



Comparison between five acellular oxidative potential measurement assays performed with detailed chemistry on PM₁₀ samples from the city of Chamonix (France)

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Abstract. Many studies have demonstrated associations between exposure to ambient particulate matter (PM) and adverse health outcomes in humans that can be explained by PM capacity to induce oxidative stress *in vivo*. Thus, assays have been developed to quantify the oxidative potential (OP) of PM as a more refined exposure metric than PM mass alone. Only a small number of studies have compared different acellular OP measurements for a given set of ambient PM samples. Yet, fewer studies have compared different assays over a year-long period and with detailed chemical characterization of ambient PM. In this study, we report on seasonal variations of the dithiothreitol (DTT), ascorbic acid (AA), electron spin resonance (ESR) and the respiratory tract lining fluid (RTLFL, composed of the reduced glutathione (GSH) and ascorbic acid (ASC)) assays over a 1-year period in which 100 samples were analyzed. A detailed PM₁₀ characterization allowed univariate and multivariate regression analyses in order to obtain further insight into groups of chemical species that drive OP measurements. Our results show that most of the OP assays were strongly intercorrelated over the sampling year but also these correlations differed when considering specific sampling periods (cold vs. warm). All acellular assays are correlated with a significant number of chemical species when considering univariate correlations, especially for the DTT assay. Evidence is also pre-

sented of a seasonal contrast over the sampling period with significantly higher OP values during winter for the DTT, AA, GSH and ASC assays, which were assigned to biomass burning species by the multiple linear regression models. The ESR assay clearly differs from the other tests as it did not show seasonal dynamics and presented weaker correlations with other assays and chemical species.

1 Introduction

Many studies have demonstrated associations between exposure to ambient particulate matter (PM) and adverse health outcomes in humans. A central mechanism to explain the harmful effects of a range of inhaled particles at the cellular level involves the production of oxidative stress through the generation of excessive reactive oxygen species (ROS) and/or inadequate antioxidant defenses (Borm et al., 2007; Kelly, 2003). The capacity of PM to elicit damaging oxidative reactions is termed oxidative potential (OP).

On this basis, probes have been developed over the last decade to quantify the OP of PM as a more refined exposure metric of PM toxicity than PM mass alone (Ayres et al., 2008; Borm et al., 2007). These probes include several

acellular assays – the most common consisting in mimicking the consumption of antioxidants (e.g., ascorbic acid, AA; reduced glutathione, GSH) or surrogates (e.g., dithiothreitol, DTT), the use of the synthetic human respiratory tract lining fluid (RTLFL) system (again to assess antioxidant depletion), probes measuring HO[•] production or the application of electron spin resonance (ESR) to quantify the ability of PM to induce specific ROS (e.g., HO[•] radicals). Oxidative potential can be considered as an integrative metric of PM characteristics (size, composition, surface area, etc.) potentially linked to the particles' toxicity through oxidative stress. Therefore, they could help to delineate those particle properties (components and sources) responsible for observed health effects. However, each of these assays is sensitive to different panels of ROS-generating compounds, and results are also sensitive to the assay design. Indeed a consensus has yet to emerge regarding a standard in vitro test system or combination of tests that would be most appropriate for PM-related health impact evaluation (Ayres et al., 2008).

Only a small number of studies have compared different acellular OP measurements for a given set of ambient PM samples (Fang et al., 2016; Janssen et al., 2014; Künzli et al., 2006; Szigeti et al., 2015; Visentin et al., 2016; Yang et al., 2014). Yet, fewer studies have compared different assays over a year-long period to gain a better understanding of seasonal variability (Fang et al., 2016; Jedynska et al., 2017; Saffari et al., 2014; Szigeti et al., 2015; Yang et al., 2015). Finally, there is little research relating the oxidative capacity of particulate pollution with detailed chemical characterization of ambient PM, in an attempt to identify the PM components or sources that may contribute most to underlying toxicity (Fang et al., 2016; Kelly et al., 2011; Saffari et al., 2014; Verma et al., 2014; Weber et al., 2018).

In this study, a series of 100 PM₁₀ samples collected over a 1-year period were screened for ROS burden using four acellular measures of OP: DTT, AA, ESR and RTLFL assays. The RTLFL assay includes three antioxidants: reduced glutathione (GSH), ascorbic acid (referred as ASC) and urate (UA). Two ascorbic acid depletion assays (AA and ASC), using different quantification techniques, were therefore included in our analyses. DTT is known to react with organic compounds but also with transition metals (Charrier and Anastasio, 2012; Lin and Yu, 2011). The ESR assay employs the spin trap 5,5-dimethyl-1-pyrroline *N*-oxide (DMPO) and is specific for reactive radical species, which for example result from partial reduction of dioxygen catalyzed by transition metal (Boogaard et al., 2012; Shi et al., 2003). The AA assays would be more specific to the oxidative potential of transition metals (Godri et al., 2011; Yang et al., 2014), but ascorbic acid is known to react with organics such as quinones (Shang et al., 2012; Visentin et al., 2016). All of these assays have shown some correlations with health outcomes in epidemiological studies (Abrams et al., 2017; Bates et al., 2015; Fang et al., 2016; Strak et al., 2017; Weichenthal et al., 2016a, b; Yang et al., 2016). Detailed PM₁₀ characterization

was performed in parallel, by analyzing up to 130 chemical species that incorporated a broad array of organic species and trace elements (Chevrier, 2016).

In this paper, we report on seasonal variations within the redox activity assays as well as on the correlation among the different assays over a 1-year period. Univariate and multivariate regression analyses were applied in order to obtain (a) further insight into groups of chemical species that drive OP measurements and (b) evaluate whether differences could be detected between the individual assays.

2 Material and methods

2.1 Site description and sampling

In Europe, particulate matter sanitary alerts are based on PM₁₀ measurements. PM₁₀ was collected in the downtown area of Chamonix (45°55'21.53" N, 6°52'11.68" E; Auvergne-Rhône Alpes, France; 1035 m.a.s.l.) in the Alpine Arve valley. This urban location is heavily impacted in winter by biomass burning (wood combustion used for domestic heating) and traffic emissions. Further, because of their topography, specific weather conditions and anthropogenic activities, the European daily limit value for PM₁₀ is often exceeded in many sites in the Alpine valleys during the winter period (Chevrier, 2016), including this site in Chamonix.

In the framework of the DECOMBIO project (biomass burning contribution to PM₁₀ in the Arve valley) PM₁₀ sampling and detailed chemical characterizations have been achieved and are described elsewhere (Chevrier, 2016). Briefly, ambient particles were collected by filtration during 24 h (24 × 30 m³ h⁻¹) with a DIGITEL DA-80 on 150 mm quartz filters (Tissuquartz Pallflex) using the European standard protocol NF EN 16450. DIGITEL DA-80 was automatically programmed to stock before and after sampled filters, and the samples were then collected every week. The filters were calcined at 500 °C for 8 h before use. After sampling, the filters were folded, wrapped in aluminum foils, sealed in polyethylene bags, and stored at -25 °C until chemical analyses and at 4 °C until OP analysis. PM₁₀ mass measurements were achieved at the sampling site with TEOM-FDMS (tapered element oscillating microbalance-filter dynamics measurement system) as part of the regular Atmo-AURA network of air quality observation (<https://www.atmo-auvergnerhonealpes.fr/donnees/acces-par-station>).

2.2 Chemical analyses

2.2.1 PM₁₀ chemical composition

Briefly, collected PM₁₀ samples were measured for the following elements and components: elemental and organic carbon (EC, OC), BC and the distinction between wood burning (BC_{wb}) and fossil fuel BC (BC_{ff}), soluble anions and cations (NO₃⁻, SO₄²⁻, Cl⁻, MSA, oxalate and NH₄⁺,

Mg²⁺, Na⁺, Ca²⁺, K⁺), a large range of inorganic elements (Al, Fe, Ti, As, Ba, Cd, Ce, Cr, Cu, La, Li, Mn, Mo, Ni, Pb, Rb, Sb, Sn, Sr, V, Zn and Zr), sugar alcohols (arabitol, sorbitol, and mannitol, also called \sum Polyols), monosaccharide anhydrides (levoglucosan, mannosan and galactosan, \sum monosaccharides), humic-like substances (HULIS), and polar and apolar organics tracers (alkanes (\sum alkanes)), hopanes (\sum hopanes), methoxyphenols (\sum methoxyphenols), polycyclic aromatic hydrocarbons (\sum PAHs), substituted derivatives (methyl-PAHs) and sulfur polycyclic aromatic hydrocarbons (\sum PASHs). More detailed information is available in the Supplement (Sect. S1).

2.2.2 Oxidative potential assays

A total of 98 PM₁₀ samples collected, from November 2013 to October 2014, were analyzed for redox activity. Since studies have shown a nonlinear DTT response to both PM concentrations (Charrier et al., 2016) or from different chemical species added to the assay (Calas et al., 2017; Wang et al., 2017), for the DTT and AA assays (single compound assay), extractions were achieved for each sample to a final concentration of 10 $\mu\text{g ml}^{-1}$, allowing samples inter-comparison as same extractions at constant mass were used. For extraction procedure, PM samples were extracted using a Gamble + DPPC (dipalmitoylphosphatidylcholine) solution and vortexed at maximum speed during 2 h at 37°C (Calas et al., 2017). This solution allows for the extraction of PM in an environment closer to physiological conditions. Our previous results (Calas et al., 2017) also show an improvement of PM suspension during the assay, thus facilitating the OP DTT measurement. In this study, nonlinear response to PM concentrations was observed in the case of the DTT assay but not in the case of the AA assays (more details can be found in the Sect. S2 and Fig. S1). Conversely, for the RTLf and ESR assays, no extraction procedure was performed. Filter punches of 0.196 cm² were used in a “direct” measurement (cf. individual subsections for more information).

PM OP measurements with the DTT, AA (single compound assay) and ESR assays were performed at the Grenoble Alpes University in early 2016. The RTLf assay was measured at the King’s College of London between late 2014 and 2015.

DTT assay

A semi-automated procedure was used with a plate-reader TECAN spectrophotometer Infinite[®] M200 Pro and 96-well CELLSTAR[®] multiwell plates from Greiner Bio-One[®], the assay was modified from the DTT assay of Cho et al. (2005). DTT depletion was monitored for 30 min by adopting the following procedure: (1) measurement of the matrix absorbance (Abs_{mat}) of particles and substrate at 412 nm (205 μL of phosphate buffer and 40 μL of PM suspension), (2) injection of 12.5 nmol of DTT (50 μL of 0.25 mM DTT solution

in phosphate buffer), and (3) for each sample (lab blank included) quantification of DTT immediately ($t = 0$) and after 15 and 30 min of exposure (50 μL of 1 mM 5,5′-dithiobis(2-nitrobenzoic acid) (DTNB) in phosphate buffer) in triplicate. Positive controls consisted in a 1,4-naphthoquinone (1,4-NQ) solution and DTT depletions by 1,4-NQ (40 μL of 24.7 μM stock solution) were quantified only once at each measurement time because of the good repeatability between triplicates (Calas et al., 2017).

The rate of DTT loss (nmol min^{-1}) was determined from the slope of the linear regression of calculated nmol of consumed DTT vs. time. The amount of remaining DTT was calculated following Eq. (1):

$$n_{\text{DTT},i} = \frac{\text{Abs}_i \cdot n_{\text{DTT},0}}{\text{Abs}_{t0}}, \quad (1)$$

where $n_{\text{DTT},i}$ is the amount of DTT (nmol) at $t = i$, Abs_i is the absorbance at $t = i$, $n_{\text{DTT},0}$ is the initial amount of DTT (nmol) and Abs_{t0} is the absorbance at $t = 0$.

The linear regression was considered acceptable when $R^2 > 0.90$ and when less than 70 % of the initial amount of DTT had been oxidized (Li et al., 2009). Matrix absorbance (Gamble + DPPC solution and PM) was subtracted from the final absorbance and DTT loss in the lab blanks was subtracted from DTT loss of the samples in order to obtain the actual DTT depletion of the samples. Normalization per cubic meter (noted -v for volume), representative of human exposure, was chosen and OP DTTv ($\text{nmol DTT min}^{-1} \text{ m}^{-3}$) was calculated by multiplying DTT depletion per μg of PM added in the wells by PM₁₀ mass concentration ($\mu\text{g m}^{-3}$). The measurement quality was estimated by calculating the coefficient of variation (CV %) of positive controls (1,4-NQ). The CV was below 2 % ($n = 13$). For the DTT assay, a total of 98 samples were analyzed. Field blanks (six samples distributed over the sampling year) were also analyzed and observed contamination remained constant over the year (more details can be found in the Sect. S2 and Fig. S2).

AA assay (single compound assay)

A semi-automated procedure using the same plate reader than for the DTT assay was applied using Greiner UV-Star[®] 96-well plates; this assay was based on the modified assay from Zielinski et al. (1999) and Mudway et al. (2004). Matrix absorbance (Abs_{mat}) measurement of PM and substrate at 265 nm was performed (120 μL of Milli-Q water and 80 μL of PM suspension). Then, 24 nmol of AA (100 μL of 0.24 mM AA solution in Milli-Q water) was injected and absorbance was read at 2 min and then every 4 min for 30 min. Positive controls (80 μL of 24.7 μM 1,4-NQ solution) were quantified in duplicates. The rate of AA loss (nmol min^{-1}) was determined from the slope of the linear regression of calculated nmol of consumed AA vs. time. The amount of remaining AA was calculated in the same way as for DTT (Eq. 1). The linear regression was considered acceptable

when $R^2 > 0.90$. The matrix absorbance was subtracted from the final absorbance and the AA loss in the lab blanks (blank filter) was subtracted from the AA loss of the samples in order to obtain the actual AA depletion of the samples. AAV ($\text{nmol AA min}^{-1} \text{ m}^{-3}$) was calculated in the same way as for DTT. The measurement quality was estimated by calculating the coefficient of variation (CV %) of the positive control (1,4-NQ). The CV was below 2 % ($n = 7$). For the AA single compound assay a total of 98 samples were analyzed. Field blanks were also analyzed and no differences with lab blanks were observed (Sect. S2 and Fig. S3).

ESR assay

A modified ESR assay involving no filter extraction was used (Hellack et al., 2014). This alternative method is highly correlated with that involving filter extraction (Hellack et al., 2014; Yang et al., 2014). This approach is based on the generation of HO^\bullet radicals in the presence of hydrogen peroxide, via Fenton-type reactions and their trapping by the nitron DMPO. Filter punches (0.196 cm^2) were placed in 1 mL tubes, and 125 μL Milli-Q water, 125 μL H_2O_2 (0.5 M) and 250 μL of DMPO (0.05 M, ESR grade) were added. Tubes were subsequently vortexed for 15 s before being placed in incubation at 37°C for 40 min under agitation (detailed information in Sect. 3, Fig. S4). Suspensions were then vortexed again for 15 s and 35 μL of the suspension was transferred to a capillary tube to measure the hydroxyl radical (HO^\bullet) formation catalyzed by PM_{10} . The EPR spectra were recorded on a Bruker EMXplus spectrometer equipped with a high-sensitivity resonator operating at 9.44 GHz at 293 K. Preliminary results have shown a strong decrease in the ESR response when Gamble + DPPC solution was used instead of Milli-Q water, thus justifying the use of Milli-Q water in this study (Sect. S3 and Fig. S5). Filter blanks and a suspension of CuO (2.73 M) were used for background correction and as a positive control, respectively. The 26 % ($n = 7$) coefficient of variation of the positive control was attributed from changes in the dispersion of this insoluble form of Cu. Due to limited remaining samples, extractions could not be performed at identical concentrations for all samples. Corrections were applied to the ESR signal when nonlinear pattern vs. PM mass was observed (Sect. S3). For the ESR assay, a series of 75 samples only was analyzed due to sample availability. OP ESR was expressed in arbitrary units (A.U.) per cubic meter and will be further referred to as OP ESRv.

RTLTF assay

For the RTLTF assay, 0.5 mL of synthetic RTLTF containing equimolar concentrations (200 μM) of ascorbic acid (ASC), urate (UA) and reduced glutathione (GSH) were added to triplicate tubes containing 0.196 cm^2 filter punch and incubated for 4 h at 37°C under constant mixing and as preliminary results indicated a linear RTLTF response to PM concen-

trations added to the assay (internal report). The synthetic RTLTF was prepared in Chelex 100 treated high-performance liquid chromatography (HPLC) grade water (pH 7). Particle-free controls at 0 and 4 h (C0 and C4), together with negative (carbon black M120) and positive (CRM NIST1648a) controls, and lab filter blanks were incubated in parallel (Mudway et al., 2004). After 4 h incubation, the micro-tubes were immediately centrifuged at 13 000 rpm for 1 h at 4°C , followed by removal of aliquots into 100 mM pH 7.5 phosphate buffer (for GSH analysis) or 5 % meta-phosphoric acid (for ASC and UA analysis). All tubes were immediately stored at -70°C . GSH analysis was derived from the total glutathione and oxidized glutathione (GSSG) analysis using a spectrophotometric enzyme-linked DTNB recycling method based on a modified method described by Baker et al. (1990). The ASC and UA analysis used a reversed-phase HPLC system (HPLC, Gilson Scientific, UK) with electrochemical detection (EG&G, Princeton Applied Research, model 400) following the procedure described by Iriyama et al. (1984). Field blanks were used for background correction before estimating the consumption of antioxidants. The consumption rate (% OP) of the antioxidants ASC (% OP ASC), UA (% OP UA) and GSH (% OP GSH) were obtained by reference to the 4 h particle-free control (C4). Conversions from % OP to OP per μg of PM and then to OP per cubic meter were also calculated and will be referred to as OP GSHv and OP ASCv. Results for UA are not presented, since PM did not deplete urate (Mudway et al., 2004; Zielinski et al., 1999). The lab blanks and carbon black negative control displayed minimal (< 5 %) background oxidation. The positive PM control (CRM NIST 1648a) reacted with up to 35 % consumption of the ASC. The same 98 PM_{10} samples as those used in the AA (single compound assay) and the DTT assays were analyzed. Ascorbic acid depletion is then measured in the AA assay and in the RTLTF assay. We named them differently: AA in the AA assay and ASC in the RTLTF assay, for the difference of experimental design and results.

3 Data analyses

All statistical analyses were carried out using the R statistical software 3.4.0. Nonparametric Mann–Whitney tests were used in order to evaluate the statistical significance between the cold and the warm periods. Nonparametric Spearman's rank correlations (r_s) were chosen to assess the strength of possible monotonic relationship between the different OP values and the concentration of the different pollutants measured. The linear model (lm) function in R was used for the multiple regressions. P values < 0.05 were considered statistically significant.

Table 1. Median ratios between cold (late November to late February) and warm (late May to late September) periods for PM₁₀ mass concentration and OP measurements expressed per m³ (OPv).

	Cold/warm period (1st, 3rd) ¹
PM ₁₀ mass concentration	2.6* (2.3, 3.7)
OP DTTv	2.1* (1.7, 2.9)
OP AAv	7.1* (6.0, 9.1)
OP ESRv	0.8 (0.7, 1.4)
OP GSHv	5.0* (4.6, 5.8)
OP ASCv	8.3* (7.5, 8.7)

* $p < 0.001$, nonparametric Mann–Whitney test; ¹ 1st and 3rd quartiles of the temporal ratio.

4 Results and discussion

4.1 PM₁₀ mass concentration and temperature in Chamonix

Figure 1a shows the mass concentration of PM₁₀ over the sampling period in Chamonix, taking into account the days with filters analyzed for OP (Table S1). Corresponding temperatures are illustrated in Fig. 1b. The data are clearly characterized by two periods – a cold period (late November to late February, $n = 30$) and the warm period (late May to late September, $n = 29$), with average temperatures of 1.4 and 14.9 °C, respectively. PM₁₀ concentrations were 2.6 times higher during the cold period (Table 1) ($29 \pm 14 \mu\text{g m}^{-3}$ and $10 \pm 2 \mu\text{g m}^{-3}$ for the cold and warm periods, respectively). The significant differences (Fig. 1c) in PM₁₀ concentration between these two periods can be explained by different PM₁₀ sources and frequent temperature inversions in this narrow valley in winter. Investigation of the former using a positive matrix factorization (PMF) approach (Chevrier, 2016) indicates that, during winter, the dominant emission source is biomass burning (60 % of PM mass on average), with 10 % due to traffic and about 18 % related to secondary inorganic aerosols (SIA). In summer, the main sources are biogenic activity (40 %), SIA (35 %) and traffic (10 %).

4.2 OP temporal variation over the 12-month sampling period and during cold or warm periods

Methodological results of the nonlinear DTT response to PM concentrations are in agreement with Charrier et al. (2016) and were avoided thanks to isomass concentration extraction. For the AA, ASC and GSH assays, nonlinear issues were not encountered. Finally, the nonlinear ESR response to PM concentrations was solved with back correction of the ESR signal using a linear curve. Altogether these methodological points allowed avoiding the non-additive effects in OP assays and allowed PM samples inter-comparison.

The seasonal variation in OPv (OP measurements expressed per m³) is reported in Fig. 2a, c, e, g, i. Over-

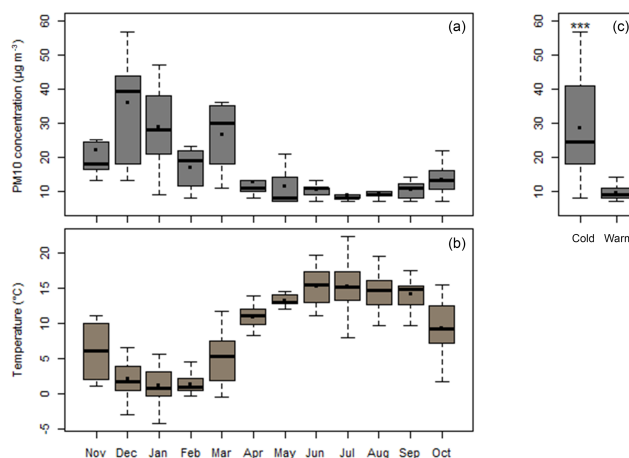


Figure 1. Temporal variation in (a) PM₁₀ mass concentration and (b) temperature over the 12-month sampling period. (c) Average PM₁₀ mass concentration during cold and warm periods. Boxplot representations with medians (horizontal line) and means (black square). *** $p < 0.001$, nonparametric Mann–Whitney test.

all, higher and more variable OPv values were observed in the cold period than during the warm period where overall values remain low and close to each other. However, patterns did differ depending on the OP measurement method used. Significant differences between cold and warm periods were observed for the DTT, AA, GSH and ASC assays (Fig. 2b, d, f, h, j), but not for OP ESR (Fig. 2f). For the ESR assay, the yearly pattern could be attributed to a possible scavenging effect of HO[•] by carbonaceous materials (Hellack et al., 2015) leading to the observed non-seasonal variation. Table 1 presents the median ratios calculated between the two periods. OP ESRv values were equivalent in the two periods with a ratio of the medians of 0.8 (close to 1 and the interquartile range includes 1) while for the other assays, the ratio was always higher than 2. The OPv DTT was only 2 times higher during the cold period compared with the warm period, whilst higher contrasts of 5, 7.1 and 8.3 were found using the GSH, AA and ASC assays, respectively.

4.3 Comparison of the OP measurement assays

Spearman correlations (r_s) between the assays were calculated over the sampling year. Correlations were considered as strong for $r_s > 0.70$ and moderate for $r_s (> 0.45 \text{ and } < 0.70)$, which are commonly criteria that can be found in the literature (Janssen et al., 2014; Yang et al., 2014). Table 2 shows that OPv values are all strongly correlated ($r_s > 0.7$) with the exception of OP ESRv values for which correlations ranged from 0.16 (OP ASCv) to 0.45 (OP DTTv). All assays are also strongly correlated with PM₁₀ ($r_s > 0.77$), except for OP ESRv ($r_s = 0.33$). The strong correlations between OP DTTv and OP AAv are in agreement with another study on PM₁₀ (Janssen et al., 2014), whereas they were not observed

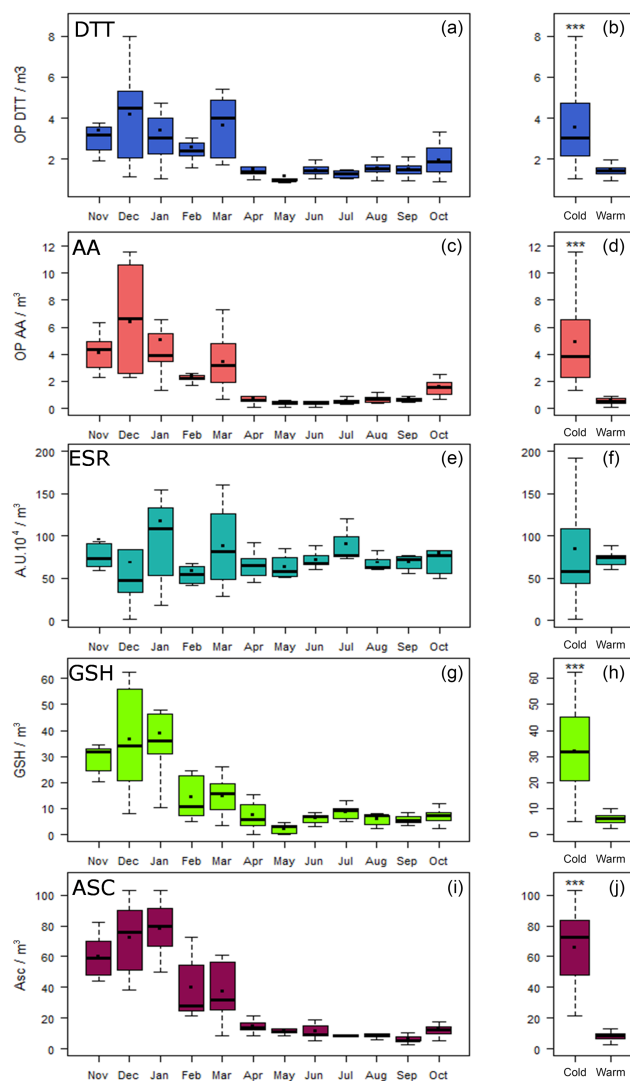


Figure 2. Seasonal variation of the five OPv (a, OP DTT; c, OP AA; e, ESR; g, OP GSH; i, OP ASC) and OPv differences between cold and warm periods (b, OP DTT; d, OP AA; f, OP ESR; h, OP GSH; j, OP ASC). Boxplot representations with medians (horizontal line) and means (black square). *** $p < 0.001$; nonparametric Mann–Whitney test.

by Fang et al. (2016) on $PM_{2.5}$. The weak correlations found between the OP ESR assay and both OP DTT and OP GSH assays are also in agreement with other studies on $PM_{2.5}$ (Künzli et al., 2006; Yang et al., 2014). However, the OP ESR assay is usually highly correlated with OP AA and ASC assays in both PM_{10} and $PM_{2.5}$ studies (Janssen et al., 2014; Künzli et al., 2006; Yang et al., 2014).

As shown in Table 3, these correlations also vary with the period, with higher r_s values during the cold period ($0.54 < r_s < 0.92$) than in the warm period for which the highest correlation ($r_s = 0.65$) was found between OP DTTv and OP AA. Temporal variations of correlations were also

observed by Fang et al. (2016) on $PM_{2.5}$. During the cold period, r_s between OP methods and PM_{10} ranged from moderate (0.59) for OP ESRv to strong (> 0.7) for the other OP measurements. During the warm period, a strong correlation was observed between PM_{10} and OP DTTv.

To better understand the evolution of OP results, OPv were related to the chemical composition of PM_{10} by using both univariate and multiple regressions analyses.

4.4 Univariate data analyses

Spearman correlations between OPv and the chemical composition of PM_{10} (expressed in $ng\ m^{-3}$ or $\mu g\ m^{-3}$) were calculated over the sampling year and are summarized in Table 4. The highest cumulative score of correlations above 0.45 is seen for OP DTTv (37/48; detailed information on cumulative scores is available in the Supplement, Sect. S4). Conversely, few correlations above 0.45 (5/48) are seen for OP ESRv. The other four assays (OP AA, OP GSH and OP ASC) exhibit very similar correlations with the chemical species.

Whereas correlations are evident between OP DTTv, OP AA, OP GSH, OP ASC, and some organic and inorganic species, in agreement with other studies regarding both $PM_{2.5}$ and PM_{10} (Janssen et al., 2014; Saffari et al., 2014; Yang et al., 2014), for OP ESRv correlations are limited to transition metals (Cr, Cu, Mo, Zr) and Ba. All these metals may be associated with traffic emissions, including brake wear (Hulskotte et al., 2014; Sanders et al., 2003). These results can be explained by the specificity of the ESR assay towards hydroxyl radical generation and in the case of Ba (not redox-active) linked to its co-emission with the former redox-active compounds. These results agree with the study of Boogaard et al. (2012) where the OP ESRv of PM collected near major urban road was highly correlated with Cr, Cu and Ba. However, OP ESRv is also associated with organic compounds (OC, PAH) in other studies (Janssen et al., 2014; Yang et al., 2014).

While many chemical species have a cumulative score of 4/5 over the sampling year, Cu is the only species with moderate to strong correlations with all OP measurements ($N = 5/5$). Several studies, including one in a subway system (Moreno et al., 2017) already pointed out the high impact of Cu concentrations on OP.

4.5 Multiple linear regression models

Multiple linear regressions were investigated in order to obtain further insight into the set of chemical species that can be among the dominant factors to the different OP measurements and for OP variations within time.

Such analysis requires the removal of extreme values and also requires transformations of the OP data in order to obtain distributions as close as possible to the normal one (detailed information about data set preparation can be found in

Table 2. Spearman correlation of the five OPv measurement assays over the sampling year.

	PM ₁₀ <i>n</i> = 98	OP DTTv <i>n</i> = 98	OP AAv <i>n</i> = 98	OP ESRv <i>n</i> = 75	OP GSHv <i>n</i> = 98	OP ASCv <i>n</i> = 95
PM ₁₀						
OP DTTv	0.88***					
OP AAv	0.81***	0.82***				
OP ESRv	0.33**	0.45***	0.27 *			
OP GSHv	0.77***	0.73***	0.80***	0.26 *		
OP ASCv	0.80***	0.72***	0.82***	0.16	0.80***	

*** $p < 0.001$ level, ** $p < 0.01$ level, * $p < 0.05$ level

Table 3. Spearman correlation of the OPv in the cold (upper part) and the warm period (lower part).

	PM ₁₀	OP DTTv ¹	OP AAv ¹	OP ESRv ²	OP GSHv ¹	OP ASCv ³
PM ₁₀		0.92***	0.91***	0.59***	0.87***	0.90***
OP DTTv	0.71***		0.89***	0.61***	0.79***	0.72***
OP AAv	0.43*	0.65***		0.54***	0.85***	0.79***
OP ESRv	0.088	0.17	0.36		0.56**	0.59**
OP GSHv	0.44*	0.29	0.36	0.63*		0.92***
OP ASCv	0.38*	0.37*	−0.072	−0.29	0.17	

*** $p < 0.001$ level, ** $p < 0.01$ level, * $p < 0.05$ level, ¹ $n = 30$ (cold period), $n = 29$ (warm period), ² $n = 30$ (cold period), $n = 14$ (warm period), ³ $n = 27$ (cold period), $n = 29$ (warm period).

the supporting information Sect. S5 and Table S2). The multiple linear regression models were obtained using the linear model (lm) function in R software. Forward variable selection and BIC number criteria were used to select the predictors that give the better explanation of the variance of the OPv (Sect. S5 model realizations and validations). The models obtained, for the set of samples excluding the extreme values, are presented in the following Eqs. (2, 3, 4, 5, and 6).

$$\begin{aligned} \log(\text{OPDTTv}) = & -0.440 + 0.101 \times \text{Cu} + 0.0259 \\ & \times \sum \text{PAH} - 0.0247 \times \text{Mg}^{2+} + 0.273 \times \text{As} + 0.318 \\ & \times \text{Mo} + 0.00549 \times \text{MSA} + 0.002 \times \sum \text{Polyols} \\ & + 0.00135 \times \text{Na}^+ \\ R^2 = & 0.81 \end{aligned} \quad (2)$$

$$\begin{aligned} \log(\text{OPAAv}) = & 0.433 + 0.000387 \\ & \times \sum \text{Monosaccharides} + 0.756 \times \text{Ni} \\ & - 0.372 \times \text{Mo} + 0.000538 \times \text{Fe} \\ R^2 = & 0.93 \end{aligned} \quad (3)$$

$$\begin{aligned} \log(\text{OPAAv}) = & -1.43 + 0.00163 \\ & \times \sum \text{Monosaccharides} - 0.00171 \times \text{Ca}^{2+} \\ & + 1.69 \times \text{Sb} + 0.117 \times \text{Cu} + 0.00208 \times \text{Na}^+ \\ & + 0.00119 \times \text{Fe} - 0.0435 \times \text{Zn} \\ R^2 = & 0.87 \end{aligned} \quad (4)$$

$$\begin{aligned} \text{OPESRv} = & 339953 + 73037 \times \text{Cu} - 6725 \\ & \times \sum \text{Alkanes} - 441351 \times \text{Cd} + 20795 \times \text{Ti} \\ R^2 = & 0.62 \end{aligned} \quad (5)$$

$$\begin{aligned} \sqrt{\text{OPGSHv}} = & 1.21 + 0.00161 \times \sum \text{Monosaccharides} \\ & + 0.210 \times \text{Cu} - 0.0642 \times \text{Mg}^{2+} + 0.0268 \times \text{MSA} \\ R^2 = & 0.48 \end{aligned} \quad (6)$$

$$\begin{aligned} \log(\text{OPASCv}) = & 1.79 + 0.000659 \times \text{NO}_3^- - 0.00733 \\ & \times \sum \text{Polyols} - 0.00292 \times \text{Na}^+ + 0.000247 \times \text{OC}^* \\ & + 0.103 \times \text{Cu} \\ R^2 = & 0.74 \end{aligned} \quad (7)$$

Table 4. Spearman correlations between the OPv measurement assays and the chemical composition of PM₁₀ (in bold: > 0.7, in italic: > 0.45).

	DTT	AA	ESR	GSH	ASC	N (5)
Cl ⁻	<i>0.68</i> ***	0.73 ***		<i>0.61</i> ***	0.77 ***	4
NO ₃ ⁻	<i>0.61</i> ***	<i>0.59</i> ***		<i>0.58</i> ***	0.74 ***	4
SO ₄ ²⁻	<i>0.33</i> ***		0.27*			
Na ⁺	<i>0.47</i> ***	<i>0.42</i> ***		<i>0.34</i> ***	<i>0.46</i> ***	2
NH ₄ ⁺	<i>0.38</i> ***	0.25*		<i>0.32</i> **	<i>0.33</i> ***	
K ⁺	0.83 ***	0.85 ***	0.34**	0.79 ***	0.77 ***	4
Mg ²⁺	<i>0.56</i> ***	0.30**	0.43***	0.30**	0.28**	1
Ca ²⁺	<i>0.53</i> ***	0.24*	0.32**	0.21*	0.32**	1
Al			0.24*			
As	<i>0.62</i> ***	<i>0.44</i> ***	0.31**	<i>0.43</i> ***	<i>0.41</i> ***	1
Ba	<i>0.68</i> ***	<i>0.46</i> ***	<i>0.57</i> ***	<i>0.42</i> ***	<i>0.35</i> ***	3
Cd	<i>0.64</i> ***	<i>0.65</i> ***		<i>0.63</i> ***	<i>0.72</i> ***	4
Ce	<i>0.50</i> ***	0.30**	0.35**	<i>0.33</i> ***	0.27**	1
Cr	<i>0.55</i> ***	<i>0.35</i> ***	<i>0.54</i> ***	<i>0.34</i> ***		2
Cu	0.87 ***	0.76 ***	<i>0.48</i> ***	0.70 ***	<i>0.64</i> ***	5
Fe	0.71 ***	<i>0.48</i> ***	0.44**	0.43**	0.38**	2
La	<i>0.44</i> ***	0.23*	0.37**	0.29**	0.27**	
Li	0.22*					
Mn	<i>0.53</i> ***	0.22*	0.41***	0.22*	0.21*	1
Mo	<i>0.65</i> ***	<i>0.38</i> ***	<i>0.46</i> ***	<i>0.40</i> ***	0.27**	2
Ni	<i>0.44</i> ***		0.37**			
Pb	<i>0.61</i> ***	<i>0.42</i> ***	0.30**	<i>0.43</i> ***	<i>0.37</i> ***	1
Rb	0.84 ***	0.76 ***	0.33**	0.70 ***	0.71 ***	4
Sb	0.79 ***	<i>0.66</i> ***	0.43***	<i>0.59</i> ***	<i>0.52</i> ***	4
Sn	0.70 ***	0.70 ***	0.29*	0.69 ***	0.83 ***	4
Sr	<i>0.55</i> ***	0.29**	0.44***	0.27**	0.28**	1
Ti	<i>0.36</i> ***		0.40***			
V		-0.28**	0.35**	-0.25*	-0.31**	
Zn	0.84 ***	<i>0.66</i> ***	0.40***	<i>0.65</i> ***	<i>0.64</i> ***	4
Zr	0.70 ***	<i>0.50</i> ***	<i>0.50</i> ***	<i>0.48</i> ***	<i>0.33</i> ***	4
BC	0.82 ***	0.90 ***	0.31*	0.75 ***	0.78 ***	4
BC _{wb}	0.77 ***	0.93 ***	0.27*	0.74 ***	0.87 ***	4
BC _{ff}	0.79 ***	0.84 ***	0.33**	0.70 ***	0.70 ***	4
OC	0.83 ***	0.87 ***	0.26*	0.81 ***	0.86 ***	4
DOC	0.79 ***	0.87 ***	0.29*	0.86 ***	0.87 ***	4
HULIS	0.85 ***	0.87 ***	0.26*	0.81 ***	0.86 ***	4
EC	0.82 ***	0.93 ***	0.25*	0.79 ***	0.84 ***	4
TC	0.84 ***	0.91 ***	0.26*	0.82 ***	0.87 ***	4
MSA	-0.26**	-0.53***		-0.36***	-0.34***	
Oxalate						
∑ polyols		-0.33***	0.38***	-0.27**	-0.51***	
∑ monosaccharides	0.74 ***	0.94 ***		0.78 ***	0.87 ***	4
∑ PAHs	<i>0.77</i> ***	0.92 ***		0.78 ***	0.88 ***	4
∑ alkanes	<i>0.63</i> ***	<i>0.61</i> ***		<i>0.54</i> ***	<i>0.77</i> ***	4
∑ methyl-PAHs	<i>0.67</i> ***	0.85 ***		0.72 ***	0.87 ***	4
∑ PASHs	<i>0.60</i> ***	0.76 ***		0.71 ***	0.76 ***	4
∑ hopanes	<i>0.66</i> ***	<i>0.67</i> ***		<i>0.58</i> ***	0.79 ***	4
∑ methoxyphenols	0.72 ***	0.93 *		0.77 ***	0.82 ***	4
N(48)	37	27	5	25	25	

*** $p < 0.001$ level, ** $p < 0.01$ level, * $p < 0.05$ level, $p < 0.10$ level. N = cumulative score of correlations (moderate and strong).

Two distinct models were associated for the AA single compound assay since the distribution is highly bimodal with two normal distributions: one in a cold period and one in a warmer period (Sect. S5 data set preparation). Strong coefficients of determination were found for the OP DTTv, AAv and ASCv (R^2 adj > 0.70), indicating that the variance of these OPv was well explained. For the ESRv and GSHv, R^2 values were lower: 0.62 and 0.48, respectively (Table S4).

Next, these models have then been applied to the general data set (Fig. S6), but, when strongly overestimated, values were removed again (Sect. S5 application to the overall data set). The models tend to underestimate OP values (negative intercept and slope = 0.84) for the ASC assay. For the other assays, the models tend to overestimate high OP values and to underestimate low OP values (negative intercept and slope between 1.11 and 1.51). However, the coefficients of determination range from 0.73 (OP ASCv) to 0.89 (OP AAv). Altogether, these results indicate that, on average, the models correctly represent the OP of PM participating in the ROS exposure during the overall year.

Finally, the contribution of each predictor during cold and warm periods has been investigated (Fig. 3) (Sect. S5 contribution of each predictor). The intercepts, attributed to unknown species, were significantly > 0 in all models. Moreover, for some species, a negative contribution was found (Table S4) that can be explained by an antagonist effect of some atmospheric components on OP: soot particles for Hellack et al. (2015), Gram-positive bacteria for Samake et al. (2017) or metal–organic binding interactions (Verma et al., 2012; Tuet et al., 2016; Wang et al., 2017), or because of the weighting assignation of species in the models. During the cold period, traffic tracers or indicators (like Cu, Fe, Mo, Ti, or \sum PAHs) are important positive factors to explain all OP measurements. Also, and with the exception of the ESR assay, biomass burning tracers (\sum Monosaccharides, including levoglucosan) or indicators such as \sum PAHs or OC* (corresponding to total OC minus the molecular species measured), both strongly related to biomass burning emission (Bonvalot et al., 2016; Chevrier, 2016), are prominent positive factors for all assays for this cold period. These results agree with the observations on the seasonal evolution of OPv, with much larger values in the cold period when biomass burning emissions are dominant. Additionally, literature results on DTT reactivity towards organic compounds indicate higher impact of biomass burning species when compared to other organic compounds (Verma et al., 2015). During the warm period, traffic tracers and especially copper are important positive factors in OP measurements. In addition, road dust and industrial tracers (Zn, Ca^{2+}) for the AA assay as well as primary biogenic indicators (\sum polyols, MSA) for the DTT and GSH assays seem to be prominent factors of these OP measurements.

Several limitations can be attributed to this study. Most important, all of these results have been obtained for a specific location and cannot be generalized as chemical composition

of PM₁₀ strongly differs from one location to another. PM₁₀ chemistry is different from PM_{2.5} and the associations reported here are only valid for PM₁₀. Some components that might mainly reside in the coarse mode are positive factors in the multiple linear regression models (e.g., Ti in OP ESRv). They can display a different final health impact, since a fraction of PM₁₀ does not penetrate all the way to the lung. Also, the results of the ESR assay warrant caution due to our back correction of the ESR signal linked to the nonlinear response of the assay. Finally, the multiple model result for the GSH assay should be considered with caution since a normal distribution was not reached in the first step of the analysis. Moreover, multiple nonlinear regression models should also be investigated since several studies have shown that oxidative potentials from different PM components are not always additive (Tuet et al., 2016, 2018; Wang et al., 2017; Xiong et al., 2017; Yu et al., 2018). Finally, these analyses are only relevant for PM₁₀ when some health studies are now taking PM_{2.5} into account. Additional studies addressing the comparison of OP results associated with PM₁₀ and PM_{2.5} are needed (Gali et al., 2017; Styszko et al., 2017).

However, all results point out that biomass burning and vehicular emissions in winter are the main sources correlated with ROS generation. In summer, ROS burden is associated with vehicular emissions, biogenic emissions, road dust and industrial sources. All these results also suggest the association of assays in order to take into account this wide range of OP determinants. To achieve this, the DTT and the AA assays, or the DTT and the RTLf assays, can be associated to get the best information, which is in agreement with other studies on acellular OP measurements (Janssen et al., 2014; Yang et al., 2014). However, a definitive proposition for the best association of assays will most probably come from a final benchmark against epidemiological study outcomes (Fang et al., 2016; Strak et al., 2017; Weichenthal et al., 2016a).

5 Conclusions

Our results show that most of the OP assays were strongly intercorrelated over the sampling year but also these correlations differed when considering specific sampling periods (cold vs. warm).

All acellular assays are correlated with a significant number of chemical species when considering nonparametric rank correlations, especially for the DTT assay. Finally, copper seems to be a unifying determinant in the OP assays.

Evidence is also presented of a seasonal contrast over the sampling period with significantly higher OPv values during winter for the DTT, AA, GSH and ASC assays, which were assigned to biomass burning species by the multiple linear regression models. However, during the cold period and for the DTT assay, \sum PAHs, which can be associated with both traffic and biomass wood burning emissions, were found to be

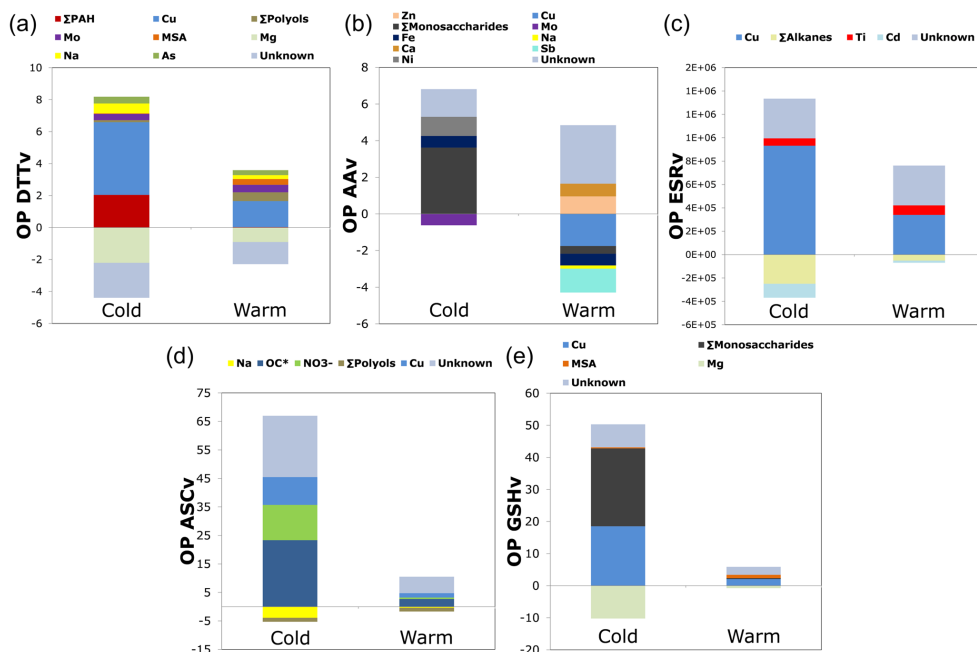


Figure 3. Multivariable linear regression analyses results. Contribution of predictors during cold and warm periods (a, OP DTTv; b, OP AAV; c, OP ESRv; d, OP ASCv; e, OP GSHv). The intercepts, attributed to unknown species, were significantly > 0 in all models. Some species were assigned with negative contributions, explained by a weighting assignment of species so as to model OP differences between warm and cold periods or a possible antagonist effect of some atmospheric components on OP contribution.

significant factors. The ESRv clearly differs from the other tests as it did not show seasonal dynamics and presented weaker correlations with other assays and with the chemical species. Nevertheless, ESR assay results are mostly associated with traffic tracer species. Finally, the combination of two models was used to fit the results of the AA assay, which is necessary to provide the best explanation of OP variance, with a bimodal distribution of the initial measurements. This indicates that the strong changes in the chemistry of the PM are probably leading to nonlinear processes in the link between chemistry and OP.

Overall, these results suggest the combination of the assays in order to take into account a wide range of determinants of OP. In the case of the Chamonix city, DTT associated with AA assays or DTT combined with RTLF assay were able to provide the most exhaustive information about OP determinants (Cu) and sources associations (biomass burning, vehicular emissions).

Rank correlations and multiple linear regressions are useful tools to determine the most prominent species driving the redox activity of ambient PM. However, to go further in identifying the assay or combination leading to the best information relying on source dynamics, multiple linear regressions analysis requires large data sets. Finally, more source apportionment approaches through positive matrix factorization methods are needed in order to assign dominant PM emission sources in the OP assays

Data availability. Data are available upon request.

Supplement. The supplement related to this article is available online at: <https://doi.org/10.5194/acp-18-7863-2018-supplement>.

Competing interests. The authors declare that they have no conflict of interest.

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