



Target Sequence Cloning Protocol

(Standard de-salting oligos are sufficient)

PX330-based plasmids, including PX458-462 – SpCas9 (or SpCas9n D10A nickase) + single guide RNA:

To clone the guide sequence into the sgRNA scaffold, synthesize two oligos of the form:



PX260 and PX334 – SpCas9 (or SpCas9n D10A nickase) + CRISPR array + tracrRNA:

To clone the guide sequence into the sgRNA scaffold, synthesize two oligos of the form:



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Oligo annealing and cloning into backbone vectors:

1. Digest 1ug of plasmid with *Bbs*I for 30 min at 37°C;
 4. Set up ligation reaction and incubate at room temperature for 10 min:

1 ug	Plasmid
1 ul	FastDigest <i>Bbs</i> I (Fermentas)
1 ul	FastAP (Fermentas)
2 ul	10X FastDigest Buffer
X ul	<i>ddH</i> ₂ O
20 ul	total

2. Gel purify digested plasmid using QIAquick Gel Extraction Kit and elute in EB.
 3. Phosphorylate and anneal each pair of oligos

1 ul	oligo 1 (100uM)
1 ul	oligo 2 (100uM)
1 ul	10X T4 Ligation Buffer (NEB)
6.5 ul	ddH ₂ O
<u>0.5 ul</u>	<u>T4 PNK (NEB)</u>
10 ul	total

Anneal in a thermocycler using the following parameters:

37°C 30 min
95°C 5 min and then ramp down to
25°C at 5°C/min

4. Set up ligation reaction and incubate at room temperature for 10 min:

X ul	<i>BbsI</i> digested plasmid from step 2 (50ng)
1 ul	phosphorylated and annealed oligo duplex from step 3 (1:200 dilution)
5 ul	2X Quickligation Buffer (NEB)
X ul	<u>ddH₂O</u>
10 ul	subtotal
1 ul	Quick Ligase (NEB)
11 ul	total

5. (optional) Treat ligation reaction with PlasmidSafe exonuclease to prevent unwanted recombination products:

11 ul ligation reaction from **step 4**
1.5 ul 10X PlasmidSafe Buffer
1.5 ul 10mM ATP
1 ul exonuclease

15 ul total

Incubate reaction at 37C for 30 min.

- ## 6. Transformation