RESEARCH ARTICLE

Serial Analysis of Gene Expression Reveals Promising Therapeutic Targets for Liver Fluke-associated Cholangiocarcinoma

Kanlayanee Sawanyawisuth^{1*}, Chaisiri Wongkham¹, Norie Araki², Qi Zhao³, Gregory J Riggins³, Sopit Wongkham^{1*}

Abstract

Cholangiocarcinoma (CCA) continues to be a serious health problem and is the most common fatal cancer in northeastern Thailand. Comprehensive gene expression analysis was here used to identify possible therapeutic targets for CCA treatment. We assessed liver fluke-associated CCA tissue using serial analysis of gene expression (SAGE) and compared the data to normal liver tissue as a part of the Cancer Genome Anatomy Project (CGAP). The analysis identified 509 differentially expressed genes. Of 142 up-regulated examples, we selected candidates including TMSB10, GAL3, VDR, CYPA and CD147 for further validation in CCA tissues by immunohistochemistry. VDR, CYPA and CD147 were confirmed to be consistently overexpressed in the samples tested. The therapeutic and diagnostic potential of these genes warrants further investigation.

Keywords: Cholangiocarcinoma - bile duct cancer - serial analysis of gene expression - SAGE

Asian Pacific J Cancer Prev, 13, 89-93

Introduction

Cholangiocarcinoma (CCA) is the most common liver cancer in the northeastern Thailand where the incidence rate is the highest in the world (Shin et al., 2010). Liver fluke (Opisthorchis viverrini) infection has been proven to be the major risk factor of CCA in this endemic area (Sripa et al., 2007). Most CCA patients present at the invasion/ metastasis stage and this leads to a high mortality rate (Patel, 2002). Discovery of new promising therapeutic targets may be the alternative approach for CCA treatment. Progression of tumor cells toward a high malignancy phenotype and metastasis is a multi-event cascade involving alterations in the expression of various genes. Thus, global gene expression profiling is the appropriate tool to study this complex disease. Serial analysis of gene expression (SAGE) (Velculescu et al., 1995) has been used to identify differentially expressed genes in many types of human cancer. This quantitative method for high throughput gene expression analysis can be compared with experiments from different laboratories done at different times (Boon et al., 2002). The Cancer Genome Anatomy Project (CGAP) provides a SAGE database of various normal and cancer tissues and also several online tools on this user-friendly website http://cgap.nci.nih.gov/SAGE. To identify the potential therapeutic targets, we performed a large scale gene expression profiling of liver flukeassociated CCA tissues using SAGE and then compared these profiles with the gene profile of normal liver provided from the SAGE public database. Among the differentially expressed genes, we focused on the up-regulated genes whose altered expression may lead directly or indirectly to an increased malignancy phenotype. We finally selected 5 candidate genes (TMSB10, GAL3, VDR, CYPA and its receptor CD147) for further validation of SAGE analysis in different sets of CCA tissues by immunohistochemistry.

Materials and Methods

CCA patient samples for SAGE

The paired CCA tissues were selected to establish two SAGE libraries: an intrahepatic metastatic nodule (SAGE_ Liver_Cholangiocarcinoma_B_K1) and a primary CCA tumor tissue (SAGE_Liver_Cholangiocarcinoma_B_ K2D). The tissues were obtained from a Thai male, 55 years old with an intrahepatic mass forming CCA. The histology proved to be a poorly differentiated adenocarcinoma (Figure 1). Tumor invaded the intrahepatic vein, and invaded beyond Glisson's capsule involving diaphragm, right adrenal gland and peritoneum. The cut sections for establishing SAGE contained more than a 60% CCA cell population as analyzed by histology.

SAGE library construction and analysis

Construction of the SAGE library was performed following the instructions of the Micro-SAGE protocol

¹Department of Biochemistry, and Liver Fluke and Cholangiocarcinoma Research Center, Faculty of Medicine, Khon Kaen University, Khon Kaen, Thailand, ²Department of Tumor Genetics and Biology, Graduate School of Medical Sciences, Kumamoto University, Kumamoto, Japan, ³Department of Neurosurgery, School of Medicine, Johns Hopkins University, Baltimore, USA *For correspondence: kanlayanee@kkumail.com, sopit@kku.ac.th



Figure 1. Histopathological Examination of the Samples Used for SAGE Library Construction. Hematoxylin and eosin staining for (A) an intrahepatic metastatic nodule and (B) a primary CCA

(Datson et al., 1999). RNA was isolated using RNeasy (Qiagen) according to manufacturer's instructions. Five µg of total RNA was used as starting material. The SAGE library clones containing inserts were purified and sequenced by Agencourt through the SAGE sequencing service (CGAP collaboration, GR). Sequence data from SAGE libraries were posted on the CGAP website http:// cgap.nci.nih.gov/SAGE.

The SAGE Differential Gene Expression Display (DGED) tool was used to compare SAGE data between the pools of two CCA samples with a normal human liver which was provided on the CGAP website. DGED results were reported only when odds ratio (fold change) was significantly greater than 2 or significantly less than 0.5. The odds ratio was calculated by the following formula [(Tag count sum/ total tag count of CCA libraries)/ (Tag count/ total tag count of normal liver library)].

Immunohistochemistry

Surgical specimens from CCA patients who underwent operations in Srinagarind hospital, Faculty of Medicine, Khon Kaen University, Thailand were collected. Informed consent was obtained from all patients before surgery, and the research protocol (#HE471214) was approved by the Human Research Ethics Committee at Khon Kaen University. All specimens were histologically proved and subjected to the standard protocol. Briefly, deparaffinized 4 µm sections of formalin-fixed specimens were submitted to heat-induced antigen retrieval, incubated with the specific antibodies, and processed using the 3,3'-diaminobenzidine (DAB) as the chromatogen. The sections were then counterstained with hematoxylin and mounted with coverslips. The optimized dilutions used were previously reported (Seubwai et al., 2007; Junking et al., 2008; Obchoei et al., 2011)

Results

SAGE profiles of CCA

SAGE data of two CCA tissues generated by us were pooled and compared with SAGE data of normal liver tissues (library name: SAGE_Liver_normal_B_1) provided on the CGAP website by the SAGE DGED online tool. The numbers of total tags and unique tags of each library are shown in Table 1. At least 46,000 total tags were reported in CCA libraries. Using the criteria mentioned in the method, we identified 620 SAGE tags as differentially expressed. Of these, 111 tags were undefined or mapped to incomplete cDNA clones on the SAGE database while 509 tags (142 up-regulated and 367 down-regulated) were assigned to genes. These genes were analyzed by Ingenuity pathway analysis (IPA; Ingenuity[®] Systems, http://www.ingenuity.com) in order to determine enriched functional categories. As seen in Figure 2, the three most abundant categories found in this analysis were lipid metabolism, energy production and molecular transport.

Selection of therapeutic targets for CCA

To identify the therapeutic targets for CCA, we selected 4 up-regulated genes, TMSB10, LGALS3 (GAL3), VDR and PPIA (CYPA) as shown in Table 2 based on these criteria: (i) gene function involved in carcinogenesis/ metastasis; (ii) had specific drugs or inhibitors which could be used for targeted therapy; and (iii) the availability of antibodies that label formalin-fixed paraffin-embedded archival material. We further previewed the expression levels of these selected candidate genes in normal and tumor tissues of various gastrointestinal tract organs such as stomach, pancreas and colon by the SAGE Anatomic viewer tool as shown in Figure 3.

Validation of SAGE data by immunohistochemistry (IHC) of selected candidates

Selected up-regulated genes in CCA including TMSB10, GAL3, VDR and CYPA were investigated in an independent set of CCA samples by the IHC method in order to validate the SAGE data using a different technique and to localize the origin of cell types that expressed the selected genes. The data demonstrated that TMSB10 and GAL3 were detected in normal and malignant biliary cells. High intensity of positive staining of these two proteins was demonstrated in most of CCA tissues. Moreover, VDR and CYPA were significantly overexpressed at the protein

Table 1. SAGE Library Information

Library	CCA_K1	CCA_K2D	Normal liver
Total clones	2,592	2,688	2,688
Total tags	60,319	46,853	66,308
Unique tags	40,476	20,722	15,496
No. of differentially expressed	509		
Up-regulated		142	
Down-regulated		367	



Figure 2. Functional Categories Assigned to Differentially Expressed Genes by IPA. There are 433 genes included in the gene ontology classification

SAGE tag	CCA	Normal	Odds	Unigene cluster	Gene symbol	Gene name
	(K1+K2D)) liver	ratio			
GGGGAAATCG	48	0	NaN	Hs.446574	TMSB10	Thymosin beta 10
TTCACTGTGA	100	1	61.93	Hs.531081	LGALS3, GAL3	Lectin, galactoside-binding, soluble, 3
GAGAAACCCT	25	2	7.74	Hs.524368	VDR	Vitamin D (1,25- dihydroxyvitamin D_3) receptor
CCTAGCTGGA	232	52	2.76	Hs.356331	PPIA, CYPA	Peptidylprolyl isomerase A (cyclophilin A)

Table 2. Information of 4 Selected Candidates

*NaN stands for not a number



Figure 3. Expression Levels of 4 Selected Up-Regulated Genes in Other Gastrointestinal (GI) Tract Organs.25.0 Up-regulation of these genes is also demonstrated in other GI tract cancers, indicating cancer associated function. Data obtained from SAGE anatomic viewer tool. N, normal; T, tumor; Stomach (S): SAGE_Stomach_normal_epithelium_B_ body1 and SAGE_Stomach_carcinoma_B_G189. Pancreas (P): SAGE_Pancreas_normal_B_1 and SAGE_Pancreas_ adenocarcinoma_B_91-16113. Colon (C): SAGE_Colon_ normal_B_NC1and SAGE_Colon_adenocarcinoma_B_Tu102

level in CCA tissues compared to normal bile ducts. We next investigated the cell surface signaling receptor of CD147 or extracellular matrix metalloproteinase inducer (EMMPRIN). CD147 expression was also highly elevated in CCA tissues and correlated with CYPA expression (Figure 4).

Discussion

The identification of genes differentially expressed in CCA tissues relative to normal tissues provides a basis for the development of novel strategies to detect and treat this highly lethal cancer. Because SAGE data can be compared between studies and laboratories, we constructed SAGE libraries of CCA and then compared to the publicly available SAGE library of normal liver tissue by using the SAGE DGED online tool on the CGAP website. We identified 509 genes that are differentially expressed in CCA. Of these 142 were up-regulated and 367 downregulated relative normal liver tissues. These genes were functionally categorized based on their gene ontology classification. The most frequent tags correspond to genes involved in lipid metabolism, energy production and molecular transport. These findings were as expected since liver is a major organ in energy metabolism. Consistently, alteration of several lipid-related pathways including apolipoproteins (APOA1, APOA2) and fatty acid metabolism (fatty acid binding protein 1, liver; FABP1) were documented in intrahepatic cholangiocarcinoma (ICC) (Nishino et al., 2008).

Among 142 up-regulated genes, we validated 4 selected genes from SAGE comparison including TMSB10,



(TMSB10, GAL3, VDR, CYPA and CD147) are shown. The positive staining of The SB10 and GAL3 was found in normal bile duct of liver and CCA tissues. Overexpression of VDR, CYPA and 20147 was biviously detected in CA tissues. DAB staining; or ginal maggification 20X

GAL3, VBR and CVPA using immunohistochemistry. Their expression levels of normal and tumor tissues in a variety of gastroinestinal tract organs were analyzed by the SAGE Anatomic viewer (http://cgap.nci.nih.gov/ SAGE/Aratomic Viewer). The results indicated that most of the differentially expressed genes in CCA were also differentially expressed in other cancers. Noteworthy, the pattern of 4 selected genes of CCA were similar to those of stomach and pancreatic cancers.

IHC analysis demonstrated that TMSB10 and GAL3 were constitutively expressed in normal bile duct epithelia even their SAGE tag counts were very low. Although a count of zero tags indicates absence of detection in SAGE analysis, there may still be expression below the level of detection, which in this case is approximately 1 transcript per 50,000. The expression of these 2 genes was not drastically changed among the normal bile duct epithelia, but markedly enhanced in CCA cells. However, we additionally investigated TMSB10 protein expression in match pairs of primary and metastatic CCA. Preliminary results showed that under expression of TMSB10 has been found in metastatic tumors (unpublished manuscript). TMSB10 is the abbreviated gene symbol for thymosin β 10 which is widely distributed in many tissues with proven biological activities as an actin sequestering protein involved in cell motility. It has been reported to correlate with tumor biology such as cell proliferation, apoptosis, angiogenesis and metastasis behavior of several types of human cancers (Chen et al., 2005; Sribenja et al., 2009). The roles of TMSB10 and its signaling pathway in CCA cell migration and metastasis is now under investigation

Kanlayanee Sawanyawisuth et al

in our laboratory.

GAL3, a β -galactoside-binding lectin, is a multifunctional protein implicated in a variety of biological functions, including tumor cell adhesion, proliferation, differentiation, cancer progression and metastasis (Castronovo et al., 1996). Our previous published study suggested that low GAL3 expression was significantly associated with lymphatic invasion. Suppression of GAL3 expression in two human CCA cell lines using siRNA substantially increased cell migration and invasion of CCA cells without alterations in cell proliferation (Junking et al., 2008). Similar to what we observed, Shimonishi and colleagues reported that GAL3 tends to disappear at later stages of ICC (Shimonishi et al., 2001). Taken together, regulation of GAL3 expression may therefore be an alternative therapeutic approach to control metastasis of CCA.

The vitamin D (1,25- dihydroxyvitamin D_3) receptor or VDR belongs to the steroid/thyroid hormone nuclear receptor superfamily. VDR up-regulation has been shown in primary tumor tissues of breast, colon and pancreatic cancers (Friedrich et al., 1998; Cross et al., 2001; Albrechtsson et al., 2003).

Validation of the VDR expression pattern corresponded to SAGE analysis. VDR was rarely expressed in normal bile duct epithelia but highly expressed in 74% of CCA tissues. The survival rate of CCA patients with positive VDR expression in tumor tissue was significantly better than that of patients with negative expression of VDR. In addition, treatment with $1,25(OH)_2D_3$, an active metabolite of vitamin D₂, in the CCA cell lines with high expression of VDR significantly reduced cell proliferation in a dose-dependent manner. The effect was not shown in lower VDR expressing CCA cell lines (Seubwai et al., 2007). These results are consistent with those seen in carcinomas of gastric, breast and colon (Albrechtsson et al., 2003; Banerjee and Chatterjee, 2003; Pelczynska et al., 2005). These findings suggest that supplementation of $1,25(OH)_2D_3$ or its analogs may be a potential strategy for long-term control of tumor development and progression in CCA patients. To prove this hypothesis, Seubwai et al. recently reported that supplementation of 22-oxa-D₂ to CCA-inoculated mice effectively suppressed tumor growth and induced cellular apoptosis in tissue samples from patients with CCA analyzed by using a histodrug response assay (Seubwai et al., 2010). These data encourage further investigation of 1,25(OH)₂D₂ or its analogues as therapeutic agents in the treatment of CCA patients.

Cyclophilin A (CYPA) is an 18 kDa cytosolic protein that is thought to be the major intracellular target of the immunosuppressive drug cyclosporin A (CsA) (Handschumacher et al., 1984). Various forms of evidence exhibited elevation of CYPA in several types of cancers including non-small cell lung cancer, pancreatic adenocarcinoma, hepatocellular carcinoma, oral cancer, buccal squamous cell carcinoma and CCA (Campa et al., 2003; Howard et al., 2004; Obama et al., 2005; Yang et al., 2005; Li et al., 2006; Wang et al., 2006). Consistent with the literature, strongly positive staining of CYPA in CCA tissues has been revealed. We have recently reported that increasing of CYPA expression could accelerate CCA cell proliferation and tumor growth. Moreover, treatment of CsA, an inhibitor of CYPA inhibited CCA cell proliferation in a dose-dependent fashion (Obchoei et al., 2011). Application of CsA or CYPA deletion may be used as the molecular target for CCA therapy.

Interplay of CYPA and CD147/EMMPRIN (extracellular matrix metalloproteinase inducer) has been demonstrated to affect cell proliferation, migration and differentiation (Sherry et al., 1992; Kim et al., 2004; Yang et al., 2005). These citations led us to determine the expression pattern of CD147 in CCA tissues. Highly elevated CD147 expression in the studied CCA tissues has been detected. The results were consistent with neoplasias such as lung, breast, oral squamous cell carcinoma, melanoma and follicular thyroid carcinoma (Polette et al., 1997; Bordador et al., 2000; Kanekura et al., 2002; Omi et al., 2012). CD147, is a transmembrane glycoprotein that is categorized as a member of the immunoglobulin superfamily and can stimulate both tumor cells and nearby fibroblasts and endothelial cells to produce MMPs, facilitating cancer cell invasion (Nabeshima et al., 2006). In 2006, Chen et al. developed iodine (¹³¹I) metuximab injection (Licartin), a novel CD147-specific monoclonal antibody which was safe and active for hepatocellular carcinoma (HCC) patients in phase I/II trials (Chen et al., 2006). This information suggest that CD147 may be the potential target for cancer treatment.

In conclusion, we constructed SAGE libraries of liver fluke- associated CCA. These gene expression profiles provided candidate genes including TMSB10, GAL3, VDR, CYPA and CD147 which are potentially involved in carcinogenesis, invasion/ metastasis of CCA. Furthermore, these selected candidates may represent some promising therapeutic targets for CCA.

Acknowledgements

This work was co-supported by the Higher Education Research Promotion and National Research University Project of Thailand, Office of the Higher Education Commission, through the Health Cluster (SHeP-GMS), Khon Kaen University, the research grant from Khon Kaen University and the National Science and Technology Development Agency (NSTDA), Thailand. GJR is supported by the Irving J. Sherman Research Professorship in Neurosurgery and the Virginia and D.K. Ludwig Fund for Cancer Research. We gratefully acknowledge Junking M, Seubwai W, Obchoei S and Sribenja S who provided the IHC data. We wish to acknowledge the support of the Khon Kaen University Publication Clinic, Research and Technology Transfer Affairs, Khon Kaen University, for their assistance.

References

- Albrechtsson E, Jonsson T, Moller S, et al (2003). Vitamin D receptor is expressed in pancreatic cancer cells and a vitamin D3 analogue decreases cell number. *Pancreatology*, 3, 41-6.
- Banerjee P, Chatterjee M (2003). Antiproliferative role of vitamin D and its analogs-a brief overview. *Mol Cell Biochem*, **253**,

DOI:http://dx.doi.org/10.7314/APJCP.2012.13.KKSuppl.89 Serial Analysis of Gene Expression (SAGE) Analysis of Fluke-associated CCA

247-54.

- Boon K, Osorio EC, Greenhut SF, et al (2002). An anatomy of normal and malignant gene expression. *Proc Natl Acad Sci* USA, 99, 11287-92.
- Bordador LC, Li X, Toole B, et al (2000). Expression of emmprin by oral squamous cell carcinoma. *Int J Cancer*, **85**, 347-52.
- Campa MJ, Wang MZ, Howard B, et al (2003). Protein expression profiling identifies macrophage migration inhibitory factor and cyclophilin a as potential molecular targets in non-small cell lung cancer. *Cancer Res*, **63**, 1652-6.
- Castronovo V, Van Den Brule FA, Jackers P, et al (1996). Decreased expression of galectin-3 is associated with progression of human breast cancer. *J Pathol*, **179**, 43-8.
- Chen C, Li M, Yang H, et al (2005). Roles of thymosins in cancers and other organ systems. *World J Surg*, **29**, 264-70.
- Chen ZN, Mi L, Xu J, et al (2006). Targeting radioimmunotherapy of hepatocellular carcinoma with iodine (131I) metuximab injection: clinical phase I/II trials. *Int J Radiat Oncol Biol Phys*, **65**, 435-44.
- Cross HS, Bareis P, Hofer H, et al (2001). 25-Hydroxyvitamin D(3)-1alpha-hydroxylase and vitamin D receptor gene expression in human colonic mucosa is elevated during early cancerogenesis. *Steroids*, **66**, 287-92.
- Datson NA, van der Perk-de Jong J, van den Berg MP, et al (1999). MicroSAGE: a modified procedure for serial analysis of gene expression in limited amounts of tissue. *Nucleic Acids Res*, **27**, 1300-7.
- Friedrich M, Rafi L, Tilgen W, et al (1998). Expression of 1,25-dihydroxy vitamin D3 receptor in breast carcinoma. J Histochem Cytochem, 46, 1335-7.
- Handschumacher RE, Harding MW, Rice J, et al (1984). Cyclophilin: a specific cytosolic binding protein for cyclosporin A. Science, 226, 544-7.
- Howard BA, Zheng Z, Campa MJ, et al (2004). Translating biomarkers into clinical practice: prognostic implications of cyclophilin A and macrophage migratory inhibitory factor identified from protein expression profiles in non-small cell lung cancer. *Lung Cancer*, **46**, 313-23.
- Junking M, Wongkham C, Sripa B, et al (2008). Decreased expression of galectin-3 is associated with metastatic potential of liver fluke-associated cholangiocarcinoma. *Eur J Cancer*, 44, 619-26.
- Kanekura T, Chen X, Kanzaki T (2002). Basigin (CD147) is expressed on melanoma cells and induces tumor cell invasion by stimulating production of matrix metalloproteinases by fibroblasts. *Int J Cancer*, **99**, 520-8.
- Kim SH, Lessner SM, Sakurai Y, et al (2004). Cyclophilin A as a novel biphasic mediator of endothelial activation and dysfunction. *Am J Pathol*, **164**, 1567-74.
- Li M, Zhai Q, Bharadwaj U, et al (2006). Cyclophilin A is overexpressed in human pancreatic cancer cells and stimulates cell proliferation through CD147. *Cancer*, **106**, 2284-94.
- Nabeshima K, Iwasaki H, Koga K, et al (2006). Emmprin (basigin/CD147): matrix metalloproteinase modulator and multifunctional cell recognition molecule that plays a critical role in cancer progression. *Pathol Int*, **56**, 359-67.
- Nishino R, Honda M, Yamashita T, et al (2008). Identification of novel candidate tumour marker genes for intrahepatic cholangiocarcinoma. *J Hepatol*, **49**, 207-16.
- Obama K, Ura K, Li M, et al (2005). Genome-wide analysis of gene expression in human intrahepatic cholangiocarcinoma. *Hepatology*, **41**, 1339-48.
- Obchoei S, Weakley SM, Wongkham S, et al (2011). Cyclophilin A enhances cell proliferation and tumor growth of liver fluke-associated cholangiocarcinoma. *Mol Cancer*, **10**, 102.
- Omi Y, Shibata N, Okamoto T, et al (2012). The role of CD147

in the invasiveness of follicular thyroid carcinoma cells. *Thyroid*, **22**, 383-94.

- Patel T (2002). Worldwide trends in mortality from biliary tract malignancies. *BMC Cancer*, **2**, 10.
- Pelczynska M, Wietrzyk J, Jaroszewicz I, et al (2005). Correlation between VDR expression and antiproliferative activity of vitamin D3 compounds in combination with cytostatics. *Anticancer Res*, **25**, 2235-40.
- Polette M, Gilles C, Marchand V, et al (1997). Tumor collagenase stimulatory factor (TCSF) expression and localization in human lung and breast cancers. J Histochem Cytochem, 45, 703-9.
- Seubwai W, Wongkham C, Puapairoj A, et al (2007). Overexpression of vitamin D receptor indicates a good prognosis for cholangiocarcinoma: implications for therapeutics. *Cancer*, **109**, 2497-505.
- Seubwai W, Wongkham C, Puapairoj A, et al (2010). 22-oxa-1,25-dihydroxyvitamin D3 efficiently inhibits tumor growth in inoculated mice and primary histoculture of cholangiocarcinoma. *Cancer*, **116**, 5535-43.
- Sherry B, Yarlett N, Strupp A, et al (1992). Identification of cyclophilin as a proinflammatory secretory product of lipopolysaccharide-activated macrophages. *Proc Natl Acad Sci USA*, 89, 3511-5.
- Shimonishi T, Miyazaki K, Kono N, et al (2001). Expression of endogenous galectin-1 and galectin-3 in intrahepatic cholangiocarcinoma. *Hum Pathol*, **32**, 302-10.
- Shin HR, Oh JK, Masuyer E, et al (2010). Comparison of incidence of intrahepatic and extrahepatic cholangiocarcinoma--focus on East and South-Eastern Asia. Asian Pac J Cancer Prev, 11, 1159-66.
- Sribenja S, Li M, Wongkham S, et al (2009). Advances in thymosin beta10 research: differential expression, molecular mechanisms, and clinical implications in cancer and other conditions. *Cancer Invest*, 27, 1016-22.
- Sripa B, Kaewkes S, Sithithaworn P, et al (2007). Liver fluke induces cholangiocarcinoma. PLoS Med, 4, e201.
- Velculescu VE, Zhang L, Vogelstein B, et al (1995). Serial analysis of gene expression. *Science*, 270, 484-7.
- Wang AG, Yoon SY, Oh JH, et al (2006). Identification of intrahepatic cholangiocarcinoma related genes by comparison with normal liver tissues using expressed sequence tags. *Biochem Biophys Res Commun*, 345, 1022-32.
- Yang H, Li M, Chai H, et al (2005). Effects of cyclophilin A on cell proliferation and gene expressions in human vascular smooth muscle cells and endothelial cells. *J Surg Res*, **123**, 312-9.