Supplementary Information

Comparative assessment of vaccine vectors encoding ten malaria antigens identifies two protective liver-stage candidates

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Figure S1. ChAd63-MVA *P. falciparum* vaccines can provide protection in BALB/c mice. (a-j) Eight vaccinated and eight naïve mice were challenged with 1000 chimeric sporozoites i.v. The Kaplan-Meier curves illustrate the time to 1% parasitaemia, whilst statistical significance between the survival curves was assessed using the Log-Rank (Mantel-Cox) Test, (a) PfCSP p=0.03, (b) PfTRAP p=0.3, (c) PfLSAP1 p=0.2, (d) PfETRAMP5 p=0.3, (e) PfCeITOS p=0.03, (f) PfUIS3 p=0.0001, (g) PfLSAP2 p<0.0001, (h) PfFalstatin p=0.007, (i) PfLSA1 p<0.0001 and (j) PfLSA3 p=0.01. For the PfLSA3 challenge, the chimeric sporozoite dose was increased to 2000 sporozoites per mouse in order to infect all naïve controls.



Figure S2. ChAd63-MVA *P. falciparum* vaccines can provide protection in CD-1 mice. (a-j) Eight to ten vaccinated and eight to ten naïve mice were challenged with 1000 chimeric sporozoites i.v. The Kaplan-Meier curves illustrate the time to 1% parasitaemia, whilst statistical significance between the survival curves was assessed using the Log-Rank (Mantel-Cox) Test: (a) PfCSP p=0.001, (b) PfTRAP p=0.02, (c) PfLSAP1 p=0.16, (d) PfETRAMP5 p=0.45, (e) PfCeITOS p=0.097, (f) PfUIS3 p=0.25, (g) PfLSAP2 p=0.0009 (h) PfFalstatin p<0.0001, (i) PfLSA1 p<0.0001 and (h) PfLSA3 p=0.15. For the PfLSA3 challenge, the chimeric sporozoite dose was increased to 2000 sporozoites per mouse in order to infect all naïve controls.



Figure S3. All chimeric *P. berghei* parasite clones contain the correct genotype.
(a) Schematic representation showing the introduction of the constructs that contain the '*P. falciparum* antigen expression-cassette' into the 230p locus of the *P. berghei*

ANKA GIMO mother line by GIMO-transfection using negative selection (5-FC). Black arrows: location of PCR primers used for diagnostic PCR-analysis (see panel c). CDS, coding sequence. (b) Southern analysis of chromosomes (Chr) of chimeric parasite lines separated by pulsed-field gel electrophoresis to confirm integration of the DNA construct in the GIMO locus (*230p* on Chr 3), shown as the removal of the h*dhfr::yfcu* SM cassette in cloned chimeric parasites compared to a control probe recognising Chr 5. As an additional control (Ctrl), parasite line 2117cl1 is also shown as it retains h*dhfr::yfcu* SM in the *230p* locus on Chr 3. (c) Diagnostic PCR analysis of chimeric parasite lines confirming correct integration of the *P. falciparum* antigen expression cassettes. Correct integration in all lines is shown by the absence of the h*dhfr::yfcu* SM, the presence of the *P. falciparum* gene coding sequence and the correct integration of the construct into the genome at both the 5'and 3'regions (5' int and 3' int).



Figure S4. All chimeric *P. berghei* parasites express the inserted antigen at the liver-stage of infection. As each chimeric parasite also contained a GFP::luciferase reporter cassette, luciferase expression in chimeric *P. berghei* parasites was measured at 44 hours post-injection of 1000 sporozoites i.v. into seven to eight BALB/c mice, using the IVIS 200 *in vivo* imager. All chimeric parasites expressed luciferase at the liver-stage, providing an indirect confirmation of antigen expression at the liver-stage.

	Chimeric P. berghei	WT P. berghei		
PbCSP	1	e voe		
PfLSA1	5.5	10 10		
PfLSA3	5 1			
PfCelTOS	$\langle \rangle$			
PfUIS3) ~	÷		
PfLSAP1	3 B			
PfLSAP2	()			
PfETRAMP5	γ. ζ. →			
PfFalstatin	* .			
PfCSP	> , [′]			
PfTRAP	, ~ .	at de		

Figure S5. All chimeric *P. berghei* **sporozoites express the inserted** *P. falciparum* **antigen.** Chimeric salivary-gland sporozoites were stained with sera from vaccinated mice or with monoclonal antibodies where available (PbCSP 3D11 and PfCSP 2A10), with green fluorescence indicating the presence of the protein (Alexa Fluor 488). As a control, wild-type (WT) *P. berghei* sporozoites were stained with the same antibodies or sera.



Figure S6. Infectivity of the chimeric parasites in mice compared to wild-type *P. berghei.* Time to 1% parasitaemia of the chimeric parasites compared to wild-type parasites following injection of 1000 sporozoites i.v, into (**a**) 7-22 BALB/c and (**b**) 8-10 CD-1 mice. Both median and individual data points are shown. All chimeric parasites are statistically comparable to wild-type (Mann-Whitney test), except for PfUIS3, p<0.0001, and PfLSA3, p=0.02, both in BALB/c, and PfCSP, p=0.009, and PfLSA3, p=0.0004, in CD-1 mice.

Antigen	Gene ID ^a	Size ^b	Predicted Structure	tPA	Final size ^b
PfLSAP1	PF3D7_1201300	318	Signal peptide, two	No	347
			transmembrane domains		
PfETRAMP5	PF3D7_0532100	543	Signal peptide,	Yes	662
			transmembrane domain,		
			repetitive region		
PfCelTOS	PF3D7_1216600	546	Signal peptide	Yes	665
PfUIS3	PF3D7_1302200	687	Signal peptide, two	No	716
			transmembrane domains		
PfLSAP2	PF3D7_0202100	906	Non-secretory,	Yes	1025
			transmembrane domain		
PfFalstatin	PF3D7_0911900	1239	Signal peptide	Yes	1358
PfLSA1	PF3D7_1036400	3486	Signal peptide, repetitive	Yes	1502
			regions		
PfLSA3	PF3D7_0220000	4674	Non-secretory, two	No	4259
			transmembrane domains,		
			repetitive regions		

Table S1.	Vaccine	construct	details.
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^a PlasmoDB Gene ID. ^b Size in base pairs.