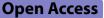
REVIEW



From dysbiosis to defense: harnessing the gut microbiome in HIV/SIV therapy

Jason M. Brenchley^{1*} and Sergio Serrano-Villar^{2*}

Abstract

Background Although the microbiota has been extensively associated with HIV pathogenesis, the majority of studies, particularly those using omics techniques, are largely correlative and serve primarily as a basis for hypothesis generation. Furthermore, most have focused on characterizing the taxonomic composition of the bacterial component, often overlooking other levels of the microbiome. The intricate mechanisms by which the microbiota influences immune responses to HIV are still poorly understood. Interventional studies on gut microbiota provide a powerful tool to test the hypothesis of whether we can harness the microbiota to improve health outcomes in people with HIV.

Results Here, we review the multifaceted role of the gut microbiome in HIV/SIV disease progression and its potential as a therapeutic target. We explore the complex interplay between gut microbial dysbiosis and systemic inflammation, highlighting the potential for microbiome-based therapeutics to open new avenues in HIV management. These include exploring the efficacy of probiotics, prebiotics, fecal microbiota transplantation, and targeted dietary modifications. We also address the challenges inherent in this research area, such as the difficulty in inducing long-lasting microbiome alterations and the complexities of study designs, including variations in probiotic strains, donor selection for FMT, antibiotic conditioning regimens, and the hurdles in translating findings into clinical practice. Finally, we speculate on future directions for this rapidly evolving field, emphasizing the need for a more granular understanding of microbiome-immune interactions, the development of personalized microbiome-based therapies, and the application of novel technologies to identify potential therapeutic agents.

Conclusions Our review underscores the importance of the gut microbiome in HIV/SIV disease and its potential as a target for innovative therapeutic strategies.

Background

During progressive human immunodeficiency virus (HIV) infection of humans and Simian immunodeficiency virus (SIV) of Asian macaque nonhuman primates (NHPs), ongoing viral replication and systemic

*Correspondence:

Jason M. Brenchley

jbrenchl@mail.nih.gov

Sergio Serrano-Villar

sergio.serrano@salud.madrid.org

 $^{\rm 1}$ Barrier Immunity Section, Lab of Viral Diseases, NIAID, NIH, Bethesda, MA, USA

² Department of Infectious Diseases, Hospital Universitario Ramon y Cajal, IRYCIS and CIBERInfec, Madrid, Spain

inflammation contribute to the development of immunodeficiency, and most infections, if left untreated, will lead to individuals becoming susceptible to opportunistic infections. In people with HIV (PWH) treated with antiretroviral therapy (ART) and with sustained viral suppression, residual inflammation contributes to increased susceptibility to cardiovascular disease and development of some cancers [1]. Thus, there has been significant interest in understanding the causes of residual inflammation in PWH on ART.

The causes of inflammation in progressive HIV and SIV infections are multifaceted and include the following: co-infections [2], HIV or SIV themselves [3], and microbial translocation [4]. However, the degree to which each



This is a U.S. Government work and not under copyright protection in the US; foreign copyright protection may apply 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicate dotherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/byl4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/ zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data. of these contributes to inflammation in PWH and SIVinfected Asian macaques on ART is unclear. Therefore, targeted therapeutic interventions have been considered to specifically reduce individual causes of inflammation in PWH and SIV-infected NHPs. Here, we review therapeutic interventions to reduce microbial translocationinduced inflammation in PWH or SIV-infected Asian macaques.

Normal functions of the gut microbiome

The microbiota of the gut comprises approximately 10¹⁴ diverse bacterial organisms. Bacterial genes are at least 100 times greater than the human genome. This microbiota consists of approximately 1000 species of bacteria, belonging to two main groups: the Bacillota and the Bacteroidota. Humans have coevolved with the gut microbiota to maintaining a symbiotic relationship. Indeed, germ-free animals are more susceptible to infections and have reduced vascularity, digestive enzyme activity, muscle wall thickness, serum immunoglobulin levels, smaller Peyer's patches, and fewer intraepithelial lymphocytes (reviewed in [5]). The beneficial roles of the gut microbiota are multifactorial and include immunological, structural, and metabolic functions [6].

The microbiota competes with pathogenic bacterial species and metabolizes dietary carbohydrates into short-chain fatty acids, a major nutritional source for the colonic epithelia [7]. Finally, most plasma cells secrete dimeric or oligomeric IgA, which can transcytose directly into the lumen. These antibodies are specific for luminal bacteria and viruses, suggesting that the microbiota shapes the specificity of luminal IgA [8]. Taken together, the microbiota provides invaluable functions to the host.

The gastrointestinal tract in HIV/SIV infections

Early after HIV was discovered as the causative agent of acquired immunodeficiency disease syndrome (AIDS), significant gastrointestinal manifestations were noted [9]. The observed abnormalities including malabsorption and lymphocyte depletion led to the conclusion as follows: "The histologic findings suggest that a specific pathologic process occurs in the lamina propria of the small intestine and colon in some patients with the syndrome" [9]. This finding was incredibly insightful in its anticipation of subsequent discoveries. Over the years, we have learned that the mechanisms underlying the observed gut abnormalities are multifaceted. Indeed, there is overwhelming evidence to suggest that CD4+T cells are disproportionately infected by the virus and depleted from the lamina propria of the gut compared to other anatomic sites of PWH and SIV-infected Asian macaques [10–16]. Moreover, progressive HIV/SIV infections lead to a massive depletion of CD4 + T cells in the gut of HIV/

SIV-infected individuals, especially of CD4+T cells producing the effector cytokines IL-17 and/or IL-22 [17–19]. Most epithelial cells in the body express the IL-22 receptor, and IL-22 is a growth factor supporting their survival and division. Moreover, IL-17 is a chemoattractant for neutrophils to anatomic sites where bacterial and fungal antigens are present [20–23]. Thus, preferential loss of lymphocytes producing these effector cytokines may provide mechanistic insights into damage to the structural barrier of the gut [20–23].

Is there an HIV-associated dysbiosis?

In addition to the observed immunological perturbations being hypothesized to contribute to HIV-associated enteropathy, increasing information suggests that the composition of the gut microbiome can also contribute. Intestinal microbial dysbiosis — an imbalance in the composition of the individual bacteria within the gut microbiome — has been noted in PWH from different geographic locations [24–30], reviewed in [31]. However, dysbiosis of the bacterial gut microbiome in SIV-infected macaques is less obvious [32–36], suggesting confounding variables may contribute to the bacterial dysbiosis observed in PWH [37–39].

Indeed, alteration to the composition of the gut bacterial microbiome seems to contribute to many diseases, ranging from conditions which affect the gut itself [40] to neurological diseases [41]. Understanding mechanisms underlying bacterial dysbiosis observed in outbred populations of humans is complicated by the multiple confounding variables that can influence susceptibilities to particular diseases and the composition of the gut microbiome. Recent meta-analyses suggested that, in many cases, dysbiosis associated with diseases were attributed to confounding variables such as age, sex, body mass index, diet, and alcohol use [42]. Indeed, in microbiome analyses of PWH after controlling for risk factors associated with HIV acquisition, recent studies have demonstrated that the degree to which the microbiome becomes dysbiotic is predicted by the nadir CD4+T-cell count [39]. These data suggest that the greater immune impairment associated with HIV infection contributes to dysbiosis.

Vertical transmission of a dysbiotic microbiome to HIV-exposed but uninfected infants may contribute to their increased morbitidies [43]. Research indicates variations in the microbiome composition, including fecal samples, between children born to HIV-positive mothers compared to those from HIV-negative mothers [44–46]. These microbial differences between HIV-exposed and unexposed children seem to mirror those found in their mothers and may be moderated by breastfeeding [47]. This suggests a possible transmission of the maternal

microbiome, although it remains unclear whether these patterns are directly due to microbial transfer or influenced by environmental or lifestyle factors related to living with HIV, such as treatment with cotrimoxazole.

The impact of ART on the gut microbiome in PWH is another growing area of study, complicated by ART's association with immunological improvement and the diversity of treatment combinations and indication bias. While some studies have begun to unravel the effects of ART on the microbiome, results remain varied. For example, in a longitudinal study in PWH starting ART based on integrase inhibitors or non-nucleoside reverse transcriptase inhibitors followed for 96 weeks, participants exhibited only minor changes in microbiota composition, regardless of the drug class [48]. Similarly, in another study in PWH starting ART at an advanced disease state, no substantial changes on gut microbiota composition were observed after 48 weeks of follow-up [49]. A recent study in Zimbabwe highlighted the role of geographical and baseline microbiota factors in ART's effects, particularly in rural areas where ART with cotrimoxazole led to a reduction in alpha diversity after 24 weeks, with more significant changes in individuals having higher baseline diversity [50]. This suggests a potential direct drug effect on microbiome diversity. Indeed, a recent in vitro study elucidates an unexpected facet of ART — its antibacterial activity. Certain antiretroviral drugs exhibited in vitro antibacterial action against gut and vaginal commensal bacteria, particularly against multidrug-resistant strains [51]. Collectively, these findings indicate that ART's influence on the gut microbiome is complex and requires further research to fully grasp its implications on the health of PWH.

Are HIV-associated changes in the microbiome a harmful adaptation?

Recent advancements in profiling the functions of the microbiome, using omics technologies such as transcriptomics, proteomics, and metabolomics, have shed light on its relationship with inflammation. One study revealed that certain bacteria, including Bifidobacteria, exert functions with a crucial role in preventing mucosal defects and reducing microbial translocation, thereby aiding immune recovery. Others, like Succinivibrionaceae and Erysipelotrichaceae, buffer pro-inflammatory molecules and accumulate antiviral compounds, highlighting their potential to reduce inflammation and inhibit viral replication [52]. A study involving women with or at high risk of HIV focused on gut microbial species in relation to carotid artery atherosclerosis. It found associations between certain microbes like Fusobacterium nucleatum and carotid artery plaque, along with a microbial metabolite, imidazole-propionate (ImP), linked to plaque and pro-inflammatory markers [53].

Moreover, in a study combining metagenomics and metatranscriptomics, we found that the HIV-associated microbiota adapts to an inflamed environment by overexpressing oxidative stress resistance pathways but also shifting towards pro-inflammatory gram-negative bacteria enriched in lipopolysaccharide biosynthesis pathways and potentially pathogenic strains [54]. These findings suggest intricate interactions between bacterial metabolism and HIV immunopathogenesis. Furthermore, they pose a thought-provoking question: might what we perceive as dysbiosis actually represent an adaptation of the microbiome for the host, with the balance between beneficial and harmful adaptations contributing to the extent of persistent inflammation in PWH?

Additionally, the role of the microbiome in HIV elite controllers offers fascinating insights. A study investigating the fecal metabolome and microbiome in elite controllers revealed an enrichment of certain dipeptides and the genus *Prevotella*, suggesting a unique microbiome profile that might contribute to their natural control of HIV replication [55]. Another study found that ECs have a richer gut microbiota with distinct bacterial signatures and metabolic profiles, potentially playing a role in their ability to control HIV [56]. These findings highlight the potential of leveraging microbiome insights to develop novel therapeutic strategies aimed at enhancing immune control against HIV.

Therapeutic interventions

To better understand the degree to which dysbiosis (irrespective of the cause, disease, or underlying anatomic or physiological predispositions to disease) contribute to inflammation and pathology, interventional studies aimed at reducing dysbiosis are powerful tools. Here, we review microbiome-altering therapeutic interventions administered to PWH and SIV-infected NHPs, how these have resulted in microbiome alterations and influences they've had on immunity, and future directions for microbiome-directed therapies in PWH (Fig. 1).

Antibiotics in SIV-infected NHPs and PWH Previous research in NHPs

The most common family of therapeutic interventions aimed at altering the microbiome's composition is antibiotics and is commonly used to reduce levels of bacteria thought to contribute to disease. Oral antibiotics significantly influence the composition of the gut microbiome [57, 58]. In SIV-uninfected NHPs, antibiotics have been experimentally given, followed by longitudinal assessment of the microbiome and immunological function. Enrofloxacin, cephalexin, paromomycin, and clindamycin

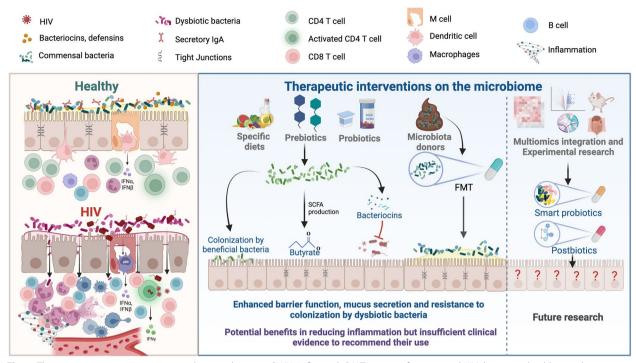


Fig. 1 Therapeutic interventions targeting the microbiome in PWH. Left panel: GALT immune function and HIV disruption: healthy conditions, complex microbiome and balanced immune response in the GALT; HIV effects, altered microbiome, damaged enterocyte barrier, increased bacterial translocation, CD4 T-cell loss, CD8 T-cell expansion, neutrophil infiltration, local and systemic inflammation. Right panel: Intervention strategies and future directions: diet and prebiotics, Mediterranean diet and prebiotics promote growth of beneficial bacteria; probiotics, targeted addition of beneficial bacteria; FMT, replacing patient's microbiota with a donor's healthy microbiome; current state of evidence, interventions potentially reduce inflammation but lack sufficient clinical backing for recommendation; research prospects, future studies aim to produce targeted microbial consortia or postbiotics with heightened precision and efficacy

all individually disrupt the composition of the gut microbiome, reduce levels of bacteria producing short-chain fatty acids, and induce some level of inflammation [58]. Importantly, the sources of NHPs that provide animals for biomedical sciences are diverse, and gut symptomologies are common in animals as they are transferred across facilities [57]. Indeed, we previously found significant alteration to the gut microbiome as NHPs are transferred from outdoor provisioned environments to indoor facilities [59]. In an attempt to ameliorate any alteration to the gut microbiome associated with acclimatizing to new environments, combinatorial use of enrofloxacin, paromomycin, fenbendazole, and azithromycin has been considered before study enrollment [57]. This multimodal, broad-spectrum, antibiotic use led to significant alterations to the gut microbiome that were fairly long-lived. Moreover, antibiotic use was associated with decreased gut inflammation and more consistent intrarectal SIV acquisition, but its influence on SIV disease progression was not analyzed.

Based on our current knowledge of microbial-immune interaction in PWH- and SIV-infected NHPs, significant alteration to the gut microbiome is anticipated to affect progressive HIV or SIV infections. Indeed, treatment of NHPs with vancomycin, an antibiotic against grampositive bacteria, led to significantly decreased levels of bacteria within the Bacillota phylum (which produce energy-rich short-chain fatty acids) and increased levels of bacteria within the Pseudomonadota phylum (which contain highly motile and potentially pathogenic gram negative facultatively anaerobic rods) [60]. While repetitive vancomycin treatment maintained this dysbiotic microbiome and resulted in a significant reduction in the continuity of the epithelial barrier of the gut, surprisingly, there was no influence on canonical metrics of SIV disease progression [60] (Table 1). However, subsequent work demonstrated that vancomycin-induced dysbiosis was sufficient to increase the number of transmitted founder viruses after low-dose intrarectal SIV challenge in NHPs [61]. Another study in acutely SIV-infected macaques found that early treatment with rifaximin and sulfasalazine reduced microbial translocation and immune activation. Still, this effect diminished over time and was ineffective in chronically infected macaques [62].

Population	Intervention	Study design and sample size	Outcomes	Ref
Rhesus macaques	Vancomycin	5 SIV + vancomycin 5 SIV control	Increased damage to GI tract epithelium	[60]
Rhesus macaques	Vancomycin	6 vancomycin before SIV challenge 6 control	Increased transmitter founder viruses after vancomycin	[61]
Pigtail macaques	Probiotics	5 (SIV + ART alone) 5 (SIV + ART + probiotics)	Increased GI tract CD4+T-cell reconstitution	[63]
Pigtail macaques	Probiotics + IL-21	5 (SIV + ART alone) 5 (SIV + ART + probiotics + IL-21)	Increased GI tract CD4+T-cell reconstitution, transient Th17 increases	[64]
Rhesus macaques	FMT	6 FMT + SIV 6 control SIV	Increased Th17 cells	[65]
Rhesus macaques	High-fat diet	8 HFD+SIV 52 historical SIV controls	Increased SIV disease progression	[96]
Pigtail macaque	High-fat diet	7 historical SIV controls 5 HFD+ SIV 3 HFD	Increased SIV disease progression	[67]
Rhesus macaques	Butyrate	6 (SIV + ART alone) 7 (SIV + ART + butyrate)	Slight increase in jejunal H3 acetylation and modest microbiome alterations	[68]
PWH on ART	Fermented skimmed milk supplemented with Lactobacillus rhannosus GG (108 cfu/ mL), Bifidobacterium animalis subsp. Lactis B-12 (108 cfu/mL), and Lactobacillus acido- philus La-5 (107 cfu/mL) Duration: 8 weeks	Randomized, double-blind, placebo con- trolled trial N = 25 Probiotics: N = 12 Placebo: N = 7 Control: N = 6	Mild D-dimer, CRP, and IL-6 reduction. No changes in microbial translocation (LPS, sCD14) or T-cell activation markers	[69]
PWH on ART	Saccharomyces boulardii (two capsules three times a day or 6 × 10 ⁷ living bacteria) Duration: 12 weeks	Randomized,d ouble-blind, placebo con- trolled trial N = 35 Probiotics: N = 19 Placebo: N = 16	Decreased LBP No changes in inflammation markers or T-cell counts	[70]
PWH diagnosed with CD4 < 350 cells/µL or AIDS starting ART	A synbiotic mixture of prebiotics, <i>Saccha-</i> <i>romyces boulardii</i> , oligonutrients, essential amino acids, omega-3 fatty acids Duration: 48 weeks	Randomized, double-blind, placebo con- trolled trial N = 59 Synbiotics: N = 32 Placebo: N = 27	No effects on primary (CD4 count, CD4/ CD8 ratio) or secondary (T-cell activation, bacterial translocation, inflammation, or fecal microbiota structure) endpoints	[49]
PWH on ART with CD4 < 500 cells/ µL and < 50 cells/µL increase 6 months before enrolment	L <i>actobacillus casei</i> Shirota Duration: 12 weeks	Randomized, double-blind, placebo con- trolled trial N=45 • Probiotics: N=23	No effects on CD4 count, CD4/CD8 ratio, T-cell activation, or sCD14	[7 1] 714,353

Population	Intervention	Study design and sample size	Outcomes	Ref
PWH on ART on ART with CD4 > 200 cells/µL	Probiotic visbiome For weeks 2–4, one sachet orally daily For weeks 4–26, one sachet orally twice daily	Randomized, double-blind, placebo con- trolled trial N=73 • Probiotics: N=42 • Placebo: N=31	No effect on primary (sCD14) or secondary (markers of inflammation and gut perme- ability) enpoints	[72]
PWH on ART and CD4 < 350/ counts µL despite virologic suppression	Probiotic visbiome Duration: 48 weeks	Randomized, double-blind, placebo con- trolled trial • Probiotics: <i>N</i> = 18 • Placebo: <i>N</i> = 10	No effect on primary outcome (CD8 activa- tion: HLA-DR+/CD38 + cells) or secondary (markers of inflammation, immune recon- stitution, bacterial translocation, and gut permeability) endpoints CD4 + T-cell activation (HLA-DR +) increased	[73]
PWH on ART and CD4 < 500 cells/µL	Probiotics: Lactiplantibacillus plantarum and Pediococcus acidilactici Prebiotics: Pectin, inulin, oat, acacia, maltodextrin polydextrose, and partially hydrolyzed guar gum Duration: 6 months	Randomized, double-blind, placebo con- trolled trial 71 patients • Probiotics: N= 21 • Prebiotics + Probiotics: N= 32 • Placebo: N= 18	With synbiotics, slight increases of CD4 + counts and CD4/CD8 ratio, and decreased sCD14	[74]
PWH on ART and CD4/CD8 ratio < 1	Fecal microbiota transplant from three selected donors	Randomized, double-blind, placebo con- trolled trial N = 29 patients • FMT: N = 14 • Placebo: N = 15	Decreased in IFABP, increased alpha diversity, and increased Lachnospiraceae and Rumi- nococcaceae abundance No effects on CD4 counts, CD4/CD8 ratio, or other markers of inflammation and bacte- rial translocation	[75]
MSM on ART	Supplemented Mediterranean diet (SMD) with extra-virgin olive oil and nuts	Randomized, open controlled trial N = 60, according to SMD • High adherence: N = 30 • Medium adherence: N = 20 • Low adherence: N = 8	Improved lipid profiles and reduced immune [76] activation with high adherence to SMD	[76]

Previous research in PWH

Rifaximin is a nonabsorbable antibiotic with established efficacy to reduce bacterial translocation and improves encephalopathy in patients with decompensated liver cirrhosis [77]. It has been considered as a possible modality to ameliorate persistent immune dysfunction in ART-suppressed PWH. However, in a trial that randomized PWH with poor immunologic recovery (CD4 counts < 350 cells/ μ L) to receive rifaximin or no therapy for 4 weeks, rifaximin elicited a minimal impact on bacterial translocation and CD8+T-cell activation [78] and no effects on the rectal microbiome composition [79].

Cotrimoxazole, an antibiotic indicated to prevent opportunistic infections in PWH with advanced immune suppression, has also been proposed as beneficial in reducing bacterial translocation and inflammation. The COSTOP trial, which evaluated the impact of discontinuing cotrimoxazole on 174 ART-suppressed PWH, found that cotrimoxazol cessation increased inflammation markers (C-reactive protein and IL-6). However, since it did not significantly alter microbial translocation markers, this raises the question of whether cotrimoxazole's impact on inflammation might be due to direct effects on prevalent infections, such as malaria and Pneumocystis *jirovecii*, in specific environments [80]. In children with HIV, maintaining cotrimoxazole has also been associated with decreased inflammatory markers but increased antibiotic resistance genes in their gut microbiota [81, 82].

Taken together, current evidence suggests that the composition of the gut microbiome influences susceptibility to SIV (and presumably HIV) acquisition but is less critical for disease progression. How the composition and function of the gut microbiome contribute to inflammation and comorbidities in PWH on ART is unclear, and further work is merited. Moreover, studies using antibiotics seem to suggest impact on the gut microbiome, while their use should be carefully thought of because of broad alterations to the microbiome and potential development of antibacterial resistance. An alternative approach would entail use of bacteriophage therapy, which has been shown to be more specific in targeting bacterial pathogens, and reducing antibacterial resistance might be considered [83].

Prebiotics and probiotics Previous research in NHPs

One of the most popular therapeutic interventions to increase the number of beneficial bacteria within the gut microbiome is the oral administration of live probiotic organisms [84]. While disparate in composition and quantity, probiotics are largely considered safe, but how they ultimately contribute to the gut microbiome and influence disease has been variably reported [85]. Secondly, providing particular nutrients to the microbiome has been considered to increase the growth of specific bacterial taxa thought to benefit the host [85]. In NHP models, probiotic administration to SIV-uninfected animals led to increased levels of T follicular helper cells and increased levels of IgA + B cells [86]. However, while the composition of the gut microbiome appears to influence vaccine-induced immunity [87], administration of probiotics before vaccination with HIV antigens did not result in improved vaccine-induced immunity [88].

In SIV-infected animals treated with ART, adjuvant therapy with probiotics increased the reconstitution of gut-resident CD4+T cells [63], and if IL-21 cytokine therapy was also included, there were transient increases in the levels of IL-17-producing lymphocytes within gut tissues [64]. Indeed, the levels of Lactobacillus within the guts of SIV-infected animals were related to the frequencies of Th17 cells [89]. Finally, treating chronically SIV-infected animals with *Lactobacillus*-containing probiotics led to increased gut epithelial cell repair [90]. Importantly, although these therapies were hypothesized to exert their effects by modulating the gut microbiome, there were no consistent, observable alterations. Irrespective, all studies found some beneficial influence of probiotic administration on the immunological competency of SIV-infected animals. Therefore, interventional studies in humans were subsequently considered.

Previous research in PWH

HIV is a chronic inflammatory disease, where ongoing mucosal and systemic immune dysfunction, potentially affected by impairment of the intestinal barrier [91, 92] and the microbiome [24, 26, 52, 93], leads to persistent inflammation, thereby resulting in inadequate CD4+T-cell recovery [94] and increasing the risk of mortality [95, 96]. For many years, efforts have been made to tackle the intricate defects in GALT linked to HIV, with the objective of reducing the long-term mortality risk associated with chronic inflammation using a range of interventions.

Numerous studies have explored the impact of prebiotics and probiotics in PWH on ART (Table 1). The first studies hinted at the potential benefit of these interventions to improve immune recovery. For example, in a controlled study in PWH on ART, the probiotic *Saccharomyces boulardii* decreased lipopolysaccharide-binding protein (LBP), a marker of bacterial translocation [70]. In another exploratory study, following a 6-week dietary supplementation with of prebiotics in PWH, we found modest reductions in bacterial translocation markers and T-cell activation markers, along with an improvement in thymic output and alterations in butyrate production [97]. While promising, most of these studies were exploratory, with limited sample sizes, assessed multiple outcomes, reporting effects only on some of them, and there were mixed results across studies [49, 69–74] (Table 1).

The two main controlled studies designed to assess the impact of probiotics or synbiotics on PWH on ART reported no differences in their primary outcomes [49, 98]. The PROMALTIA study explored the effects of nutritional intervention on 78 PWH starting ART at advanced stages, randomized to receive either PMT25341 (a mix of prebiotics, Saccharomyces boulardii, omega-3/6 fatty acids, and amino acids) or a placebo. The study found that the synbiotic mixture did not significantly impact CD4+T-cell counts, CD4/CD8 ratios, or gut microbiota structure compared to the placebo. Ultimately, the study concluded that while ART alone provides clear immunological benefits in advanced HIV disease, synbiotics did not enhance these benefits [49]. While more notable effects on gut microbiota composition were found in vertically infected children and adolescents receiving PMT25341 [98], again, the intervention exerted no quantifiable influence on inflammation, microbial translocation, or mucosal integrity [99].

The A5350 phase II study examined the safety, tolerability, and effects on inflammation and intestinal barrier function of Visbiome Extra Strength (ES) on inflammation and intestinal barrier function in 73 participants on ART with CD4+T-cell count>200 cells/ μ L over 24 weeks. Again, despite some changes in the microbiome, including a decrease in Gammaproteobacteria, it showed no significant impact on systemic inflammatory markers [72]. Lastly, PROOV-IT, another exploratorycontrolled study in PWH with less than 350 CD4+T cells/mm³ despite viral suppression, found a consistent lack of effect using the same probiotic [73].

In conclusion, so far, the gut microbiota remains an elusive target for prebiotic and probiotic interventions in PWH. While some potential beneficial effects are described across studies, current evidence does not support using the current formulations of prebiotics or probiotics to decrease inflammation or enhance immune recovery in PWH. In addition, the absence of regulatory oversight from major bodies, such as the Food and Drug Administration (FDA) and the European Medicines Agency (EMA), poses further challenges in their investigation as therapeutic agents. Future research focusing on second-generation probiotics, which are characterized by their diversity, specific targeting, derivation from human microbiomes, and design for distinct health outcomes, informed by a comprehensive understanding of the microbiome's role in human physiology, holds promise in advancing this field.

Fecal transplants Previous research in NHPs

As noted, probiotics have not been shown to significantly alter the composition of the gut microbiome, and they do not colonize for long periods of time [84]. An alternative approach to introducing very high numbers and species of bacterial taxa into the gut is fecal microbiota transplantation (FMT) [100]. FMT in patients suffering from Clostridioides difficile disease significantly reduces C. difficile levels, leading to clinical improvement [101]. Therefore, FMT has been considered a potential therapeutic intervention for progressive HIV/ SIV infections. In SIV-infected NHPs, animals were treated with antibiotics before fecal transplantation. Antibiotic use altered the gut microbiome to a greater extent than FMT [65]. However, even though significant bacterial dysbiosis does not occur in progressive SIV infection of Asian macaques, FMT led to increased frequencies of IL-17 and IL-22-producing lymphocytes and reduced CD4 T-cell activation within the guts of SIV-infected animals.

Previous research in PWH

Given the significant dysbiosis associated with PWH and the high safety profile associated with the treatment, there has been more interest in conducting studies in PWH than in NHPs to mitigate chronic inflammation. The first study in PWH on ART explored the effect of a single FMT delivered by colonoscopy in six participants. While microbial engraftment was measurable in this study, not all subjects experienced engraftment, which was more pronounced among the individuals with lower alpha diversity at baseline, and the effect tended to disappear during the 24-week follow-up [102]. In another uncontrolled study, six patients on ART received six weekly doses of lyophilized fecal microbiota from healthy donors, proving to be a safe and well-tolerated intervention. While microbiota diversity increased in participants with initially low diversity, the metagenomic analysis revealed no consistent changes, suggesting future trials should focus on PWH with more significant inflammation, gut damage, or dysbiosis [103].

In our pilot placebo-controlled study involving 30 PWH on ART, we asked whether repeated oral FMT could attenuate HIV-associated microbiota changes and systemic inflammation [75]. Because a hallmark of the HIV-associated microbiota is a depletion of butyrate-producing bacteria (Lachnospiraceae and Ruminococ-caceae families) [31], we specifically searched for donors with enrichment for butyrate in their feces. Weekly FMT was safe and led to increased microbiota diversity and temporary engraftment of donor microbiota, with

greater engraftment observed in those with recent antibiotic use. Additionally, FMT showed potential in reducing intestinal damage, as indicated by lower intestinal fatty acid-binding protein (IFABP), a biomarker predicting mortality in PWH on ART for long-term ART [96]. Collectively, studies in PWH indicate that FMT can lead to some changes in gut microbiota and encourage further exploration of strategies to address immune dysfunction by targeting the microbiota.

Postbiotics and live biotherapeutic products (LBPs)

If we can conclusively show that alterations in the microbiota lead to tangible clinical or immunological benefits, the subsequent step would naturally involve identifying the specific microbiota components responsible for these effects. This knowledge would pave the way for creating targeted products, like specific bacterial consortia or the emerging field of postbiotics. A prime example of progress in this area is the treatment of *C. difficile*. Following the proven effectiveness of microbiota transplants, live biotherapeutic products (LBPs)—live microorganisms to modulate the microbiome—have proved efficacy and have gained approval to become commercially available [104, 105]. There are immense research opportunities in HIV.

For example, short-chain fatty acids (SCFAs) produced by certain gut bacteria exhibit anti-inflammatory properties [106]. A postbiotic intervention that might be hypothesized to decrease intestinal permeability and reduce microbial translocation-induced inflammation is butyrate [68]. As noted, this short-chain fatty acid is an important microbiome-produced metabolite that provides energy to the epithelium. However, high doses of butyrate in ARV-treated, SIV-infected, NHPs were insufficient to reduce residual inflammation in these animals. Indeed, very few immunological benefits were noted after butyrate supplementation, possibly due to butyrate absorption within the small intestine. A less absorbed, but bioactive compound such as tributyrin or probiotic organisms which produce high levels of butyrate may provide clinical benefits [107].

PWH display lower levels of butyrate-producing bacteria, such as *Roseburia intestinalis*, which correlates with increased systemic immune activation and HIV replication [97, 108]. Adding butyrate exogenously in vitro reduced T-cell activation and HIV replication, suggesting that butyrate modulates gut immune responses in HIV [108]. We previously observed fecal butyrate increases following prebiotic dietary supplementation, which correlated with amelioration of the inflammatory biomarkers soluble CD14 and highsensitivity C-reactive protein [97]. However, in a randomized controlled trial investigating the effects of vitamin D and phenylbutyrate supplementation — a butyrate derivative — on gut-derived inflammation and microbiota composition PWH, no effects on gut-derived inflammatory markers or the microbiota in treatment-naïve PWH were found [109].

Therefore, despite the links between butyrate and microbial translocation-induced inflammation shown in studies on SIV-infected NHP and PWH, the role of butyrate as a postbiotic in PWH remains to be proven. Further research in the postbiotis field is needed.

Dietary interventions

Previous research in SIVs

In addition to oral use of probiotics, other medicinal food and other dietary interventions have been considered for potential alteration of the gut microbiome and physiology. Diets high in fat and sugars alter the composition of the gut microbiome and increase gut inflammation [110]. However, while a high-fat diet significantly increased inflammation in NHPs, only modest alteration to the gut microbiome was observed [66, 67]. Importantly, inflammation associated with the high-fat diet dramatically accelerated progressive SIV infection of NHPs, suggesting that novel therapeutic interventions aimed at reducing obesity-associated inflammation may improve the prognosis of PLWH.

Previous research in PWH

In PWH, dietary interventions, particularly the Mediterranean diet supplemented with nuts and olive oil, have shown promise in improving gut microbiota, reducing inflammation, and potentially impacting microbial translocation. A study examined 102 PWH over 12 weeks, comparing those on a supplemented Mediterranean diet with extra-virgin olive oil and nuts to a control group on a regular diet. After classifying individuals by their adherence to the Mediterranean diet, those highly adherent exhibited improved lipid profiles and reduced immune activation. Notably, gut microbiota diversity and richness increased in the high-adherence group, with significant correlations between Succinivibrio and Bifidobacterium abundances and Treg cell levels [76].

In an observational study, we identified two dietary patterns among PWH: Mediterranean like and Western like. Participants with a Mediterranean pattern showed higher Lachnospira abundance and lower levels of the inflammatory biomarkers D-dimer and soluble tumor necrosis factor receptor 2 (sTNFR2), two independent predictors of mortality [96], compared to the Westerndiet fed group. The Mediterranean pattern was also associated with lower Erysipelotrichaceae abundance and inflammatory markers, indicating its potential to reduce inflammation-related processes in PWH [111]. In summary, despite the limited availability of clinical trials evaluating the effects of dietary interventions on gut microbiota and inflammation in PWH, existing data suggest that healthier dietary choices can positively alter the microbiota and diminish inflammation in PWH. This aligns with findings from extensive research conducted in the general population [112].

Other interventions

Beyond the above interventions, there are other gutoriented strategies to reduce inflammation in PWH. However, most studies have yielded negative results. For example, mesalamine, an anti-inflammatory agent effective in ulcerative colitis, was tested in a placebocontrolled trial to assess its impact on abnormal immune activation in PWH. The study, involving 33 PWH on ART with incomplete immune recovery (CD4 < 350 cells/mm³), showed no significant change in various immune markers, including canonical markers of T-cell activation (CD38 + HLA-DR + CD4 + and CD8 + T cells) in blood and rectal tissue, or in various soluble markers of inflammation, after 12 weeks of mesalamine treatment [113].

Table 2 Challenges and solutions for limitations in SIV/HIV microbiota research

Research barrier	Specific challenge	Proposed solution
Complexity of microbial interactions	Difficulty in understanding interactions, causal- ity vs. correlation issues	Enhanced multidisciplinary studies, advanced computational modeling coupled with experi- mental validations
Incomplete definition of a healthy microbiota	Lack of consensus on "healthy" microbiome, vari- ability across populations	Standardized guidelines for microbiota research, global collaborative research
Individual variability in microbiome	Inter-individual differences, standardization issues	Personalized medicine approaches, large-scale population studies
Confounding effects of sexual practices	Isolating effects of HIV beyond confounders	Targeted research on sexual behavior impacts, controlled study designs
Confounding effects of geographical and die- tary factors	Regional microbiome variation, generalization challenges	Cross-cultural studies, inclusion of diverse dietary patterns in research
Interactions with ART	ART-microbiome interplay, regimen variability	Comprehensive ART-microbiome interaction stud- ies, standardized ART protocols
Limited longitudinal data	Scarcity of long-term studies, safety and efficacy assessments	Longitudinal study designs, posttreatment follow- up studies
Ethical and clinical trial constraints to study microbial interactions in tissue	Ethical concerns with invasive studies, clinical trial complexities	Develop innovative noninvasive techniques to sample tissues
Lack of standardized methodologies	Methodological differences, standardization need in proteomics and metabolomics	Development of universal protocols, collaborative methodological research
Technological and analytical challenges	Limitations in technology to understand the microbiota, data analysis complexities	Investment in advanced analytical tools, collabora- tion with data scientists
Generalizability of findings	Difficulty in applying findings broadly, microbi- ome composition variations	Multicentric studies, inclusion of diverse partici- pant profiles
Interdisciplinary integration	Need for integrated research, coordination challenges	Interdisciplinary research programs, collaborative platforms
Funding and resource allocation	Limited funding, resource prioritization	Advocacy for increased funding, diversification of funding sources

Table 3 Challenges and solutions for limitations in current intervention studies in SIV/HIV microbiome research
--

Limitation category	Specific limitation	Proposed solution
Prebiotics and probiotics market proliferation	Marketed without research evidence, efficacy and safety variation	Stricter regulations, evidence-based marketing
Heterogeneity in intervention studies	Dosage and duration heterogeneity, outcome inconsistency	Standardized intervention protocols, unified out- come measures
FMT challenges	Donor profile uncertainty, antibiotic precondition- ing consensus	Donor profile databases, clinical trials assessing preconditioning antibiotic regimens
Effectiveness assessment and localization	Stool vs. tissue microbiota correlation, impact assessment challenges	Advanced diagnostic tools, in-depth impact studies
Small sample sizes and short study duration	Limited sample size, short study duration	Larger, multicenter trials, extended study periods
Study design	Lack of standardized protocols, study design variability	Protocol development committees for microbiota interventions

Challenges and limitations

In the field of SIV/HIV microbiome research, we are confronted with numerous complexities. Central to these challenges is the complex interplay between the microbiome and the immune system, especially in the context of HIV. How variable is the microbiome across and within individuals? What are the fundamental characteristics of a "healthy" microbiome, and how do these vary across different populations influenced by genetics, lifestyle, and environmental factors [114]? How can we diagnose "dysbiosis"?

One significant obstacle is the individual variability in microbiome composition. This not only raises questions about the standardization of study conditions but also about the generalizability of research findings [115]. The influence of external factors like sexual practices, geographical location, and dietary habits adds further complexity. How do these factors interplay with HIV to affect the microbiome?

The scarcity of longitudinal studies poses another hurdle. What are the long-term risks of modifying the microbiome, and do the effects of interventions last over time? Ethical considerations, especially concerning invasive procedures for detailed microbiome analysis, further complicate these efforts. Moreover, interactions between ART and the gut microbiome modulations, together with the variability in ART regimens, ART length, and immune recovery, add another layer of complexity. Technological and analytical limitations, particularly in handling highly multidimensional data, are especially relevant in proteomics and metabolomics studies, essential for understanding the microbiome at a functional level.

Intervention studies in the SIV/HIV microbiome area face distinct challenges. The market is flooded with prebiotics and probiotics, often lacking sufficient research-backed evidence. How can we ensure the pharmacological properties of these products? There is also significant heterogeneity in dosage, duration, and measured outcomes across studies. What standardized outcome definitions and designs can we implement to ensure reliable evaluations and follow-ups?

FMT presents its own set of uncertainties. Which donor microbiota profiles are optimal for FMT in PWH? Do FMT donor profiles need to be individualized? Is an antibiotic conditioning regimen necessary for a more impactful microbiota modification? The lack of consensus on these issues points to the need for a more standardized and coordinated approach.

In summary, addressing these challenges calls for an interdisciplinary strategy, integrating advancements in technology and analytical methodologies. The list of challenges and potential solutions can be found in Tables 2 and 3, highlighting the need for collaborative

efforts to realize the potential of microbiome research in HIV management.

Future directions

Future directions in SIV/HIV microbiome research must focus on unexplored domains while refining our understanding of established concepts. The role of the microbiome in HIV transmission and its influence on comorbidities is fertile ground for research.

A granular understanding of microbiome-immune interactions is essential. Current knowledge is rudimentary, and future research should detail these interactions to design more effective interventions. This effort demands a meticulous examination of the molecular dialogues between microbial communities and immune responses. Such insights are vital for developing targeted therapeutic strategies, particularly in modulating immune responses and controlling inflammation.

Personalized microbiome-based therapies represent another promising frontier. The inter-individual variability of the microbiome suggests that tailored therapeutic approaches could be more effective than one-size-fits-all solutions. Research must pivot towards understanding individual microbiome profiles and their interactions with the immune system, paving the way for customized treatment protocols. For example, if we are able to identify detrimental bacteria, we should design personalized strategies to extract them from the microbiome. Exploring the potential of bacteriophage therapy to selectively target detrimental bacteria within the microbiota of PWH offers a promising avenue. This approach, if harnessed effectively, could minimize the risk of inducing antibiotic resistance genes, compared to traditional antibiotics.

Defining outcomes is another crucial area. Precise parameters for inflammation, immune recovery in immunological nonresponders, HIV transmission, and the management of comorbidities need establishment. These outcomes should not only guide research but also inform clinical practice, aiding in developing more effective treatment strategies.

Technological advancements are pivotal in transitioning beyond current interventions like FMT and pre-/ postbiotics. The search for potential therapeutic agents from the microbiome demands not just sophisticated technologies but also a focus on exploring its "dark matter." This includes the extensive virome and genome of uncultivated taxa, which are functionally significant yet remain largely unknown [116]. Investment in advanced analytical tools, high-throughput sequencing, and bioinformatics is essential. Collaboration with data scientists and bioinformaticians, followed by experimental validation of the detected hits, will facilitate handling and interpreting the complex data sets generated by these studies.

Conclusions

Multiple recent studies have highlighted the complex interplay between the microbiome and SIV/HIV, emphasizing its significant implications for disease progression and treatment. We have gained substantial insights into how disruptions in the microbiome contribute to bacterial translocation and inflammation, paving the way for innovative therapeutic interventions.

However, this field faces notable challenges. The diversity in microbiome composition among individuals, shaped by genetics, lifestyle, and environment, complicates the development of universal treatment protocols. Confounding variables, such as diet, sexual practices, immune status, and ART, add layers of complexity, necessitating a subtle methodological approach. The requirement for long-term studies to understand the enduring impacts of microbiome alterations is clear, as is the ethical responsibility in conducting such research.

Despite these obstacles, the potential of microbiomebased therapies in HIV treatment remains promising. Exploring prebiotics, probiotics, and FMT has introduced new possibilities, though their efficacy and safety continue to be actively researched. The emergence of postbiotics and LBPs represents a new frontier in leveraging the microbiome to achieve better health outcomes in PWH. Dietary interventions have also shown promise in altering the microbiome and reducing inflammation, offering a less invasive method to modulate the microbiome. The efficacy of these interventions, along with other innovative strategies like the use of GLP-2 agonists, highlights the value of a multidisciplinary approach in addressing HIV's complexities.

Looking ahead, the future of this field depends on integrating advanced technological and analytical methods with collaborative, interdisciplinary efforts. Such a comprehensive approach is essential to unlocking the full potential of microbiome research in HIV treatment. As we advance, awareness of the current challenges and limitations highlighted in this review will lay a more robust foundation for future studies, steering us towards more effective and personalized HIV treatments (Fig. 1).

Acknowledgements

Figure 1 was created with BioRender.com.

Authors' contributions

JB and SSV equally contributed to the conceptualization, literature review, writing of the manuscript, and to table and figures preparation. Both authors reviewed the manuscript.

Funding

Open access funding provided by the National Institutes of Health. Funding for this study was provided in part by the Division of Intramural Research/ NIAID/NIH of the Instituto de Salud Carlos III and Fondos FEDER, Acción Estratégica en Salud (ICI20/00058 and PI21/00041). The content of this publication does not necessarily reflect the views or policies of DHHS, nor does the mention of trade names, commercial products, or organizations imply endorsement by the US government.

Availability of data and materials

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

S.S.V has received fees for educational presentations from Mikrobiomik. The authors declare no other competing financial interests.

Received: 23 December 2023 Accepted: 26 April 2024 Published: 21 June 2024

References

- Hsue PY, Waters DD. HIV infection and coronary heart disease: mechanisms and management. Nat Rev Cardiol. 2019. https://doi.org/10. 1038/s41569-019-0219-9.
- Schnittman SR, Hunt PW. Clinical consequences of asymptomatic Cytomegalovirus in treated human immunodeficency virus infection. Curr Opin HIV AIDS. 2021;16:168–76.
- 3. Swiecki M, Colonna M. The multifaceted biology of plasmacytoid dendritic cells. Nat Rev Immunol. 2015;15:471–85.
- Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, Rao S, Kazzaz Z, Bornstein E, Lambotte O, Altmann D, Blazar BR, Rodriguez B, Teixeira-Johnson L, Landay A, Martin JN, Hecht FM, Picker LJ, Lederman MM, Deeks SG, Douek DC. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. Nat Med. 2006;12:1365–71.
- Shanahan F. The host-microbe interface within the gut. Best Pract Res Clin Gastroenterol. 2002;16:915–31.
- O'Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Rep. 2006;7:688–93.
- Louis P, Young P, Holtrop G, Flint HJ. Diversity of human colonic butyrate-producing bacteria revealed by analysis of the butyryl-CoA:acetate CoA-transferase gene. Environ Microbiol. 2010;12:304–14.
- Brandtzaeg P, Baekkevold ES, Farstad IN, Jahnsen FL, Johansen FE, Nilsen EM, Yamanaka T. Regional specialization in the mucosal immune system: what happens in the microcompartments? Immunol Today. 1999;20:141–51.
- Kotler DP, Gaetz HP, Lange M, Klein EB, Holt PR. Enteropathy associated with the acquired immunodeficiency syndrome. Ann Intern Med. 1984;101:421–8.
- Rodgers VD, Fassett R, Kagnoff MF. Abnormalities in intestinal mucosal T cells in homosexual populations including those with the lymphadenopathy syndrome and acquired immunodeficiency syndrome. Gastroenterology. 1986;90:552–8.
- Clayton F, Reka S, Cronin WJ, Torlakovic E, Sigal SH, Kotler DP. Rectal mucosal pathology varies with human immunodeficiency virus antigen content and disease stage. Gastroenterology. 1992;103:919–33.
- 12. Ullrich R, Zeitz M, Riecken EO. Enteric immunologic abnormalities in human immunodeficiency virus infection. Semin Liver Dis. 1992;12:167–74.

- Kotler DP, Reka S, Clayton F. Intestinal mucosal inflammation associated with human immunodeficiency virus infection. Dig Dis Sci. 1993;38:1119–27.
- Clayton F, Snow G, Reka S, Kotler DP. Selective depletion of rectal lamina propria rather than lymphoid aggregate CD4 lymphocytes in HIV infection. Clin Exp Immunol. 1997;107:288–92.
- Mattapallil JJ, Douek DC, Hill B, Nishimura Y, Martin M, Roederer M. Massive infection and loss of memory CD4+ T cells in multiple tissues during acute SIV infection. Nature. 2005;434:1093–7.
- Li Q, Duan L, Estes JD, Ma ZM, Rourke T, Wang Y, Reilly C, Carlis J, Miller CJ, Haase AT. Peak SIV replication in resting memory CD4+ T cells depletes gut lamina propria CD4+ T cells. Nature. 2005;434:1148–52.
- Brenchley JM, Paiardini M, Knox KS, Asher AI, Cervasi B, Asher TE, Scheinberg P, Price DA, Hage CA, Kholi LM, Khoruts A, Frank I, Else J, Schacker T, Silvestri G, Douek DC. Differential Th17 CD4 T-cell depletion in pathogenic and nonpathogenic lentiviral infections. Blood. 2008;112:2826–35.
- Cecchinato V, Trindade C, Laurence A, Heraud J, Brenchley J, Ferrari M, Zaffiri L, Tryniszewska E, Tsai W, Vaccari M, Washington Parks R, Venzon D, Douek D, O'Shea J, Franchini G. Altered balance between Th17 and Th1 cells at mucosal sites predicts AIDS progression in simian immunodeficienty virus-infected macaques. Mucosal Immunol. 2008;1:279–88.
- Favre D, Lederer S, Kanwar B, Ma ZM, Proll S, Kasakow Z, Mold J, Swainson L, Barbour JD, Baskin CR, Palermo R, Pandrea I, Miller CJ, Katze MG, McCune JM. Critical loss of the balance between Th17 and T regulatory cell populations in pathogenic SIV infection. Plos Pathog. 2009;5:e1000295.
- Gordon SN, Cervasi B, Odorizzi P, Silverman R, Aberra F, Ginsberg G, Estes JD, Paiardini M, Frank I, Silvestri G. Disruption of intestinal CD4+T cell homeostasis is a key marker of systemic CD4+T cell activation in HIV-infected individuals. J Immunol. 2010;185:5169–79.
- 21. Klatt NR, Brenchley JM. Th17 cell dynamics in HIV infection. Curr Opin HIV AIDS. 2010;5:135–40.
- 22. Chege D, Sheth PM, Kain T, Kim CJ, Kovacs C, Loutfy M, Halpenny R, Kandel G, Chun TW, Ostrowski M, Kaul R. Sigmoid Th17 populations, the HIV latent reservoir, and microbial translocation in men on long-term antiretroviral therapy. AIDS. 2011;25:741–9.
- 23. Hunt PW. Th17, gut, and HIV: therapeutic implications. Curr Opin HIV AIDS. 2010;5:189–93.
- Vujkovic-Cvijin I, Dunham RM, Iwai S, Maher MC, Albright RG, Broadhurst MJ, Hernandez RD, Lederman MM, Huang Y, Somsouk M, Deeks SG, Hunt PW, Lynch SV, McCune JM. Dysbiosis of the gut microbiota is associated with HIV disease progression and tryptophan catabolism. Sci Transl Med. 2013;5:193ra91.
- Lozupone CA, Li M, Campbell TB, Flores SC, Linderman D, Gebert MJ, Knight R, Fontenot AP, Palmer BE. Alterations in the gut microbiota associated with HIV-1 infection. Cell Host Microbe. 2013;14:329–39.
- Dillon SM, Lee EJ, Kotter CV, Austin GL, Dong Z, Hecht DK, Gianella S, Siewe B, Smith DM, Landay AL, Robertson CE, Frank DN, Wilson CC. An altered intestinal mucosal microbiome in HIV-1 infection is associated with mucosal and systemic immune activation and endotoxemia. Mucosal Immunol. 2014;7:983–94.
- Ellis CL, Ma ZM, Mann SK, Li CS, Wu J, Knight TH, Yotter T, Hayes TL, Maniar AH, Troia-Cancio PV, Overman HA, Torok NJ, Albanese A, Rutledge JC, Miller CJ, Pollard RB, Asmuth DM. Molecular characterization of stool microbiota in HIV-infected subjects by panbacterial and order-level 16S ribosomal DNA (rDNA) quantification and correlations with immune activation. J Acquir Immune Defic Syndr. 2011;57:363–70.
- Guo X, Wang Z, Qu M, Guo Y, Yu M, Hong W, Zhang C, Fan X, Song J, Xu R, Zhang J, Huang H, Linghu E, Wang FS, Sun L, Jiao YM. Abnormal blood microbiota profiles are associated with inflammation and immune restoration in HIV/AIDS individuals. mSystems. 2023;8:e0046723.
- Rocafort M, Noguera-Julian M, Rivera J, Pastor L, Guillen Y, Langhorst J, Parera M, Mandomando I, Carrillo J, Urrea V, Rodriguez C, Casadella M, Calle ML, Clotet B, Blanco J, Naniche D, Paredes R. Evolution of the gut microbiome following acute HIV-1 infection. Microbiome. 2019;7:73.
- Vazquez-Castellanos JF, Serrano-Villar S, Latorre A, Artacho A, Ferrus ML, Madrid N, Vallejo A, Sainz T, Martinez-Botas J, Ferrando-Martinez S, Vera M, Dronda F, Leal M, Del Romero J, Moreno S, Estrada V, Gosalbes MJ, Moya

A. Altered metabolism of gut microbiota contributes to chronic immune activation in HIV-infected individuals. Mucosal Immunol. 2015;8:760–72.

- Vujkovic-Cvijin I, Somsouk M. HIV and the gut microbiota: composition, consequences, and avenues for amelioration. Curr HIV/AIDS Rep. 2019;16:204–13.
- Klase Z, Ortiz A, Deleage C, Mudd JC, Quinones M, Schwartzman E, Klatt NR, Canary L, Estes JD, Brenchley JM. Dysbiotic bacteria translocate in progressive SIV infection. Mucosal Immunol. 2015;8:1009–20.
- McKenna P, Hoffmann C, Minkah N, Aye PP, Lackner A, Liu Z, Lozupone CA, Hamady M, Knight R, Bushman FD. The macaque gut microbiome in health, lentiviral infection, and chronic enterocolitis. Plos Pathog. 2008;4:e20.
- Glavan TW, Gaulke CA, Santos Rocha C, Sankaran-Walters S, Hirao LA, Raffatellu M, Jiang G, Baumler AJ, Goulart LR, Dandekar S. Gut immune dysfunction through impaired innate pattern recognition receptor expression and gut microbiota dysbiosis in chronic SIV infection. Mucosal Immunol. 2016;9:677–88.
- Handley SA, Desai C, Zhao G, Droit L, Monaco CL, Schroeder AC, Nkolola JP, Norman ME, Miller AD, Wang D, Barouch DH, Virgin HW. SIV infectionmediated changes in gastrointestinal bacterial microbiome and virome are associated with immunodeficiency and prevented by vaccination. Cell Host Microbe. 2016;19:323–35.
- 36. Ericsen AJ, Lauck M, Mohns MS, DiNapoli SR, Mutschler JP, Greene JM, Weinfurter JT, Lehrer-Brey G, Prall TM, Gieger SM, Buechler CR, Crosno KA, Peterson EJ, Reynolds MR, Wiseman RW, Burwitz BJ, Estes JD, Sacha JB, Friedrich TC, Brenchley JM, O'Connor DH. Microbial translocation and inflammation occur in hyperacute immunodeficiency virus infection and compromise host control of virus replication. Plos Pathog. 2016;12:e1006048.
- Guillen Y, Noguera-Julian M, Rivera J, Casadella M, Zevin AS, Rocafort M, Parera M, Rodriguez C, Arumi M, Carrillo J, Mothe B, Estany C, Coll J, Bravo I, Herrero C, Saz J, Sirera G, Torrella A, Navarro J, Crespo M, Negredo E, Brander C, Blanco J, Calle ML, Klatt NR, Clotet B, Paredes R. Low nadir CD4+ T-cell counts predict gut dysbiosis in HIV-1 infection. Mucosal Immunol. 2019;12:232–46.
- Noguera-Julian M, Rocafort M, Guillen Y, Rivera J, Casadella M, Nowak P, Hildebrand F, Zeller G, Parera M, Bellido R, Rodriguez C, Carrillo J, Mothe B, Coll J, Bravo I, Estany C, Herrero C, Saz J, Sirera G, Torrela A, Navarro J, Crespo M, Brander C, Negredo E, Blanco J, Guarner F, Calle ML, Bork P, Sonnerborg A, Clotet B, Paredes R. Gut microbiota linked to sexual preference and HIV infection. EBioMedicine. 2016;5:135–46.
- Vujkovic-Cvijin I, Sortino O, Verheij E, Sklar J, Wit FW, Kootstra NA, Sellers B, Brenchley JM, Ananworanich J, Loeff MSV, Belkaid Y, Reiss P, Sereti I. HIV-associated gut dysbiosis is independent of sexual practice and correlates with noncommunicable diseases. Nat Commun. 2020;11:2448.
- 40. Dickson I. Gut microbiota: diagnosing IBD with the gut microbiome. Nat Rev Gastroenterol Hepatol. 2017;14:195.
- 41. Dowling LR, Strazzari MR, Keely S, Kaiko GE. Enteric nervous system and intestinal epithelial regulation of the gut-brain axis. J Allergy Clin Immunol. 2022;150:513–22.
- 42. Vujkovic-Cvijin I, Sklar J, Jiang L, Natarajan L, Knight R, Belkaid Y. Host variables confound gut microbiota studies of human disease. Nature. 2020;587:448–54.
- 43. von Mollendorf C, von Gottberg A, Tempia S, Meiring S, de Gouveia L, Quan V, Lengana S, Avenant T, du Plessis N, Eley B, Finlayson H, Reubenson G, Moshe M, O'Brien KL, Klugman KP, Whitney CG, Cohen C, Group for Enteric R, Meningeal Disease Surveillance in South A. Increased risk for and mortality from invasive pneumococcal disease in HIV-exposed but uninfected infants aged <1 year in South Africa, 2009–2013. Clin Infect Dis. 2015;60:1346–56.
- Bender JM, Li F, Martelly S, Byrt E, Rouzier V, Leo M, Tobin N, Pannaraj PS, Adisetiyo H, Rollie A, Santiskulvong C, Wang S, Autran C, Bode L, Fitzgerald D, Kuhn L, Aldrovandi GM. Maternal HIV infection influences the microbiome of HIV-uninfected infants. Sci Transl Med. 2016;8:349ra100.
- 45. Machiavelli A, Duarte RTD, Pires MMS, Zarate-Blades CR, Pinto AR. The impact of in utero HIV exposure on gut microbiota, inflammation, and microbial translocation. Gut Microbes. 2019;10:599–614.
- Amenyogbe N, Dimitriu P, Cho P, Ruck C, Fortuno ES 3rd, Cai B, Alimenti A, Cote HCF, Maan EJ, Slogrove AL, Esser M, Marchant A, Goetghebuer T, Shannon CP, Tebbutt SJ, Kollmann TR, Mohn WW, Smolen

KK. Innate immune responses and gut microbiomes distinguish HIV-exposed from HIV-unexposed children in a population-specific manner. J Immunol. 2020;205:2618–28.

- 47. Jackson CL, Frank DN, Robertson CE, Ir D, Kofonow JM, Montlha MP, Mutsaerts E, Nunes MC, Madhi SA, Ghosh D, Weinberg A. Evolution of the gut microbiome in HIV-exposed uninfected and unexposed infants during the first year of life. mBio. 2022;13:e0122922.
- Ancona G, Merlini E, Tincati C, Barassi A, Calcagno A, Augello M, Bono V, Bai F, Cannizzo ES, d'Arminio Monforte A, Marchetti G. Long-term suppressive cART is not sufficient to restore intestinal permeability and gut microbiota compositional changes. Front Immunol. 2021;12:639291.
- 49. Serrano-Villar S, de Lagarde M, Vazquez-Castellanos J, Vallejo A, Bernadino JI, Madrid N, Matarranz M, Diaz-Santiago A, Gutierrez C, Cabello A, Villar-Garcia J, Blanco JR, Bisbal O, Sainz T, Moya A, Moreno S, Gosalbes MJ, Estrada V. Effects of immunonutrition in advanced human immunodeficiency virus disease: a randomized placebo-controlled clinical trial (Promaltia Study). Clin Infect Dis. 2019;68:120–30.
- Burkhart Colorado AS, Lazzaro A, Neff CP, Nusbacher N, Boyd K, Fiorillo S, Martin C, Siebert JC, Campbell TB, Borok M, Palmer BE, Lozupone C. Differential effects of antiretroviral treatment on immunity and gut microbiome composition in people living with HIV in rural versus urban Zimbabwe. Microbiome. 2024;12:18.
- Rubio-Garcia E, Ferrando N, Martin N, Balleste-Delpierre C, Miro JM, Paredes R, Casals-Pascual C, Vila J. In vitro antibacterial activity of antiretroviral drugs on key commensal bacteria from the human microbiota. Front Cell Infect Microbiol. 2023;13:1306430.
- Serrano-Villar S, Rojo D, Martinez-Martinez M, Deusch S, Vazquez-Castellanos JF, Bargiela R, Sainz T, Vera M, Moreno S, Estrada V, Gosalbes MJ, Latorre A, Seifert J, Barbas C, Moya A, Ferrer M. Gut bacteria metabolism impacts immune recovery in HIV-infected individuals. EBioMedicine. 2016;8:203–16.
- 53. Wang Z, Peters BA, Bryant M, Hanna DB, Schwartz T, Wang T, Sollecito CC, Usyk M, Grassi E, Wiek F, Peter LS, Post WS, Landay AL, Hodis HN, Weber KM, French A, Golub ET, Lazar J, Gustafson D, Sharma A, Anastos K, Clish CB, Burk RD, Kaplan RC, Knight R, Qi Q. Gut microbiota, circulating inflammatory markers and metabolites, and carotid artery atherosclerosis in HIV infection. Microbiome. 2023;11:119.
- Vazquez-Castellanos JF, Serrano-Villar S, Jimenez-Hernandez N, Soto Del Rio MD, Gayo S, Rojo D, Ferrer M, Barbas C, Moreno S, Estrada V, Rattei T, Latorre A, Moya A, Gosalbes MJ. Interplay between gut microbiota metabolism and inflammation in HIV infection. ISME J. 2018;12:1964–76.
- 55. Sperk M, Ambikan AT, Ray S, Singh K, Mikaeloff F, Diez RC, Narayanan A, Vesterbacka J, Nowak P, Sonnerborg A, Neogi U. Fecal metabolome signature in the HIV-1 elite control phenotype: enrichment of dipeptides acts as an HIV-1 antagonist but a Prevotella agonist. J Virol. 2021;95:e0047921.
- Vesterbacka J, Rivera J, Noyan K, Parera M, Neogi U, Calle M, Paredes R, Sonnerborg A, Noguera-Julian M, Nowak P. Richer gut microbiota with distinct metabolic profile in HIV infected elite controllers. Sci Rep. 2017;7:6269.
- 57. Bochart RM, Busman-Sahay K, Bondoc S, Morrow DW, Ortiz AM, Fennessey CM, Fischer MB, Shiel O, Swanson T, Shriver-Munsch CM, Crank HB, Armantrout KM, Barber-Axthelm AM, Langner C, Moats CR, Labriola CS, MacAllister R, Axthelm MK, Brenchley JM, Keele BF, Estes JD, Hansen SG, Smedley JV. Mitigation of endemic GI-tract pathogenmediated inflammation through development of multimodal treatment regimen and its impact on SIV acquisition in rhesus macaques. Plos Pathog. 2021;17:e1009565.
- Manuzak JA, Zevin AS, Cheu R, Richardson B, Modesitt J, Hensley-McBain T, Miller C, Gustin AT, Coronado E, Gott T, Fang M, Cartwright M, Wangari S, Agricola B, May D, Smith E, Hampel HB, Gale M, Cameron CM, Cameron MJ, Smedley J, Klatt NR. Antibiotic-induced microbiome perturbations are associated with significant alterations to colonic mucosal immunity in rhesus macaques. Mucosal Immunol. 2020;13:471–80.
- Flynn JK, Ortiz AM, Herbert R, Brenchley JM. Host genetics and environment shape the composition of the gastrointestinal microbiome in nonhuman primates. Microbiol Spectr. 2023;11:e0213922.

- Ortiz AM, Flynn JK, DiNapoli SR, Vujkovic-Cvijin I, Starke CE, Lai SH, Long ME, Sortino O, Vinton CL, Mudd JC, Johnston L, Busman-Sahay K, Belkaid Y, Estes JD, Brenchley JM. Experimental microbial dysbiosis does not promote disease progression in SIV-infected macaques. Nat Med. 2018;24:1313–6.
- Ortiz AM, Baker PJ, Langner CA, Simpson J, Stacy A, Flynn JK, Starke CE, Vinton CL, Fennessey CM, Belkaid Y, Keele BF, Brenchley JM. Experimental bacterial dysbiosis with consequent immune alterations increase intrarectal SIV acquisition susceptibility. Cell Rep. 2023;42:112020.
- Pandrea I, Xu C, Stock JL, Frank DN, Ma D, Policicchio BB, He T, Kristoff J, Cornell E, Haret-Richter GS, Trichel A, Ribeiro RM, Tracy R, Wilson C, Landay AL, Apetrei C. Antibiotic and antiinflammatory therapy transiently reduces inflammation and hypercoagulation in acutely SIV-infected pigtailed macaques. Plos Pathog. 2016;12:e1005384.
- 63. Klatt NR, Canary LA, Sun X, Vinton CL, Funderburg NT, Morcock DR, Quinones M, Deming CB, Perkins M, Hazuda DJ, Miller MD, Lederman MM, Segre JA, Lifson JD, Haddad EK, Estes JD, Brenchley JM. Probiotic/ prebiotic supplementation of antiretrovirals improves gastrointestinal immunity in SIV-infected macaques. J Clin Invest. 2013;123:903–7.
- Ortiz AM, Klase ZA, DiNapoli SR, Vujkovic-Cvijin I, Carmack K, Perkins MR, Calantone N, Vinton CL, Riddick NE, Gallagher J, Klatt NR, McCune JM, Estes JD, Paiardini M, Brenchley JM. 2015. IL-21 and probiotic therapy improve Th17 frequencies, microbial translocation, and microbiome in ARV-treated, SIV-infected macaques. Mucosal Immunol 2015. https:// doi.org/10.1038/mi.2015.75.
- Hensley-McBain T, Zevin AS, Manuzak J, Smith E, Gile J, Miller C, Agricola B, Katze M, Reeves RK, Kraft CS, Langevin S, Klatt NR. Effects of fecal microbial transplantation on microbiome and immunity in Simian immunodeficiency virus-infected macaques. J Virol. 2016;90:4981–9.
- Mansfield KG, Carville A, Wachtman L, Goldin BR, Yearley J, Li W, Woods M, Gualtieri L, Shannon R, Wanke C. A diet high in saturated fat and cholesterol accelerates Simian immunodeficiency virus disease progression. J Infect Dis. 2007;196:1202–10.
- 67. He T, Xu C, Krampe N, Dillon SM, Sette P, Falwell E, Haret-Richter GS, Butterfield T, Dunsmore TL, McFadden WM Jr, Martin KJ, Policicchio BB, Raehtz KD, Penn EP, Tracy RP, Ribeiro RM, Frank DN, Wilson CC, Landay AL, Apetrei C, Pandrea I. High-fat diet exacerbates SIV pathogenesis and accelerates disease progression. J Clin Invest. 2019;129:5474–88.
- Ortiz AM, Simpson J, Langner CA, Baker PJ, Aguilar C, Brooks K, Flynn JK, Vinton CL, Rahmberg AR, Hickman HD, Brenchley JM. Butyrate administration is not sufficient to improve immune reconstitution in antiretroviral-treated SIV-infected macaques. Sci Rep. 2022;12:7491.
- 69. Stiksrud B, Nowak P, Nwosu FC, Kvale D, Thalme A, Sonnerborg A, Ueland PM, Holm K, Birkeland SE, Dahm AE, Sandset PM, Rudi K, Hov JR, Dyrhol-Riise AM, Troseid M. Reduced levels of D-dimer and changes in gut microbiota composition after probiotic intervention in HIV-infected individuals on stable ART. J Acquir Immune Defic Syndr. 2015;70:329–37.
- Villar-Garcia J, Hernandez JJ, Guerri-Fernandez R, Gonzalez A, Lerma E, Guelar A, Saenz D, Sorli L, Montero M, Horcajada JP, Knobel FH. Effect of probiotics (Saccharomyces boulardii) on microbial translocation and inflammation in HIV-treated patients: a double-blind, randomized, placebo-controlled trial. J Acquir Immune Defic Syndr. 2015;68:256–63.
- Tenore SB, Avelino-Silva VI, Costa PR, Franco LM, Sabino EC, Kalil J, Cerqueira NB, Nakagawa Z, Kallas EG. Immune effects of Lactobacillus casei Shirota in treated HIV-infected patients with poor CD4+T-cell recovery. AIDS. 2020;34:381–9.
- Presti RM, Yeh E, Williams B, Landay A, Jacobson JM, Wilson C, Fichtenbaum CJ, Utay NS, Dube MP, Klingman KL, Estes JD, Flynn JK, Loftin A, Brenchley JM, Andrade A, Kitch DW, Overton ET. A randomized, placebo-controlled trial assessing the effect of VISBIOME ES probiotic in people with HIV on antiretroviral therapy. Open Forum Infect Dis. 2021;8:ofab550.
- Rousseau RK, Walmsley SL, Lee T, Rosenes R, Reinhard RJ, Malazogu F, Benko E, Huibner S, Kovacs CM, Singer J, Kim CJ, Kaul R. Randomized, blinded, placebo-controlled trial of De Simone formulation probiotic during HIV-associated suboptimal CD4+ T cell recovery. J Acquir Immune Defic Syndr. 2022;89:199–207.
- 74. Blazquez-Bondia C, Parera M, Catala-Moll F, Casadella M, Elizalde-Torrent A, Aguilo M, Espadaler-Mazo J, Santos JR, Paredes R, Noguera-Julian

M. Probiotic effects on immunity and microbiome in HIV-1 discordant patients. Front Immunol. 2022;13:1066036.

- Serrano-Villar S, Talavera-Rodriguez A, Gosalbes MJ, Madrid N, Perez-Molina JA, Elliott RJ, Navia B, Lanza VF, Vallejo A, Osman M, Dronda F, Budree S, Zamora J, Gutierrez C, Manzano M, Vivancos MJ, Ron R, Martinez-Sanz J, Herrera S, Ansa U, Moya A, Moreno S. Fecal microbiota transplantation in HIV: a pilot placebo-controlled study. Nat Commun. 2021;12:1139.
- 76. Pastor-Ibanez R, Blanco-Heredia J, Etcheverry F, Sanchez-Palomino S, Diez-Fuertes F, Casas R, Navarrete-Munoz MA, Castro-Barquero S, Lucero C, Fernandez I, Leal L, Benito JM, Noguera-Julian M, Paredes R, Rallon N, Estruch R, Torrents D, Garcia F. Adherence to a supplemented Mediterranean diet drives changes in the gut microbiota of HIV-1-infected individuals. Nutrients. 2021;13:1141.
- Bass NM, Mullen KD, Sanyal A, Poordad F, Neff G, Leevy CB, Sigal S, Sheikh MY, Beavers K, Frederick T, Teperman L, Hillebrand D, Huang S, Merchant K, Shaw A, Bortey E, Forbes WP. Rifaximin treatment in hepatic encephalopathy. N Engl J Med. 2010;362:1071–81.
- Tenorio AR, Chan ES, Bosch RJ, Macatangay BJ, Read SW, Yesmin S, Taiwo B, Margolis DM, Jacobson JM, Landay AL, Wilson CC, Team A. Rifaximin has a marginal impact on microbial translocation, T-cell activation and inflammation in HIV-positive immune non-responders to antiretroviral therapy - ACTG A5286. J Infect Dis. 2015;211:780–90.
- 79. Williams BB, Green SJ, Bosch RJ, Chan ES, Jacobson JM, Margolis DM, Engen P, Landay AL, Wilson CC, team Ap. Four weeks of treatment with rifaximin fails to significantly alter microbial diversity in rectal samples of HIV-infected immune non-responders (ACTG A5286) which may be attributed to rectal swab use. Pathog Immun. 2019;4:235–50.
- Kyosiimire-Lugemwa J, Anywaine Z, Abaasa A, Levin J, Gombe B, Musinguzi K, Kaleebu P, Grosskurth H, Munderi P, Pala P. Effect of stopping cotrimoxazole preventive therapy on microbial translocation and inflammatory markers among human immunodeficiency virus-infected Ugandan adults on antiretroviral therapy: the COSTOP Trial Immunology Substudy. J Infect Dis. 2020;222:381–90.
- D'Souza AW, Moodley-Govender E, Berla B, Kelkar T, Wang B, Sun X, Daniels B, Coutsoudis A, Trehan I, Dantas G. Cotrimoxazole prophylaxis increases resistance gene prevalence and alpha-diversity but decreases beta-diversity in the gut microbiome of human immunodeficiency virus-exposed, uninfected infants. Clin Infect Dis. 2020;71:2858–68.
- 82. Bourke CD, Gough EK, Pimundu G, Shonhai A, Berejena C, Terry L, Baumard L, Choudhry N, Karmali Y, Bwakura-Dangarembizi M, Musiime V, Lutaakome J, Kekitiinwa A, Mutasa K, Szubert AJ, Spyer MJ, Deayton JR, Glass M, Geum HM, Pardieu C, Gibb DM, Klein N, Edens TJ, Walker AS, Manges AR, Prendergast AJ. Cotrimoxazole reduces systemic inflammation in HIV infection by altering the gut microbiome and immune activation. Sci Transl Med. 2019;11:eaav0537.
- Mirza KA, Nietzsche S, Makarewicz O, Pletz MW, Thieme L. Bacteriophage-mediated decolonization of Klebsiella pneumoniae in a novel Galleria mellonella gut colonization model with Enterobacteriaceae. Sci Rep. 2024;14:318.
- Xiao Y, Zhai Q, Zhang H, Chen W, Hill C. Gut colonization mechanisms of Lactobacillus and Bifidobacterium: an argument for personalized designs. Annu Rev Food Sci Technol. 2021;12:213–33.
- Jadhav A, Bajaj A, Xiao Y, Markandey M, Ahuja V, Kashyap PC. Role of diet-microbiome interaction in gastrointestinal disorders and strategies to modulate them with microbiome-targeted therapies. Annu Rev Nutr. 2023;43:355–83.
- Manuzak JA, Hensley-McBain T, Zevin AS, Miller C, Cubas R, Agricola B, Gile J, Richert-Spuhler L, Patilea G, Estes JD, Langevin S, Reeves RK, Haddad EK, Klatt NR. Enhancement of microbiota in healthy macaques results in beneficial modulation of mucosal and systemic immune function. J Immunol. 2016;196:2401–9.
- Lynn DJ, Benson SC, Lynn MA, Pulendran B. Modulation of immune responses to vaccination by the microbiota: implications and potential mechanisms. Nat Rev Immunol. 2022;22:33–46.
- Klatt NR, Broedlow C, Osborn JM, Gustin AT, Dross S, O'Connor MA, Coronado E, Barnette P, Hensley-McBain T, Zevin AS, Muir R, Roederer A, Wangari S, Iwayama N, Ahrens CY, Smedley J, Moats C, Lynch RM, Haddad EK, Haigwood NL, Fuller DH, Manuzak JA. Effects of persistent modulation of intestinal microbiota on SIV/HIV vaccination in rhesus macaques. NPJ Vaccines. 2021;6:34.

- Vujkovic-Cvijin I, Swainson LA, Chu SN, Ortiz AM, Santee CA, Petriello A, Dunham RM, Fadrosh DW, Lin DL, Faruqi AA, Huang Y, Apetrei C, Pandrea I, Hecht FM, Pilcher CD, Klatt NR, Brenchley JM, Lynch SV, McCune JM. Gut-resident Lactobacillus abundance associates with IDO1 inhibition and Th17 dynamics in SIV-infected macaques. Cell Rep. 2015;13:1589–97.
- Crakes KR, Santos Rocha C, Grishina I, Hirao LA, Napoli E, Gaulke CA, Fenton A, Datta S, Arredondo J, Marco ML, Sankaran-Walters S, Cortopassi G, Giulivi C, Dandekar S. PPARalpha-targeted mitochondrial bioenergetics mediate repair of intestinal barriers at the hostmicrobe intersection during SIV infection. Proc Natl Acad Sci U S A. 2019;116:24819–29.
- Tincati C, Merlini E, Braidotti P, Ancona G, Savi F, Tosi D, Borghi E, Callegari ML, Mangiavillano B, Barassi A, Bulfamante G, d'Arminio Monforte A, Romagnoli S, Chomont N, Marchetti G. Impaired gut junctional complexes feature late-treated individuals with suboptimal CD4+T-cell recovery upon virologically suppressive combination antiretroviral therapy. AIDS. 2016;30:991–1003.
- Chung CY, Alden SL, Fu P, Funderburg NT, Levine AD. Progressive proximal-to-distal reduction in expression of the tight junction complex in colonic epithelium of virally-suppressed HIV+ individuals. Plos Pathog. 2014;10:e1004198.
- Li SX, Sen S, Schneider JM, Xiong KN, Nusbacher NM, Moreno-Huizar N, Shaffer M, Armstrong AJS, Severs E, Kuhn K, Neff CP, McCarter M, Campbell T, Lozupone CA, Palmer BE. Gut microbiota from high-risk men who have sex with men drive immune activation in gnotobiotic mice and in vitro HIV infection. Plos Pathog. 2019;15:e1007611.
- Somsouk M, Estes JD, Deleage C, Dunham RM, Albright R, Inadomi JM, Martin JN, Deeks SG, McCune JM, Hunt PW. Gut epithelial barrier and systemic inflammation during chronic HIV infection. AIDS. 2015;29:43–51.
- Tenorio AR, Zheng Y, Bosch RJ, Krishnan S, Rodriguez B, Hunt PW, Plants J, Seth A, Wilson CC, Deeks SG, Lederman MM, Landay AL. Soluble markers of inflammation and coagulation but not T-cell activation predict non-AIDS-defining morbid events during suppressive antiretroviral treatment. J Infect Dis. 2014;210:1248–59.
- Hunt PW, Sinclair E, Rodriguez B, Shive C, Clagett B, Funderburg N, Robinson J, Huang Y, Epling L, Martin JN, Deeks SG, Meinert CL, Van Natta ML, Jabs DA, Lederman MM. Gut epithelial barrier dysfunction and innate immune activation predict mortality in treated HIV infection. J Infect Dis. 2014;210:1228–38.
- Serrano-Villar S, Vazquez-Castellanos JF, Vallejo A, Latorre A, Sainz T, Ferrando-Martinez S, Rojo D, Martinez-Botas J, Del Romero J, Madrid N, Leal M, Mosele JI, Motilva MJ, Barbas C, Ferrer M, Moya A, Moreno S, Gosalbes MJ, Estrada V. The effects of prebiotics on microbial dysbiosis, butyrate production and immunity in HIV-infected subjects. Mucosal Immunol. 2017;10:1279–93.
- Sainz T, Gosalbes MJ, Talavera-Rodriguez A, Jimenez-Hernandez N, Prieto L, Escosa L, Guillen S, Ramos JT, Munoz-Fernandez MA, Moya A, Navarro ML, Mellado MJ, Serrano-Villar S. Effect of a nutritional intervention on the intestinal microbiota of vertically HIV-infected children: the Pediabiota Study. Nutrients. 2020;12:2112.
- Sainz T, Diaz L, Rojo D, Clemente MI, Barbas C, Gosalbes MJ, Jimenez-Hernandez N, Escosa L, Guillen S, Ramos JT, Munoz-Fernandez MA, Navarro ML, Mellado MJ, Serrano-Villar S. Targeting the gut microbiota of vertically HIV-infected children to decrease inflammation and immunoactivation: a pilot clinical trial. Nutrients. 2022;14:992.
- 100. Zhang ZJ, Lehmann CJ, Cole CG, Pamer EG. Translating microbiome research from and to the clinic. Annu Rev Microbiol. 2022;76:435–60.
- Yadegar A, Pakpoor S, Ibrahim FF, Nabavi-Rad A, Cook L, Walter J, Seekatz AM, Wong K, Monaghan TM, Kao D. Beneficial effects of fecal microbiota transplantation in recurrent Clostridioides difficile infection. Cell Host Microbe. 2023;31:695–711.
- Vujkovic-Cvijin I, Rutishauser RL, Pao M, Hunt PW, Lynch SV, McCune JM, Somsouk M. Limited engraftment of donor microbiome via one-time fecal microbial transplantation in treated HIV-infected individuals. Gut Microbes. 2017;8:440–50.
- 103. Utay NS, Monczor AN, Somasunderam A, Lupo S, Jiang ZD, Alexander AS, Finkelman M, Vigil KJ, Lake JE, Hanson B, DuPont HL, Arduino RC. Evaluation of six weekly oral fecal microbiota transplants in people with HIV. Pathog Immun. 2020;5:364–81.

- 104. Feuerstadt P, Louie TJ, Lashner B, Wang EEL, Diao L, Bryant JA, Sims M, Kraft CS, Cohen SH, Berenson CS, Korman LY, Ford CB, Litcofsky KD, Lombardo MJ, Wortman JR, Wu H, Aunins JG, McChalicher CWJ, Winkler JA, McGovern BH, Trucksis M, Henn MR, von Moltke L. SER-109, an oral microbiome therapy for recurrent Clostridioides difficile infection. N Engl J Med. 2022;386:220–9.
- 105. Orenstein R, Dubberke ER, Khanna S, Lee CH, Yoho D, Johnson S, Hecht G, DuPont HL, Gerding DN, Blount KF, Mische S, Harvey A. Durable reduction of Clostridioides difficile infection recurrence and microbiome restoration after treatment with RBX2660: results from an open-label phase 2 clinical trial. BMC Infect Dis. 2022;22:245.
- Li M, van Esch B, Wagenaar GTM, Garssen J, Folkerts G, Henricks PAJ. Pro- and anti-inflammatory effects of short chain fatty acids on immune and endothelial cells. Eur J Pharmacol. 2018;831:52–9.
- 107. Hayashi A, Nagao-Kitamoto H, Kitamoto S, Kim CH, Kamada N. The butyrate-producing bacterium Clostridium butyricum suppresses Clostridioides difficile infection via neutrophil- and antimicrobial cytokine-dependent but GPR43/109a-independent mechanisms. J Immunol. 2021;206:1576–85.
- Dillon SM, Kibbie J, Lee EJ, Guo K, Santiago ML, Austin GL, Gianella S, Landay AL, Donovan AM, Frank DN, Mc CM, Wilson CC. Low abundance of colonic butyrate-producing bacteria in HIV infection is associated with microbial translocation and immune activation. AIDS. 2017;31:511–21.
- 109. Missailidis C, Sorensen N, Ashenafi S, Amogne W, Kassa E, Bekele A, Getachew M, Gebreselassie N, Aseffa A, Aderaye G, Andersson J, Brighenti S, Bergman P. Vitamin D and phenylbutyrate supplementation does not modulate gut derived immune activation in HIV-1. Nutrients. 2019;11:1675.
- Djuric Z. Dietary approaches for normalizing dysbiosis induced by highfat, obesogenic diets. Curr Opin Clin Nutr Metab Care. 2023;26:293–301.
- 111. Manzano M, Talavera-Rodriguez A, Moreno E, Madrid N, Gosalbes MJ, Ron R, Dronda F, Perez-Molina JA, Lanza VF, Diaz J, Moreno S, Navia B, Serrano-Villar S. Relationship of diet to gut microbiota and inflammatory biomarkers in people with HIV. Nutrients. 2022;14:1221.
- 112. Estruch R, Ros E, Salas-Salvado J, Covas MI, Corella D, Aros F, Gomez-Gracia E, Ruiz-Gutierrez V, Fiol M, Lapetra J, Lamuela-Raventos RM, Serra-Majem L, Pinto X, Basora J, Munoz MA, Sorli JV, Martinez JA, Fito M, Gea A, Hernan MA, Martinez-Gonzalez MA, Investigators PS. Primary prevention of cardiovascular disease with a Mediterranean diet supplemented with extra-virgin olive oil or nuts. N Engl J Med. 2018;378:e34.
- 113. Somsouk M, Dunham RM, Cohen M, Albright R, Abdel-Mohsen M, Liegler T, Lifson J, Piatak M, Gorelick R, Huang Y, Wu Y, Hsue PY, Martin JN, Deeks SG, McCune JM, Hunt PW. The immunologic effects of mesalamine in treated HIV-infected individuals with incomplete CD4+ T cell recovery: a randomized crossover trial. Plos One. 2014;9:e116306.
- 114. Gacesa R, Kurilshikov A, Vich Vila A, Sinha T, Klaassen MAY, Bolte LA, Andreu-Sanchez S, Chen L, Collij V, Hu S, Dekens JAM, Lenters VC, Bjork JR, Swarte JC, Swertz MA, Jansen BH, Gelderloos-Arends J, Jankipersadsing S, Hofker M, Vermeulen RCH, Sanna S, Harmsen HJM, Wijmenga C, Fu J, Zhernakova A, Weersma RK. Environmental factors shaping the gut microbiome in a Dutch population. Nature. 2022;604:732–9.
- Vandeputte D, De Commer L, Tito RY, Kathagen G, Sabino J, Vermeire S, Faust K, Raes J. Temporal variability in quantitative human gut microbiome profiles and implications for clinical research. Nat Commun. 2021;12:6740.
- 116. Rodriguez Del Rio A, Giner-Lamia J, Cantalapiedra CP, Botas J, Deng Z, Hernandez-Plaza A, Munar-Palmer M, Santamaria-Hernando S, Rodriguez-Herva JJ, Ruscheweyh HJ, Paoli L, Schmidt TSB, Sunagawa S, Bork P, Lopez-Solanilla E, Coelho LP, Huerta-Cepas J. Functional and evolutionary significance of unknown genes from uncultivated taxa. Nature. 2024;626:377–84.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.