7, 116.2, 84.8, 78.4, 77.5, 46.7, 44.0, 37.5, 36.6, 27.7, 27.6, 26.5, 25.5 ppm.

Synthesis of tri-BOC-Spm (2)

The trifluoroacetate protecting group of **2-Tfa** (1.41 g, 2.35 mmol in 5 mL MeOH) was removed with NaOH after stirring for 15 h at room temperature. The solution was concentrated in vacuum and the residue was extracted with dichloromethane. The organic phase was washed with H₂O and then dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure to give the desired product **2** as a yellow oil (1.089 g, 92% yield). ¹H NMR (500 MHz, acetone-*d*₆): δ 5.98 (br s, 1H), 3.30-3.23 (m, 8H), 3.17 (t, *J* =7.0 Hz, 2H), 3.08 (m, 2H), 1.80 (m, 2H), 1.70 (br s, 2H), 1.53 (br s, 4H), 1.46 -1.41 (m, 27H) ppm.; ESI-MS: Mass Calcd. for C₂₅H₅₁N₄O₆ 503.38. Found *m/z* = 503.4 [M+H]⁺.

Synthesis of 3

To a solution of H₂TCPP (200 mg, 0.252 mmol) in dry DMF(2 mL), a solution of HATU (445 mg, 1.17 mmol in 1 mL of DMF) was added dropwise and stirred for 20 min at room temperature under N₂ atmosphere. Then a solution of **2** (510 mg, 1.02 mmol) in dry DMF (2 mL) was added to the reaction mixture. After 40 min *N*,*N*-diisopropylethylamine (200 μ L, 1.17 mmol) was added to the reaction, which was stirred for 24 h at rt. The solvent was removed under reduced pressure and CH₂Cl₂ was added obtaining a precipitate that was removed by filtration. The resulting solution was concentrated under vacuum and purified by column chromatography (CH₂Cl₂/EtOH 90:10) to afford the desired compound **3** as a purple powder (107.6 mg, 16% yield). ¹H NMR (500 MHz, CDCl₃): δ 8.84 (s, 8H), 8.31(br s, 16H), 3.63-3.00 (br s, 48H), 1.91 (br s, 8H), 1.69 (br s, 8H), 1.55-1.28 (m, 124 H), -2.72 (s, 2H) ppm. ¹³C NMR (126 MHz, CDCl₃) 134.5, 125.5, 80.1, 79.6, 55.5, 46.8, 43.5, 37.5, 28.5, 28.47, 28.43, 27.7 ppm. MALDI: Mass Calcd. for C₁₄₈H₂₂₃N₂₀O₂₈ 2728.66. Found *m/z* 2728.92 [M+H]⁺

Synthesis of 1

To a solution of **3** in CH₂Cl₂ (0.04 mmol in 1.5 mL), TFA (20 eq.) was added dropwise under nitrogen and the reaction was stirred at rt for 2h. The reaction was monitored by TLC (CH₂Cl₂/EtOH 90:10) and the solvent was removed under reduced pressure affording the corresponding trifluoroacetate salt compound **1** as a green-purple solid in nearly quantitative yield. ¹H NMR (500 MHz, CD₃OD): δ 8.88 (br, s, 8H), 8.37 (d, *J* = 8.0 Hz, 8H), 8.33 (d, *J* = 8.0 Hz, 8H), 3.71 (br, s, 8H), 3.26-3.06 (m, 40H), 2.19-2.09 (m, 16H), 1.91 (br, s, 16H) ppm. ¹³C NMR (126 MHz, CD₃OD): δ 170.8, 136.4, 135.2, 127.3, 146.5, 121.4, 46.6, 45.8, 37.8, 27.8, 25.3, 24.4, 24.3 ppm. ESI-MS: Mass Calcd. for C₈₈H₁₂₇N₂₀O₄ 1528.03. Found *m/z* 1527.7 [M+H]⁺ (100%), 1453.7 [M-(C₃H₉N₂)]⁺, 1399.6 [M-(C₇H₁₇N₂]⁺, 1325.6 [M-(C₁₀H₂₆N₄]⁺.















[1] a) Geall, A. J.; Blagbrough, I. S.; Tetrahedron, 2000, 56, 2449–2460. b) US2007197658A1.



UV–vis spectra of a solution of $H_2TCPPSpm4$ (2 μM) increasing the pH from 1.3 to 11.1



Fluorescence spectra of a solution of $H_2TCPPSpm4$ (2 μM) increasing the pH from 1.3 to 11.1. The λ ex 424 nm



Fluorescence vs pH, for the continuous titration at 675 nm (solid black square), of 2 µM H₂TCPPSpm4 solution.



SEM images of $H_2TCPPSpm4$ deposited on silicon by drop casting of 100µl of 100 µM solution at pH=1,5 (a), pH=5 (b) and pH=8)c)

SEM images show silicon surface after evaporation of 100 μ l of H₂TCPPSpm4 aqueous solution (10 μ M) at different pH. Clearly, working at pH 1,5 (a) the substrate surface doesn't reveal the presence of any aggregates, while a more structured surface is well evident upon increasing pH value. In particular, small spherical nanostructures are visible at pH 5 (b), while at pH 8 a high density of particles is well detectable (c).

EDS analyses of regions shown in (b) and (c) confirm the carbonaceous nature of such small nanostructures.



EDS spectra of silicon inside (red line) and outside (black line) the H2TCPP-Spm4 drop spot (1 µM, pH=8).

Nanoaggregate dimensions (below tenth of nanometers) confirm the RLS measurements that have shown a signal associable to small aggregates, as expected by the use of extremely diluted solutions.