

Search keywords

Phylogeny and Evolution of Lepidoptera

Charles Mitter,^{1,*} Donald R. Davis,² and Michael P. Cummings³

¹Department of Entomology, University of Maryland, College Park, Maryland 20742; email: cmitter@umd.edu

²Department of Entomology, National Museum of Natural History, Smithsonian Institution, Washington, DC 20560

³Laboratory of Molecular Evolution, Center for Bioinformatics and Computational Biology, University of Maryland, College Park, Maryland 20742

Annu. Rev. Entomol. 2017. 62:265-83

First published online as a Review in Advance on November 16, 2016

The *Annual Review of Entomology* is online at ento.annualreviews.org

This article's doi: 10.1146/annurev-ento-031616-035125

Copyright © 2017 by Annual Reviews. All rights reserved

*Corresponding author

Keywords

Hexapoda, insect, systematics, classification, butterfly, moth, molecular systematics

Abstract

Until recently, deep-level phylogeny in Lepidoptera, the largest single radiation of plant-feeding insects, was very poorly understood. Over the past two decades, building on a preceding era of morphological cladistic studies, molecular data have yielded robust initial estimates of relationships both within and among the $\sim\!43$ superfamilies, with unsolved problems now yielding to much larger data sets from high-throughput sequencing. Here we summarize progress on lepidopteran phylogeny since 1975, emphasizing the superfamily level, and discuss some resulting advances in our understanding of lepidopteran evolution.

INTRODUCTION

Lepidoptera, arguably the largest single radiation of plant-feeding insects, also function ecologically as pollinators and prey. In addition, they have substantial impact on humans and many other species. Lepidoptera also provide important model systems for studies of genetics, physiology, development, ecology, and evolutionary biology.

A robust phylogenetic framework is essential for all attempts to understand the diversity, adaptations, and ecological roles of Lepidoptera. Until recently, however, deep-level lepidopteran phylogeny remained poorly known, and relationships within the \sim 43 superfamilies had only begun to receive concerted study. Following an era of morphological cladistic studies, molecular data since the 1990s have yielded robust initial estimates of relationships both within and among superfamilies. Using still larger data sets from high-throughput sequencing, researchers are now addressing previously unsolved problems. Here we summarize progress on lepidopteran phylogeny since 1975, focusing on the superfamily level (18).

OVERVIEW OF APPROACHES IN LEPIDOPTERAN PHYLOGENETICS FROM 1975 TO 2016

Modern lepidopteran phylogenetic research began in the 1970s and 1980s with detailed studies of anatomy coupled with early application of Hennigian phylogenetics (65, 66). This now-classic work focused mostly on the earliest-diverging lineages, the so-called nonditrysians, yielding a well-resolved hypothesis on the major subdivisions of the order (65, 66). Morphological-phylogenetic works then followed on later-originating major clades (83, 84) and subgroups therein (3, 4, 47, 58, 61, 69, 90, 101, 124). An edited volume (63) summarized progress as of 1998 and presented the first comprehensive working hypothesis of phylogeny within and among superfamilies.

Early molecular phylogenetic studies on Lepidoptera in the 1990s used the highly abundant sequences of the mitochondrial genome and nuclear ribosomal DNA (12, 14, 125, 134). Nuclear protein-coding genes were introduced several years later (13, 16, 38, 138), and their number expanded over the next 15 years (17, 93, 106, 130), with up to 19–26 in some recent studies (17, 111, 151). Much of the progress described below stems from this multigene approach based on the polymerase chain reaction. Although these methods remain useful, others are increasingly supplementing or superseding them. For deeper divergences, transcriptomes and genomes yielding hundreds to thousands of genes show strong resolving potential (8, 9, 11, 53). Whole mitochondrial genomes are also providing strong signals at multiple levels (57, 126, 140). The newest methods capture target DNA regions using conserved hybridization probes (37, 73), followed by high-throughput sequencing. In forthcoming lepidopteran studies, the anchored hybridization (73) approach has yielded hundreds of loci and strong resolution at modest pertaxon cost (10). This method seems likely to become widely applied for large-scale phylogenetic studies.

Morphological studies continue (100), and the first morphological character matrix across the Ditrysia (45) now permits quantitative comparisons of morphological versus molecular phylogenetic signal. In the next section, we attempt to synthesize all lines of evidence in summarizing the current understanding of relationships among lepidopteran superfamilies. We exclude species currently unplaced to superfamily or family as well as the little-known superfamilies Simaesthistoidea and Whalleyanoidea (127). **Figure 1** summarizes our conclusions and provides representative live-specimen images for the major lineages.

SYSTEMATIC POSITION AND MAJOR SUBDIVISIONS OF THE LEPIDOPTERA

The well-established sister group to Lepidoptera is Trichoptera (caddis flies), most of whose larvae are aquatic. Numerous synapomorphies support the monophyly of Lepidoptera (66). The most recent authoritative summary of lepidopteran diversity (127), our source for all such numbers except where otherwise specified, recognizes 157,424 extant species in 43 superfamilies and 133 families. The most securely established large subgroup of Lepidoptera is the clade Ditrysia, which contains 29 superfamilies and 98% of lepidopteran species (127).

Early-Diverging Lineages: The Nonditrysians

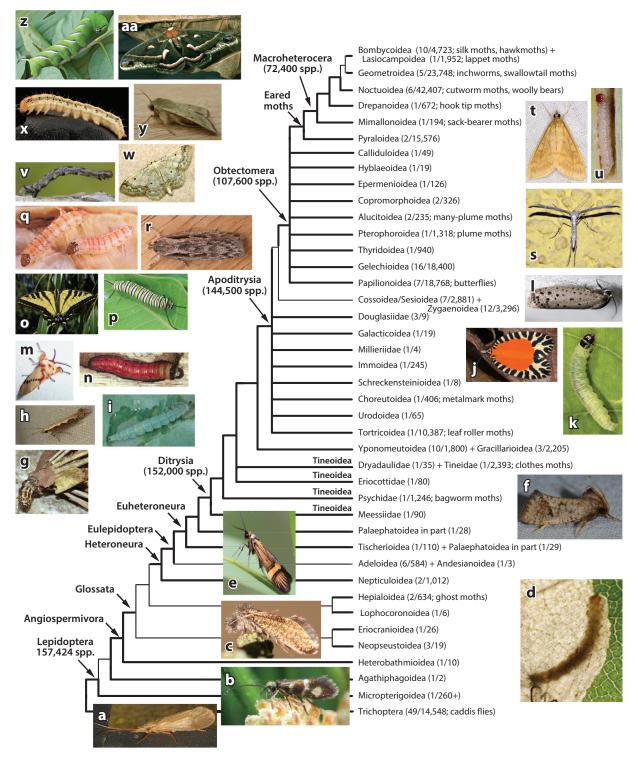
Nonditrysian Lepidoptera are a paraphyletic group of 14 superfamilies and ~21 families, most of which are species poor (23, 64, 109). The adults are mostly small (wingspan <10 mm), and most larvae are internal feeding. The morphological hypothesis (65, 66) features a nested series of clades corresponding to stepwise acquisition of traits characterizing most Lepidoptera. Molecular evidence strongly supports most of these clades, strongly contradicts several, and remains ambiguous about a few. The largest securely established clade is the newly named Angiospermivora (109), comprising all lepidopterans except Micropterigidae and Agathiphagidae and feeding in the larval stage almost always on angiosperms.

Micropterigidae, unlike most other Lepidoptera, retain functional mandibles in the adult, which feeds on pollen and fern spores. The larvae are also unusual: They dwell on or in the ground and feed mainly on liverworts or detritus. Agathiphagidae include just two Australo-Pacific species. The adult mandibles are apparently nonfunctional. The larvae feed on seeds of the conifer *Agathis*. According to morphology, Micropterigidae branch off first, and Agathiphagidae branch off second, from the remaining Lepidoptera. This phylogeny is compatible with stepwise progression from an ancestral ground-dwelling, nonphytophagous larva, to arboreal feeding on gymnosperms, and from thence onto angiosperms (62). An early molecular study strongly agreed (138), but multigene studies sometimes group micropterigids with agathiphagids, complicating the ecological interpretation (63, 109, 111). Transcriptomics strongly reasserts the hypothesis of a basal divergence between Micropterigidae and all others, but the question is not fully settled.

Within Angiospermivora, there is strong evidence for a clade Glossata, characterized by acquisition of the typical adult proboscis, that excludes only Heterobathmiidae. The morphological hypothesis (65, 66) postulates three nested basal divergences within Glossata, giving rise to Coelolepida (hollow wing scales), Myoglossata (proboscis with intrinsic musculature), and Neolepidoptera (musculate, crochet-bearing larval abdominal prolegs). These putative clades, however, are strongly contradicted by molecular evidence, implying homoplasy in the early history of their defining traits (9, 64, 109). In contrast, two other nested subclades within Glossata have strong molecular support: Heteroneura (differing venation in forewings versus hindwings, frenular wing coupling) and Eulepidoptera (origin of the pilifers, bristle-bearing organs at the proboscis base, and an advanced mechanism locking the proboscis halves). Molecular evidence further supports a clade Euheteroneura (109), containing Ditrysia plus Tischerioidea and Palaephatoidea. Finally, phylogenomic analysis resolves a subset of South American Palaephatidae as the sister group to Ditrysia, rendering palaephatoids paraphyletic (9).

Overview of Ditrysia

Delimiting subgroups is more difficult in Ditrysia than in nonditrysians. Three nested major subdivisions are now recognized (**Figure 1**). The group Apoditrysia was proposed for all Ditrysia



except Tineoidea, Yponomeutoidea, Gracillarioidea, and Gelechioidea (83, 84). Subsequently, Gelechioidea were shown to belong within Apoditrysia (8, 49, 53, 93, 111). Within Apoditrysia, Minet (83) proposed a group Obtectomera, defined initially by a pupa with fused abdominal segments but later modified to accommodate other partially conflicting characters (84). Molecular evidence strongly supports a revised Obtectomera that includes Gelechioidea (8, 53, 93, 111). Finally, within Obtectomera there is robust molecular support (8, 53, 93, 111) for a group Macroheterocera (big moths) that includes such familiar taxa as Geometroidea, Noctuoidea, and Bombycoidea.

Early-Diverging Ditrysia: The Nonapoditrysians

Molecular data (8, 53, 93, 108, 111) now strongly confirm the proposed monophyly of Ditrysia apart from Tineoidea (84) but argue strongly against monophyly of Tineoidea, which is defended by a single putative synapomorphy (24). Morphology analyzed alone and in combination with multigene data supports tineoid monophyly, but very weakly (45). In contrast, in multigene and transcriptomic analyses excluding all or most synonymous change, tineoids are paraphyletic with respect to the remaining Ditrysia (93, 108, 111). Tineoid monophyly, seen when synonymous change is included (108, 111), is an artifact of base composition convergence (111). The strong molecular evidence appears to outweigh the morphological evidence, and the superfamily classification will probably need to change. The newly recognized family Meessiidae (108), formerly included in Tineidae, appears to be the earliest-branching ditrysian lineage known (but see below), followed by Psychidae (bagworm moths). Either Tineidae sensu stricto + Dryadaulidae or these families plus Eriocottidae appear to be sister group to the remaining Ditrysia. Finally, there is strong molecular evidence that the earliest-diverging lineage of nontineoid ditrysians is Yponomeutoidea + Gracillarioidea (93, 111), which in turn are sister group to Apoditrysia. A majority of tineoid species feed on detritus and/or fungi, rather than on living plants. Given that

Figure 1

Estimated phylogeny of superfamilies of Lepidoptera, synthesized from multiple sources. Thicker lines denote more securely established groupings, and parentheses enclose number of families/species. Classification and species counts follow data from Reference 127 except as noted in the text. Panels provide representative images for major lepidopteran clades. Format for panel legends: major group name (and common name if available); approximate (forewing) length where available; image author and/or source; license code. (a) Trichoptera (caddis fly); D. Hobern, Wikipedia (W); CCA 2.0G. (b) Micropterigoidea (micropterigid moth); 3.5 mm; M. Kurz, W; CCA 2.0G. (c) Eriocranioidea (eriocraniid moth); 5 mm; Sydmolen; CCA 2.5G. (d) Eriocranioidea (eriocraniid larva); <5 mm; Charley Eiseman. (e) Adeloidea (adelid moth); 10 mm; Svdmolen; CCA 2.5G. (f) Tineoidea (grass tubeworm moth); 10 mm; A. Reago and C. McClarrene; CCA 2.0G. (g) Tineoidea (bagworm moth); B. Dupont; CCA 2.0G. (b) Yponomeutoidea (plutellid moth); 7 mm; D. Hobern; CCA 2.0G. (i) Yponomeutoidea (diamondback moth larva); M. Shepard, G.R. Carner, and P.A.C. Ooi, Insects and their Natural Enemies Associated with Vegetables and Soybean in S.E. Asia, Bugwood Network (BW); CCA 3.0. (1) Galacticoidea (mimosa webworm); 10 mm; Ash, W; CCA 2.0G. (k) Tortricoidea (tortricid moth); 14 mm; G. Wise, W; CCA 2.0G. (l) Tortricoidea (tortricid larva); 7 mm; G. Csoka, Hungary Forest Research Institute, BW. (m) Zygaenoidea (flannel moth); 13 mm; gailhampshire; CCA 2.0G. (n) Cossoidea (carpenter worm); 30 mm; J. Solomon, USDA Forest Service, BW; CCA 3.0. (o) Papilionoidea (western tiger swallowtail); Calibas; public domain (PD). (p) Papilionoidea (monarch butterfly); USDA Forest Service; PD. (q) Gelechioidea (pink bollworm); P. Greb, USDA Agricultural Research Service; BW; CCA 3.0. (r) Gelechioidea (gelechiid moth); D. Hobern, W; CCA 2.0G. (s) Pterophoroidea (plume moth); F. Welter-Schultes, W; CC0 1.0 PD. (t) Pyraloidea (European corn borer); 13 mm; © entomart; PD. (u) Pyraloidea (European corn borer); Clemson University USDA Cooperative Extension Slide Series, BW; CCA 3.0. (v) Geometroidea (inchworm); 20 mm; Sydmolen, W; CCA 2.5G. (w) Geometroidea (inchworm moth); 12 mm; © entomart; PD. (x) Noctuoidea (corn earworm); 25 mm; USGS Bee Inventory and Monitoring Lab; CCA 2.0G. (y) Noctuoidea (Old World bollworm); 20 mm; D. Hobern; CCA 2.0G. (z) Bombycoidea (privet hawkmoth); 85 mm; © entomart; PD. (aa) Bombycoidea (cecropia moth); 80 mm; E.L. Manigault, Clemson University Donated Collection, BW; CCA 3.0. Key to license codes: CC = Creative Commons; A = Attribution (only); number (e.g., 2.0) = version; G = generic.

tineoids appear to represent several successive basal divergences in Ditrysia, the latter may have gone through an initial evolutionary phase in which saprophagy/mycophagy were dominant, prior to reradiating on living angiosperms (108).

Early-Diverging Apoditrysia: The Nonobtectomerans

The boundary between obtectomeran and nonobtectomeran Apoditrysia is still being clarified, primarily by molecular data. Several groups not traditionally considered Obtectomera appear to belong there, including Gelechioidea, Alucitoidea, and Epermeniidae. In contrast, some presumptive obtectomerans, such as Immidae (66), no longer appear to belong to that clade (8, 53).

The paraphyletic nonobtectomeran Apoditrysia, approximately nine superfamilies and 25 families (~16,500 species), have proven exceptionally difficult to resolve. From morphology, only two groupings of superfamilies have been postulated: (a) the pairing of Cossoidea + Sesioidea (sometimes treated as a single superfamily) (127) and (b) the grouping of Cossoidea/Sesioidea together with Zygaenoidea (84, 117) to form what we informally term the CSZ complex. Monophyly of the CSZ complex has been difficult to assess in molecular studies, in part because the two highly divergent ectoparasitic families of Zygaenoidea are so unstable in position (8, 111). If these are excluded, however, the grouping has strong molecular support (53, 111). In contrast, morphological analyses (45) do not support monophyly. The CSZ complex is probably the sister group to Obtectomera (53, 111), though more evidence is needed. Essentially no other relationships among lower apoditrysian superfamilies have been definitively established, though transcriptomic data are showing promise (8, 53).

Relationships within Obtectomera

The most securely established large clade (~88,000 species) within Obtectomera, supported by all molecular studies to date, consists of Pyraloidea plus Macroheterocera (here taken to include Mimallonoidea) (8, 53). We refer to these informally as the eared moths, because adults of most macroheterocerans, and nearly all pyraloids, bear a pair of lateral tympanic organs somewhere near the boundary between the metathorax and abdomen. Similar structures are rare elsewhere in Lepidoptera, although other types of ultrasound detectors may be widespread (22, 87). Butterflies, formerly placed with big moths in Macrolepidoptera (65, 66), are now decisively excluded from this clade.

Phylogenomics is gradually clarifying macroheteroceran relationships. Mimallonidae (the sack-bearers) appear to be sister group to the remaining macroheterocerans (8, 53, 126). The next split is between Drepanoidea sensu lato (127) and Geometroidea + Noctuoidea + Bombycoidea (8, 126). Within the latter triplet, one study supported Geometroidea + Noctuoidea (53), but another favored Geometroidea + Bombycoidea (8). Morphological characters may favor the latter pairing (104), which is also consistently found in mitogenomic studies (57, 60, 140).

Relationships among the other nine obtectomeran superfamilies remain problematic (93, 111). Initial transcriptomic analyses appeared to provide strong resolution but included few superfamilies (8, 53) and were sometimes in conflict. In the absence of more complete taxon sampling, strong conclusions cannot be drawn.

PROGRESS ON THE MAJOR LEPIDOPTERAN SUPERFAMILIES

Each of the superfamilies containing $\geq 1,500$ species (as well as several smaller ones) (114) has now been subject to one or more molecular studies that attempt to span its diversity. We here

summarize advances on these major superfamilies, focusing mainly on the family level. Progress is also rapid, however, at the subfamily level and below.

Tineoidea: Clothes Moths, Bagworm Moths, and Relatives

Recent studies (93, 108, 111) have altered the previous classification of tineoid families (24). Tineidae have proven strongly monophyletic, but only after inclusion of Acrolophidae plus removal of three aberrant genera (93, 108). Psychidae (1,350 species) are securely monophyletic, but only after inclusion of Arrhenophanidae (93, 108, 111). Relationships among the 11 psychid subfamilies are incompletely studied, but results so far (108, 113) support a basal divergence between psychine and arrhenophanine lineages. Tineidae sensu stricto are split basally into an acrolophine versus a tineine lineage, but as yet, there is almost no further resolution (108). Approximately 13% of the \sim 2,200 described species of Tineidae are unplaced to subfamily, and two of the largest previous subfamilies (24) are demonstrably polyphyletic (108). Multiple new deep ditrysian lineages may await discovery.

Yponomeutoidea: Ermine Moths and Relatives

Yponomeutoidea, which include approximately 1,800 species of small- to medium-sized moths, have long been problematic. Building on the work of Kyrki and others (29, 67, 68), a molecular study (122) provided moderate to strong support for yponomeutoid monophyly and recognized 10 families. Some relationships among families were strongly supported but not all, and further work is needed. Yponomeutoidea are one of the earliest lepidopteran groups to include both internal and external feeders and to extensively colonize herbaceous as well as woody plants. The diamondback moth (Yponomeutoidea: Plutellidae) is the first genome-sequenced nonobtectomeran (143).

Gracillarioidea: Leaf-Mining Moths

Gracillarioidea formerly included Bucculatricidae, Douglasiidae, Gracillariidae, and Roeslerstammiidae (24). Molecular evidence now removes Douglasiidae to the nonobtectomeran Apoditrysia (55, 93, 111, 122). The remaining families plus Yponomeutoidea form a robust monophyletic group, but Gracillarioidea are probably paraphyletic with respect to Yponomeutoidea. Gracillaridae (~1,900 species, wingspan 4–20 mm), mostly arboreal feeders, are the largest predominantly leaf-mining lepidopteran clade (71). Nearly all species undergo larval hypermetamorphosis (i.e., a dramatic shift in morphology upon switching from fluid feeding to tissue feeding) (21). Recent multigene studies (55, 56) robustly support eight subfamilies comprising three larger clades.

Tortricoidea: Leaf-Roller Moths and Relatives

The larvae of Tortricoidea (>10,800 species, all in Tortricidae) (41, 48) are mostly either external-feeding leaf tiers or borers in various plant parts. There are three subfamilies and 20 tribes (41, 48). A multigene study (105) strongly supported both individual monophyly of and a sister-group relationship between the two large subfamilies Tortricinae and Olethreutinae. Most tribes and relationships among them were well supported, although two tribes proved paraphyletic. Tribes vary markedly in diet breadth, correlated with internal versus external feeding and with deposition of eggs singly versus in large clusters (105).

Zygaenoidea

The definition and internal phylogeny of Zygaenoidea (currently ~3,300 species in 12 families) remain in flux (34, 35, 83, 127, 142). Zygaenoid larvae are mostly exposed external folivores, which are rare outside Obtectomera, but the small families Cyclotornidae and Epipyropidae are at least initially ectoparasites on auchenorrhynchan Hemiptera. Zygaenoid larvae are typically slow growing and heavily defended and are often aposematic, as are the adults of some zygaenoid groups (35).

A morphological analysis treating five families (34) supported a limacodid group consisting of Limacodidae, Dalceridae, Aididae, and Megalopygidae, but excluding Zygaenidae. Molecular studies with additional taxa have begun to fill out the picture (8, 93, 111). Molecular evidence strongly supports a core Zygaenoidea including all except the two ectoparasitic families, basally divided into limacodid-group versus zygaenid-group families. Most but not all family definitions and relationships within these two groups are strongly supported. In molecular analyses, the ectoparasitic families are long branched and unstably placed, never grouping with other zygaenoids. In analyses including morphology (45), however, weak support joins them with the limacodid group. Overall the evidence favors the molecular result because of its very strong support, but the conflict appears genuine.

Cossoidea + Sesioidea (Cossoidea Sensu Lato)

Monophyly for this putative clade, defended by two potential synapomorphies (32, 84), is uncertain. It is supported by some but not all multigene analyses (17, 93, 111), sometimes with inclusion of the parasitic zygaenoids, and usually with low support except when the latter are omitted. It was also supported, weakly, in analyses including morphology (45). Larvae in these groups are typically stem borers. Monophyly of each superfamily individually also remains uncertain. Four synapomorphies have been proposed for Sesioidea (32, 83, 84) but only one, variably expressed, for Cossoidea (32, 83), and none for Cossidae (32). Analyses including morphology weakly support monophyly for each superfamily (45). In contrast, in molecules-only analyses, the two are invariably intermingled (93, 111), sometimes with strong support (8), and even the family Cossidae is not monophyletic. Further work is needed.

Gelechioidea

Gelechioidea (~18,400 species, plus many undescribed; wingspan typically <10 mm) remain poorly known and may eventually prove to be the most species-rich superfamily. Gelechioid larvae are mostly host specific on living plants and are typically concealed external feeders, though some are leaf miners or borers (50). There have also been unusually numerous shifts to saprophagy/fungivory (50, 123).

Both morphological and molecular data (43, 45, 123) support monophyly of Gelechioidea, but the circumscription of and relationships among families remain problematic. Two morphological studies (46, 49) reached very different conclusions. The first multigene analyses (50, 111) differed substantially from both of these results, in part owing to very low support values. Somewhat stronger groupings were seen in a combined morphological/molecular analysis by Heikkilä et al. (43), who recognized 16 families. An independent molecular analysis (123) found a different rooting but otherwise similar branching structure. If the former tree (43) is rerooted following the latter (123), the two studies both support tentative division of the superfamily into three large assemblages, with similar internal relationships. Larval ecologies show strongly clumped

distributions on this phylogeny. Saprophagy is mostly limited to one subclade of the depressariid assemblage, whereas internal feeding is almost entirely restricted to the other two assemblages. Although progress has been made, a robust phylogeny of the families will require much further work.

Papilionoidea: Butterflies

Papilionoidea (18,768 species) are the largest clade of Lepidoptera in which adults are almost always diurnal (though diurnality has arisen repeatedly in other superfamilies). Most butterfly larvae are exposed external folivores, though some are leaf rollers or stem borers. Molecular evidence has markedly changed our understanding of butterfly relationships. There is now strong support for inclusion here of Hedylidae (2, 116), formerly placed in Geometridae. Papilionoidea as formerly defined (2), excluding Hesperiidae (skippers) and Hedylidae, are paraphyletic, because the Papilionidae are sister group to all remaining butterflies sensu lato (44, 53, 93, 111). Intensive study of among-family relationships supports the following phylogeny, expressed in standard parenthetical notation in which each pair of left and right parentheses at the same level of nesting encloses a monophyletic group: (Papilionidae, ((Hedylidae, Hesperiidae), (Pieridae, ((Lycaenidae, Riodinidae), Nymphalidae)))). There are now multiple phylogenetic studies on the individual families as well (36, 51, 119, 128, 129, 132); Lycaenidae are probably least well understood. A recently funded project (75) seeks to produce a molecular phylogeny for all butterfly species. Butterflies, in which phylogenetics is especially advanced, have been the model group for most recent applications of phylogeny to broader questions in lepidopteran evolution (see below).

Pyraloidea: Snout Moths, Grass Moths, and Relatives

Pyraloidea (15,576 species, with many more undescribed) (96, 127) are typically small- to mediumsized moths. Most feed as larvae on living plants, as concealed external folivores, or as stem borers. However, pyraloids are exceptionally ecologically diverse and also include detritivores, coprophages, predators, and parasites as well as aquatic herbivores.

Monophyly for Pyraloidea sensu stricto (81, 82) is supported by morphological analyses (45) and by all molecular studies to date (93, 110, 112). The chief synapomorphy, seen in nearly all species, is a pair of unique ventral tympanal organs on the second abdominal segment (92). Minet (81, 82) divided pyraloids into two families, Pyralidae and Crambidae, citing differences in their tympanal organs. Molecular evidence strongly supports the monophyly of each (110). Relationships among the five subfamilies of Pyralidae (5,921 species) are strongly resolved by molecular and morphological analyses (110, 124), except that Chrysauginae are poorly understood and might even include the earliest-diverging lineages in the family (110). In Crambidae, an initial molecular study (110) strongly resolved many relationships among the 21 subfamilies, including a clade associated with mustard oil plants and another associated with wet habitats. A smaller mitogenomic study on pyraloids yielded concordant results (141). Multiple nodes remain problematic, however, and further work is needed.

Geometroidea: Inchworms and Allies

Geometroidea previously included Geometridae, Uraniidae, and Sematuridae (86). Molecular evidence (8, 111) strongly favors addition of both Epicopeiidae, previously in Drepanoidea (86), and the new family Pseudobistonidae (104). Geometroids are typically medium-sized to large moths (wingspans 12–120 mm). The larvae are exposed leaf feeders, typically on woody plants.

Within geometroids, the preponderance of recent evidence (8, 104, 111) favors the following relationships: ((Uraniidae, Geometridae), (Sematuridae, (Epicopeiidae, Pseudobistonidae))). Owing to sparse sampling and conflicting results, however (53, 118), this arrangement is not definitive. The provisional phylogeny parallels the distribution of paired tympanic organs. Geometridae bear unique tympanic organs on the base of the abdomen. Uraniidae possess another unique form of this organ, differing in morphology and location between the sexes. The other families lack tympanal organs. Molecular evidence has now strongly resolved most relationships among the eight subfamilies of Geometridae (24,000 species) (1, 111, 118, 139, 144). Multigene studies also strongly resolve relationships among the three subfamilies of Uraniidae (111, 118), concordant with an earlier morphological hypothesis (72).

Bombycoid Complex: Silkworms and Allies

Minet (85) recognized a bombycoid complex consisting of Bombycoidea, Lasiocampoidea, and Mimallonoidea. Subsequent analyses, cited above, now place Mimallonoidea at or near the base of Macroheterocera, but multigene and especially transcriptomic data strongly group Lasiocampoidea (Lasiocampidae) with Bombycoidea (8, 45, 111). Within this clade, molecular evidence strongly resolves most relationships among the ten bombycoid families (11, 106, 150, 151), which include Anthelidae, formerly placed in Lasiocampoidea (85). Extensive phylogenetic information now exists on the two largest families, Saturniidae (wild silk moths) and Sphingidae (hawkmoths) (7, 52, 54, 107).

Noctuoidea: Cutworms and Allies

Noctuoidea (~42,000 species) is by far the largest lepidopteran superfamily. The adults are typically medium to large in size. Noctuoid monophyly is strongly supported by molecular evidence (93, 111, 112, 146), though (surprisingly) not by morphological analysis alone (45). Unexpectedly, molecular evidence very strongly assigns the formerly noctuoid family Doidae (59) to Drepanoidea (8, 93, 111), implying a dual origin for the characteristic noctuoid pair of tympanal organs on the last thoracic segment. Noctuoid larvae are typically exposed leaf feeders, though some groups bore in stems, flowers, or fruits, and some live on or below the soil.

Understanding of the relationships within Noctuoidea has changed substantially in recent years. Molecular data strongly establish that Noctuidae in the former broad sense (59) are paraphyletic with respect to the former Arctiidae and Lymantriidae (89, 133, 146). There is now general agreement on a system of six monophyletic families (146) consisting of the earlier-diverging Oenosandridae (8 species) and Notodontidae (3,800 species) plus a clade comprising Erebidae (24,569 species), subsuming lymantriids and arctiids; Noctuidae sensu stricto (11,772 species; the trifine noctuids) (59); Nolidae (1,738 species); and Euteliidae (520 species). Oenosandridae were argued on morphological grounds to be sister group to the remaining noctuoids (79), but molecular evidence remains inconsistent, sometimes strongly grouping oenosandrids with Notodontidae (80, 93, 111, 140, 146). Relationships among the remaining families are also unclear. The strongest evidence to date, from mitogenomics (140), favors the hypothesis (Erebidae, (Nolidae, (Euteliidae, Noctuidae))), contradicting the most robust multigene result (148). Initial phylogenetic analyses have been carried out on all families except Euteliidae (70, 80, 102, 145, 147–149), but countless questions remain.

THE TIMESCALE OF LEPIDOPTERAN PHYLOGENY

The fossil record provides only the barest outline of lepidopteran history, much of which must therefore be inferred by comparison among extant species. The oldest unambiguously lepidopteran fossil (136) is from the early Jurassic (193 Mya) and is approximately as old as the earliest trichopteran fossil (42). The earliest evidence for any subdivision of extant Lepidoptera is a much younger, lower Cretaceous fossil belonging to Glossata (125 Mya) (42). Therefore, Micropterigoidea, Agathiphagoidea, and Heterobathmioidea must have already diverged by then, though the earliest fossil micropterigid is somewhat younger (111 Mya) (135). The first reported fossil of Ditrysia is a leaf mine from the mid-Cretaceous (103 Mya) (69), assigned to a modern subfamily of Gracillariidae, though reservations about this interpretation have been expressed (42). No further securely identified fossils of Ditrysia occur until extant families start to appear in the late Paleocene (56 Mya) (42).

The lepidopteran fossil record is very poorly studied and appears to be much sparser than that of other comparably diverse insect orders (18, 42, 66, 120). The fragility and buoyancy of the scale-covered bodies and wings of Lepidoptera probably impede their preservation in lake-bed deposits (30), making compression fossils rarer than amber inclusions and trace fossils such as leaf mines. Scales and wings often obscure diagnostic features in fossils, and diagnostic features themselves are scarce even for extant lineages of Ditrysia (45). Very few lepidopteran fossils have explicitly synapomorphy-based identifications, and almost no molecular datings for lepidopteran groups have adhered strictly to the best practice (98) of using only such fossils as calibration points (25). Fortunately, lepidopteran fossils have now been extensively cataloged (27, 121), and many are being critically reexamined (26).

Despite limited fossil evidence, there have now been many molecular datings within Lepidoptera that use one or more fossils and/or other calibrations based on geography or host-plant age. Moreover, multiple dating studies have tied the origin or diversification of particular lepidopteran clades to specific Earth-history events, a potentially important advance (20, 99, 131). Most though not all divergence datings (52, 114, 139) have focused on butterflies (36, 40, 44, 94, 119). Results have varied, but the inferred ages for Papilionoidea and constituent families have typically been 140–80 Mya (lower to middle Cretaceous), two to three times older than their oldest fossils (Cenozoic; ~56–30 Mya) (42).

Several authors have examined broader divergences, with somewhat conflicting results. A multigene study across Lepidoptera (131) dated the first split between extant lineages at 215 Mya (late Triassic), the origin of Ditrysia at ~160 Mya (late Jurassic), and the origin of Apoditrysia at ~140 Mya (early Cretaceous), approximately twice as old, on average, as the corresponding oldest fossils. In contrast, a phylogenomic analysis across all insect orders (88) that included seven extant and three fossil lepidopterans yielded much younger ages of ~140 Mya, ~100 Mya, and ~80 Mya, respectively. This study (88) also estimated the origin of Lepidoptera at ~245 Mya, whereas a multigene analysis across hexapods (19) placed that origin at 270 Mya. Given the multicausal variability in divergence-time estimates (28, 40, 137) and the paucity of secure calibrations (25, 120), some caution is warranted about the conclusions drawn thus far. As the array of rigorously identified fossils expands, however, and dating methods become more sophisticated and better understood, divergence datings in Lepidoptera should become increasingly accurate and precise.

TOWARD UNDERSTANDING LEPIDOPTERAN EVOLUTION

Progress on phylogenetics is helping to answer long-standing questions about lepidopteran evolution. Space permits only a brief sketch of some promising directions. Phylogenetic studies of host-plant use have shown an overall trend across Lepidoptera (with numerous exceptions) from internal feeding (most nonditrysians) to concealed external feeding (most nonmacroheteroceran ditrysians) to fully exposed folivory (butterflies and macro-moths) (77, 103), associated with

increase in body size. The reasons for this and other trends, such as the repeated switch to detritivory/fungivory, remain unexplored. Conserved association with particular plant lineages, the basis for coevolution sensu Ehrlich & Raven (33), has proven to be pervasive though not universal across surprisingly broad plant and lepidopteran taxonomic scales (77). Strong phylogenetic and genetic/biochemical evidence is emerging for escape and radiation coevolution in some groups of butterflies (31, 39). How widely it may apply elsewhere is unclear, and some studies in other groups appear to rule it out (76). Attention has also begun to focus on the evolutionary causes and consequences of phylogenetic change in host range (15, 97). Although the emphasis on herbivory still dominates studies of lepidopteran evolution, phylogenies are starting to elucidate evolutionary patterns in several modes of defense against predators, which may be equally important and complex (7, 52, 149).

Phylogenies are also helping to trace the evolution of key lepidopteran traits that have been previously studied mainly from genetic/developmental and biochemical perspectives. Analysis of the evolution of wing-pattern formation in butterflies, a prominent model system in evo-devo (91), is being extended to other major lineages (6, 115). Similarly, phylogenies are also helping to reconstruct the biochemical diversification of lepidopteran long-distance mating pheromones (74). For example, the use of type I pheromones (long straight-chain alcohols and their mainly acetate and aldehyde derivatives) versus type II pheromones (long-chain polyunsaturated hydrocarbons and their epoxy derivatives), variable among subfamilies of the former Noctuidae sensu lato (5, 78), is more conserved than had previously appeared. Type I subfamilies belong to Noctuidae sensu stricto, whereas type II subfamilies are now placed in Erebidae (146). The latter subsumes the former Arctiidae and Lymantriidae, the only other noctuoids in which type II pheromones are known (78). Countless other lines of investigation could also be cited. Phylogeny-enabled study of lepidopteran evolution is a limitless enterprise for the future.

DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS

The progress we summarize is the work of a highly collaborative global research community that includes amateur as well as professional lepidopterists. Countless people have made important contributions, and we apologize to the many whose work we have cited sparsely or not at all owing to lack of space. We are very grateful to the numerous collaborators and other colleagues who have helped us in our own work, including especially our partners in the Leptree project, which grew out of our long-term collaboration with Jerome Regier. We thank Charley Eiseman for providing the eriocraniid larva image. The work described here was supported in part by US National Science Foundation grants DBI-0755048, DEB-1355028, DEB-0531769, and DEB-1355023 and by NIFA project MD-ENTO-0907. We dedicate this paper to the memory of N.P. Kristensen and E.S. Nielsen, pioneers in the modern study of lepidopteran phylogeny.

LITERATURE CITED

Abraham D, Ryrholm N, Wittzell H, Holloway JD, Scoble MJ, Lofstedt C. 2001. Molecular phylogeny
of the subfamilies in Geometridae (Geometroidea: Lepidoptera). Mol. Phylogenet. Evol. 20:65–77

- Ackery PR, de Jong R, Vane-Wright RI. 1998. The butterflies: Hedyloidea, Hesperioidea, and Papilionoidea. See Ref. 63, pp. 264–300
- Ackery PR, Vane-Wright RI. 1984. Milkweed Butterflies, Their Cladistics and Biology: Being an Account of the Natural History of the Danainae, Subfamily of the Lepidoptera, Nymphalidae. London: Br. Mus. Nat. Hist.
- Adamski D, Brown RL. 1989. Morphology and systematics of North American Blastobasidae (Lepidoptera: Gelechioidea). Tech. Bull. 165, Miss. Agric. For. Exp. Stn., Miss. State
- 5. Ando T, Inomata S, Yamamoto M. 2004. Lepidopteran sex pheromones. Top. Curr. Chem. 239:51-96
- Baixeras J. 2002. An overview of genus-level taxonomic problems surrounding Argyroploce Hübner (Lepidoptera: Tortricidae), with description of a new species. Ann. Entomol. Soc. Am. 95:422–31
- Barber J, Leavell BC, Keener AL, Breinholt JW, Chadwell BA, et al. 2015. Moth tails divert bat attack: evolution of acoustic deflection. PNAS 112:2812–16
- Bazinet AL, Cummings MP, Mitter KT, Mitter C. 2013. Can RNA-Seq resolve the rapid radiation of advanced moths and butterflies (Hexapoda: Lepidoptera: Apoditrysia)? An exploratory study. PLOS ONE 8:e82615
- Bazinet AL, Mitter KT, Davis DR, van Nieukerken EJ, Cummings MP, Mitter C. 2016. Phylotranscriptomics resolves ancient divergences in the Lepidoptera. Syst. Entomol. In press
- Breinholt JW, Cason CE, Lemmon AR, Lemmon EM, Xiao L, Kawahara AY. 2016. Anchored hybrid enrichment in Lepidoptera: leveraging genomic data for studies on the megadiverse butterflies and moths. Syst. Biol. In press
- Breinholt JW, Kawahara AY. 2013. Phylotranscriptomics: Saturated third codon positions radically influence the estimation of trees based on next-gen data. Genome Biol. Evol. 5:2082–92
- Brower AVZ. 1994. Phylogeny of Heliconius butterflies inferred from mitochondrial DNA sequences. Mol. Phylogenet. Evol. 3:159–74
- Brower AVZ, DeSalle R. 1998. Mitochondrial versus nuclear DNA sequence evolution among nymphalid butterflies: the utility of Wingless as a source of characters for phylogenetic inference. Insect Mol. Biol. 7:1–10
- Brown JM, Pellmyr O, Thompson JN, Harrison RG. 1994. Mitochondrial DNA phylogeny of the Prodoxidae (Lepidoptera: Incurvarioidea) indicates a rapid ecological diversification of the yucca moths. Ann. Entomol. Soc. Am. 87:795–802
- Cho S, Mitchell A, Mitter C, Regier J, Matthews M, Robertson R. 2008. Molecular phylogenetics of heliothine moths (Lepidoptera: Noctuidae: Heliothinae), with comments on the evolution of host range and pest status. Syst. Entomol. 49:581–94
- Cho S, Mitchell A, Regier JC, Mitter C, Poole RW, et al. 1995. A highly conserved nuclear gene for low-level phylogenetics: elongation factor-1α recovers morphology-based tree for heliothine moths. Mol. Biol. Evol. 12:650–56
- Cho S, Zwick A, Regier JC, Mitter C, Cummings MP, et al. 2011. Can deliberately incomplete gene sample augmentation improve a phylogeny estimate for the advanced moths and butterflies (Hexapoda: Lepidoptera)? Syst. Biol. 60:782–96
- 18. Common IFB. 1975. Evolution and classification of the Lepidoptera. Annu. Rev. Entomol. 20:183-203
- Condamine FL, Clapham ME, Kergoat GJ. 2016. Global patterns of insect diversification: towards a reconciliation of fossil and molecular evidence? Sci. Rep. 6:19208
- Condamine FL, Sperling FAH, Wahlberg N, Rasplus JY, Kergoat GJ. 2012. What causes latitudinal gradients in species diversity? Evolutionary processes and ecological constraints on swallowtail biodiversity. Ecol. Lett. 15:267–77
- Davis DR. 1987. Gracillariidae. In *Immature Insects*, Vol. 1, ed. FW Stehr, pp. 372–78. Dubuque, IA: Kendall/Hunt
- Davis DR. 1998. A world classification of the Harmacloninae, a new subfamily of Tineidae (Lepidoptera: Tineoidea). Smithson. Contrib. Zool. 597:1–81
- Davis DR, Gentili P. 2003. Andesianidae, a new family of monotrysian moths (Lepidoptera: Andesianoidea) from South America. *Invertebr. Syst.* 17:15–26
- 24. Davis DR, Robinson GS. 1998. The Tineoidea and Gracillarioidea. See Ref. 63, pp. 91-117

- de Jong R. 2007. Estimating time and space in the evolution of the Lepidoptera. Tijdschr. Entomol. 150:319–46
- de Jong R. 2016. Reconstructing a 55-million-year-old butterfly (Lepidoptera: Hesperiidae). Eur. J. Entomol. 113:423–28
- Doorenweerd C, van Nieukerken EJ, Sohn J-C, Labandeira CC. 2015. A revised checklist of Nepticulidae fossils (Lepidoptera) indicates an Early Cretaceous origin. Zootaxa 3963:295–334
- dos Reis M, Donoghue PCJ, Yang Z. 2016. Bayesian molecular clock dating of species divergences in the genomics era. Nat. Rev. Genet. 17:71–80
- Dugdale JS, Kristensen NP, Robinson GS, Scoble MJ. 1998. The Yponomeutoidea. See Ref. 63, pp. 120– 30
- 30. Duncan IJ. 1997. The taphonomy of insects. PhD Thesis, Palaeobiol. Res. Group, Univ. Bristol
- 31. Edger PP, Heidel-Fischer HM, Bekaert M, Rota J, Glöckner G, et al. 2015. The butterfly plant arms-race escalated by gene and genome duplications. *PNAS* 112:8362–66
- Edwards ED, Gentili P, Horak M, Kristensen NP, Nielsen ES. 1998. The cossoid/sesioid assemblage. See Ref. 63, pp. 181–97
- 33. Ehrlich PR, Raven PH. 1964. Butterflies and plants: a study in coevolution. Evolution 18:586-608
- Epstein ME. 1996. Revision and phylogeny of the Limacodid-group families, with evolutionary studies on slug caterpillars (Lepidoptera: Zygaenoidea). Smithson. Contrib. Zool. 582:1–102
- Epstein ME, Geertsema H, Naumann CM, Tarmann GM. 1998. The Zygaenoidea. See Ref. 63, pp. 159– 80
- Espeland M, Hall JPW, DeVries PJ, Lees DC, Cornwall M, et al. 2015. Ancient Neotropical origin
 and recent recolonisation: phylogeny, biogeography and diversification of the Riodinidae (Lepidoptera:
 Papilionoidea). Mol. Phylogenet. Evol. 93:296–306
- Faircloth BC, Branstetter MG, White ND, Brady SG. 2014. Target enrichment of ultraconserved elements from arthropods provides a genomic perspective on relationships among Hymenoptera. Mol. Ecol. Res. 15:489–501
- 38. Fang Q, Cho S, Regier J, Mitter C, Matthews M, et al. 1997. A new nuclear gene for insect phylogenetics: Dopa decarboxylase is informative of relationships within Heliothinae (Lepidoptera: Noctuidae). *Syst. Biol.* 46:269–83
- 39. Fordyce JA. 2010. Host shifts and evolutionary radiations of butterflies. Proc. R. Soc. B 277:3735-43
- Garzón-Orduña IJ, Silva-Brandão KL, Willmott KR, Freitas AV, Brower AV. 2015. Incompatible ages for clearwing butterflies based on alternative secondary calibrations. Syst. Biol. 64:752–67
- Gilligan TM, Baixeras J, Brown JW, Tuck KR. 2014. T@RTS: online world catalogue of the Tortricidae (version 3.0). Tortricid.net: Torticidae Resources on the Web. http://www.tortricid.net/catalogue.asp. Accessed March 26, 2016
- 42. Grimaldi DA, Engel MS. 2005. Evolution of the Insects. Cambridge, UK: Cambridge Univ. Press
- 43. Heikkilä M, Mutanen M, Kekkonen M, Kaila L. 2014. Morphology reinforces proposed molecular phylogenetic affinities: a revised classification for Gelechioidea (Lepidoptera). *Cladistics* 30:563–89
- Heikkilä M, Kaila L, Mutanen M, Peña C, Wahlberg N. 2011. Cretaceous origin and repeated tertiary diversification of the redefined butterflies. Proc. R. Soc. B 279:1093–99
- Heikkilä M, Mutanen M, Wahlberg N, Sihvonen P, Kaila L. 2015. Elusive ditrysian phylogeny: an account of combining systematized morphology with molecular data (Lepidoptera). BMC Evol. Biol. 15:260
- 46. Hodges RW. 1998. The Gelechioidea. See Ref. 63, pp. 131-58
- Holloway JD. 1986–2011. Moths of Borneo part 2. Kuala Lumpur: Malays. Nat. Soc./Southdene Sdn. Bhd.
- 48. Horak M. 1998. The Tortricoidea. See Ref. 63, pp. 199–215
- Kaila L. 2004. Phylogeny of the superfamily Gelechioidea (Lepidoptera: Ditrysia): an exemplar approach. Cladistics 20:303–40
- Kaila L, Mutanen M, Nyman T. 2011. Phylogeny of the mega-diverse Gelechioidea (Lepidoptera): adaptations and determinants of success. Mol. Phylogenet. Evol. 61:801–9
- 51. Kaliszewska ZA, Lohman DJ, Sommer K, Adelson G, Rand DB, et al. 2015. When caterpillars attack: biogeography and life history evolution of the Miletinae (Lepidoptera: Lycaenidae). *Evolution* 69:571–88

- Kawahara AY, Barber JR. 2015. Tempo and mode of anti-bat ultrasound and jamming in the diverse hawkmoth radiation. PNAS 112:6407–12
- Kawahara AY, Breinholt JW. 2014. Phylogenomics provides strong evidence for relationships of butterflies and moths. Proc. R. Soc. B 281:20140970
- Kawahara AY, Mignault AA, Regier JC, Kitching IJ, Mitter C. 2009. Phylogeny and biogeography of hawkmoths (Lepidoptera: Sphingidae): evidence from five nuclear genes. PLOS ONE 4:e5719
- Kawahara AY, Ohshima I, Kawakita A, Regier JC, Mitter C, et al. 2011. Increased gene sampling provides stronger support for higher-level groups within gracillariid leaf mining moths and relatives (Lepidoptera: Gracillariidae). BMC Evol. Biol. 11:182
- 56. Kawahara AY, Plotkin D, Ohshima I, Lopez-Vaamonde C, Houlihan PR, et al. 2016. A molecular phylogeny and revised higher-level classification for the leaf-mining moth family Gracillariidae and its implications for larval host use evolution. Syst. Entomol. In press. https://doi.org/10.1111/syen.12210
- 57. Kim MJ, Kang AR, Jeong HC, Kim KG, Kim I. 2011. Reconstructing intraordinal relationships in Lepidoptera using mitochondrial genome data with the description of two newly sequenced lycaenids, Spindasis takanonis and Protantigius superans (Lepidoptera: Lycaenidae). Mol. Phylogenet. Evol. 61:436–45
- Kitching IJ. 1987. Spectacles and silver Ys: a synthesis of the systematics, cladistics and biology of the Plusiinae (Lepidoptera: Noctuidae). Bull. Br. Mus. Nat. Hist. Entomol. 54:75–261
- 59. Kitching IJ, Rawlins JE. 1998. Noctuoidea. See Ref. 63, pp. 355-401
- Kong W, Yang J. 2015. The complete mitochondrial genome of Rondotia menciana (Lepidoptera: Bombycidae). J. Insect Sci. 15:48
- Kristensen NP. 1976. Remarks on the family-level phylogeny of butterflies (Insecta, Lepidoptera, Rhopalocera). 7. Zool. Syst. Evol. Res. 14:25–33
- Kristensen NP. 1997. Early evolution of the Trichoptera + Lepidoptera lineage: phylogeny and the ecological scenario. Mém. Mus. Natl. Hist. Nat. 173:253–71
- 63. Kristensen NP, ed. 1998. Lepidoptera: Moths and Butterflies, Vol. 1: Evolution, Systematics, and Biogeography. Handb. Zool. 35. Berlin: De Gruyter
- 64. Kristensen NP, Hilton DJ, Kallies A, Milla L, Rota J, et al. 2015. A new extant family of primitive moths from Kangaroo Island, Australia, and its significance for understanding early Lepidoptera evolution. Syst. Entomol. 40:5–16
- Kristensen NP, Scoble MJ, Karsholt OK. 2007. Lepidoptera phylogeny and systematics: the state of inventorying moth and butterfly diversity. Zootaxa 1668:699–747
- 66. Kristensen NP, Skalski AW. 1998. Phylogeny and palaeontology. See Ref. 63, pp. 7-25
- Kyrki J. 1984. The Yponomeutoidea: a reassessment of the superfamily and its suprageneric groups (Lepidoptera). *Insect Syst. Evol.* 15:71–84
- Kyrki J. 1990. Tentative reclassification of holarctic Yponomeutoidea (Lepidoptera). Nota Lepidopterol. 13:28–42
- Labandeira CC, Dilcher DL, Davis DR, Wagner DL. 1994. Ninety-seven million years of angiosperminsect association: paleobiological insights into the meaning of coevolution. PNAS 91:12278–82
- Lafontaine JD. 1993. Cutworm systematics: confusions and solutions. Mem. Entomol. Soc. Can. 165:189–96
- Lees DC, Kawahara AY, Rougerie R, Ohshima I, Kawakita A, et al. 2014. DNA barcoding reveals a largely unknown fauna of Gracillariidae leaf-mining moths in the Neotropics. Mol. Ecol. Resour. 14:286–96
- Lees DC, Smith NG. 1991. Foodplant associations of the Uraniinae and their systematic, evolutionary and ecological significance. J. Lepidopterists Soc. 45:296–347
- Lemmon AR, Emme S, Lemmon EC. 2012. Anchored hybrid enrichment for massively high-throughput phylogenetics. Syst. Biol. 61:721–74
- Löfstedt C, Wahlberg N, Miller JG. 2016. Evolutionary patterns of pheromone diversity in Lepidoptera.
 In Pheromone Communication in Moths: Evolution, Behavior, and Application, ed. JD Allison, RT Cardé, pp. 43–78. Oakland, CA: Univ. California Press
- Lohman D. 2015. Collaborative research: ButterflyNet—an interactive framework for comparative biology. NSF Award Abstr. 1541557, Nat. Sci. Found., Arlington, VA. https://www.nsf.gov/awardsearch/showAward?AWD_ID=1541557

- Lopez-Vaamonde C, Wikström N, Labandeira C, Godfray HCJ, Goodman SJ, Cook JM. 2006. Fossilcalibrated molecular phylogenies reveal that leaf-mining moths radiated millions of years after their host plants. 7. Evol. Biol. 19:1314–26
- Menken SBJ, Boomsma JJ, van Nieukerken EJ. 2010. Large-scale evolutionary patterns of host plant associations in the Lepidoptera. Evolution 64:1098–119
- Millar JG 2000. Polyene hydrocarbons and epoxides: a second major class of lepidopteran sex attractant pheromones. Annu. Rev. Entomol. 45:575–604
- Miller JS. 1987. Phylogenetic studies in the Papilioninae (Lepidoptera: Papilionidae). Bull. Am. Mus. Nat. Hist. 186:365–512
- Miller JS. 1991. Cladistics and classification of the Notodontidae (Lepidoptera: Noctuoidea) based on larval and adult morphology. Bull. Am. Mus. Nat. Hist. 204:1–230
- Minet J. 1982. Les Pyraloidea et leurs principales divisions systématiques. Bull. Soc. Entomol. Fr. 86:262– 80
- Minet J. 1985. Étude morphologique et phylogénétique des organs tympaniques des Pyraloidea. 2— Pyralidae, Crambidae, premiere partie (Lepidoptera Glossata). Ann. Soc. Entomol. Fr. 21:69–86
- 83. Minet J. 1986. Ebauche d'une classification modern de l'ordre des Lepidopteres. Alexanor 14:291-313
- Minet J. 1991. Tentative reconstruction of the ditrysian phylogeny (Lepidoptera: Glossata). Entomol. Scand. 22:69–95
- Minet J. 1994. The Bombycoidea: phylogeny and higher classification (Lepidoptera: Glossata). Entomol. Scand. 25:63–88
- 86. Minet J, Scoble MJ. 1998. The drepanoid/geometroid assemblage. See Ref. 63, pp. 301-20
- 87. Minet J, Surlykke A. 2003. Auditory and sound producing organs. In *Lepidoptera: Moths and Butterflies*, Vol. 2: *Morphology and Physiology*, ed. NP Kristensen, pp. 289–323. Berlin: De Gruyter
- 88. Misof B, Liu S, Meusemann K, Peters RS, Donath A, et al. 2014. Phylogenomics resolves the timing and pattern of insect evolution. *Science* 346:763–67
- 89. Mitchell A, Mitter C, Regier JC. 2006. Systematics and evolution of the cutworm moths (Lepidoptera: Noctuidae): evidence from two protein-coding nuclear genes. Syst. Entomol. 31:21–46
- 90. Mitter C, Silverfine E. 1988. On the systematic position of Catocala Schrank. Syst. Entomol. 13:67-84
- Monteiro A. 2015. The origin, development, and evolution of butterfly eyespots. Annu. Rev. Entomol. 60:253–71
- 92. Munroe EG, Solis MA. 1998. The Pyraloidea. See Ref. 63, pp. 233-56
- Mutanen M, Wahlberg N, Kaila L. 2010. Comprehensive gene and taxon coverage elucidates radiation patterns in moths and butterflies. Proc. R. Soc. B 277:2839–48
- Nazari V, Zakharov EV, Sperling FAH. 2007. Phylogeny, historical biogeography, and taxonomic ranking of Parnassiinae (Lepidoptera, Papilionidae) based on morphology and seven genes. Mol. Phylogenet. Evol. 42:131–56
- Niehuis O, Yen S-H, Naumann CM, Misof B. 2006. Higher phylogeny of zygaenid moths (Insecta: Lepidoptera) inferred from nuclear and mitochondrial sequence data and the evolution of larval cuticular cavities for chemical defence. *Mol. Phylogenet. Evol.* 39:812–29
- Nuss M, Landry B, Vegliante F, Tränkner A, Mally R, et al. 2015. GlobIZ: global information system on Pyraloidea, accessed on March 15, 2016. http://www.pyraloidea.org
- Nylin S, Slove J, Janz N. 2014. Host plant utilization, host range oscillations and diversification in nymphalid butterflies: a phylogenetic investigation. Evolution 68:105–24
- Parham JF, Donoghue PCJ, Bell CJ, Calway TD, Head JJ, et al. 2012. Best practices for justifying fossil calibrations. Syst. Biol. 61:346–59
- Peña C, Wahlberg N. Prehistorical climate change increased diversification of a group of butterflies. Biol. Lett. 2008:274–78
- Penz CM, Freitas AVL, Kaminski LA, Casagrande MM, DeVries PJ. 2013. Adult and early-stage characters of Brassolini contain conflicting phylogenetic signal (Lepidoptera, Nymphalidae). Syst. Entomol. 38:316–33
- Pitkin LM. 1988. The Holarctic genus *Teleiopsis*: host-plants, biogeography and cladistics (Lepidoptera: Gelechiidae). *Entomol. Scand.* 19:143–91

- 102. Poole RW. 1995. *Noctuoidea. Noctuidea (part). Cuculliinae, Stiriinae, Psaphidinae*. Moths Am. North Mex. Ser. Fascicle 26.1. Washington, DC: Wedge Entomol. Res. Found.
- Powell JA, Mitter C, Farrell BD. 1998. Evolution of larval feeding habits in Lepidoptera. See Ref. 63, pp. 403–22
- 104. Rajaei HS, Greve C, Letsch H, Stüning D, Wahlberg N, et al. 2015. Advances in Geometroidea phylogeny, with characterization of a new family based on *Pseudobiston pinratanai* (Lepidoptera, Glossata). Zool. Scr. 44:418–36
- 105. Regier JC, Brown JW, Mitter C, Baixeras J, Cho S, et al. 2012. A molecular phylogeny for the leaf-roller moths (Lepidoptera: Tortricidae) and its implications for classification and life history evolution. PLOS ONE 7:e35574
- 106. Regier JC, Cook CP, Mitter C, Hussey A. 2008. A phylogenetic study of the "bombycoid complex" (Lepidoptera) using five protein-coding nuclear genes, with comments on the problem of macrolepidopteran phylogeny. Syst. Entomol. 33:175–89
- Regier JC, Grant MC, Peigler RS, Mitter C, Cook CP, Rougerie R. 2008. Phylogenetic relationships of wild silkmoths (Lepidoptera: Saturniidae) inferred from four protein-coding nuclear genes. Syst. Entomol. 33:219–28
- 108. Regier JC, Mitter C, Davis DR, Harrison TL, Sohn J-C, et al. 2015. A molecular phylogeny and revised classification for the oldest ditrysian moth lineages (Lepidoptera: Tineoidea), with implications for ancestral feeding habits of the mega-diverse Ditrysia. Syst. Entomol. 40:409–32
- 109. Regier JC, Mitter C, Kristensen NP, Davis DR, van Nieukerken EJ, et al. 2015. A molecular phylogeny for the oldest (non-ditrysian) lineages of extant Lepidoptera, with implications for classification, comparative morphology and life history evolution. Syst. Entomol. 40:671–704
- Regier JC, Mitter C, Solis MA, Hayden JE, Landry B, et al. 2012. A molecular phylogeny for the pyraloid moths (Lepidoptera: Pyraloidea) and its implications for higher-level classification. Syst. Entomol. 37:635– 56
- 111. Regier JC, Mitter C, Zwick A, Bazinet AL, Cummings MP, et al. 2013. A large-scale, higher-level, molecular phylogenetic study of the insect order Lepidoptera (moths and butterflies). PLOS ONE 8:e58568
- 112. Regier JC, Zwick A, Cummings MP, Kawahara AY, Cho S, et al. 2009 Toward reconstructing the evolution of advanced moths and butterflies (Lepidoptera: Ditrysia): an initial molecular study. BMC Evol. Biol. 9:280
- Rhainds M, Davis DR, Price PW. 2009. Bionomics of bagworms (Lepidoptera: Psychidae). Annu. Rev. Entomol. 54:209–26
- 114. Rota J, Peña C, Miller SE. 2016. The importance of long-distance dispersal and establishment events in small insects: historical biogeography of metalmark moths (Lepidoptera, Choreutidae). J. Biogeogr. 43:1254–65
- 115. Schachat SR, Brown RL. 2015. Color pattern on the forewing of Micropterix (Lepidoptera: Micropterigidae): insights into the evolution of wing pattern and wing venation in moths. PLOS ONE 10:e0139972
- Scoble MJ. 1986. The structure and affinities of the Hedyloidea: a new concept of the butterflies. Bull. Br. Mus. Nat. Hist. Entomol. 53:251–86
- 117. Scott JA. 1986. On the monophyly of the Macrolepidoptera, including a reassessment of their relationship to Cossoidea and Castnioidea, and a reassignment of Mimallonidae to Pyraloidea. J. Res. Lepidoptera 25:30–38
- 118. Sihvonen P, Mutanen M, Kaila L, Brehm G, Hausmann A, et al. 2011. Comprehensive molecular sampling yields a robust phylogeny for geometrid moths (Lepidoptera: Geometridae). *PLOS ONE* 6:e20356
- 119. Simonsen TJ, Zakharov EV, Djernaes M, Cotton A, Vane-Wright RI, Sperling FAH. 2011. Phylogeny, host plant associations and divergence time of Papilioninae (Lepidoptera: Papilionidae) inferred from morphology and seven genes with special focus on the enigmatic genera *Teinopalpus* and *Meandrusa*. Cladistics 27:113–37
- Sohn J-C, Labandeira C, Davis DR. 2015. The fossil record and taphonomy of butterflies and moths (Insecta, Lepidoptera): implications for evolutionary diversity and divergence-time estimates. BMC Evol. Biol. 15:12
- Sohn J-C, Labandeira C, Davis DR, Mitter C. 2012. An annotated catalog of fossil and subfossil Lepidoptera (Insecta: Holometabola) of the world. *Zootaxa* 3286:1–132

- 122. Sohn J-C, Regier JC, Mitter C, Adamski D, Landry JF, et al. 2013. A molecular phylogeny for Yponomeutoidea (Insecta, Lepidoptera, Ditrysia) and its implications for classification, biogeography and the evolution of host plant use. PLOS ONE 8:e55066
- 123. Sohn JC, Regier JC, Mitter C, Adamski D, Landry JF, et al. 2015. Phylogeny and feeding trait evolution of the mega-diverse Gelechioidea (Lepidoptera: Obtectomera): new insight from 19 nuclear genes. Syst. Entomol. 41:112–32
- 124. Solis MA, Mitter C. 1992. Review and phylogenetic analysis of the Pyralidae (sensu stricto) (Lepidoptera: Pyralidae) and placement of the Epipaschiinae. Syst. Entomol. 17:79–90
- Sperling FAH. 1993. Mitochondrial DNA phylogeny of the Papilio machaon species group (Lepidoptera: Papilionidae). Mem. Entomol. Soc. Can. 165:233

 –42
- Timmermans MJTN, Lees DC, Simonsen TJ. 2014. Towards a mitogenomic phylogeny of Lepidoptera using next generation sequence technology. Mol. Phylogenet. Evol. 79:169–78
- 127. van Nieukerken EJ, Kaila L, Kitching IJ, Kristensen NP, Lees DC, et al. 2011. Order Lepidoptera Linnaeus, 1758. Zootaxa 3148:212–21
- Wahlberg N, Leneveu J, Kodandaramaiah U, Peña C, Nylin S, et al. 2009. Nymphalid butterflies diversify following near demise at the Cretaceous/Tertiary boundary. Proc. R. Soc. B 276:4295–302
- Wahlberg N, Rota J, Braby MF, Pierce NP, Wheat CW. 2014. Revised systematics and higher classification of pierid butterflies (Lepidoptera: Pieridae) based on molecular data. Zool. Scr. 43:641–50
- Wahlberg N, Wheat CW. 2008. Genomic outposts serve the phylogenomic pioneers: designing novel nuclear markers for genomic DNA extractions of Lepidoptera. Syst. Biol. 57:231–42
- Wahlberg N, Wheat CW, Peña C. 2013. Timing and patterns in the taxonomic diversification of Lepidoptera (butterflies and moths). PLOS ONE 8:e80875
- 132. Warren AD, Ogawa JR, Brower AVZ. 2008. Phylogenetic relationships of subfamilies and circumscription of tribes in the family Hesperiidae (Lepidoptera: Hesperioidea). *Cladistics* 24:1–35
- Weller SJ, Pashley DP, Martin JA, Constable JL. 1994. Phylogeny of noctuoid moths and the utility of combining independent nuclear and mitochondrial genes. Syst. Biol. 43:194–211
- 134. Weller SJ, Friedlander TP, Martin JA, Pashley DP. 1992. Phylogenetic studies of ribosomal RNA variation in higher moths and butterflies (Lepidoptera: Ditrysia). Mol. Phylogenet. Evol. 1:312–37
- 135. Whalley PES. 1978. New taxa of fossil and recent Micropterigidae with a discussion of their evolution and a comment on the evolution of the Lepidoptera (Insecta). *Ann. Transvaal Mus.* 31:72–86
- Whalley PES. 1985. The systematics and paleogeography of the Lower Jurassic insects of Dorset, England. Bull. Br. Mus. Nat. Hist. Geol. 39:107

 –89
- Wheat CW, Wahlberg N. 2013. Critiquing blind dating: the dangers of over-confident date estimates in comparative genomics. Trends Ecol. Evol. 28:636–42
- Wiegmann BM, Mitter C, Regier JC, Friedlander TP, Wagner DM, Nielsen ES. 2000. Nuclear genes resolve Mesozoic-aged divergences in the insect order Lepidoptera. Mol. Phylogenet. Evol. 15:242–59
- 139. Yamamoto S, Sota T. 2007. Phylogeny of the Geometridae and the evolution of winter moths inferred from a simultaneous analysis of mitochondrial and nuclear genes. *Mol. Phylogenet. Evol.* 44:711–23
- 140. Yang X, Cameron SL, Lees DC, Xue D, Han H. 2015. A mitochondrial genome phylogeny of owlet moths (Lepidoptera: Noctuoidea), and examination of the utility of mitochondrial genomes for lepidopteran phylogenetics. *Mol. Phylogenet. Evol.* 85:230–37
- 141. Ye F, Shi Y, Xing L, Yu H, You P. 2013. The complete mitochondrial genome of *Paracymoriza prodigalis* (Leech, 1889) (Lepidoptera), with a preliminary phylogenetic analysis of Pyraloidea. *Aquat. Insects* 35:71–88
- 142. Yen S-H, Robinson GS, Quicke DLJ. 2005. The phylogenetic relationships of Chalcosiinae (Lepidoptera, Zygaenoidea, Zygaenidae). Zool. J. Linn. Soc. 143:161–341
- 143. You M, Yue Z, He W, Yang X, Yang G, et al. 2013. A heterozygous moth genome provides insights into herbivory and detoxification. *Nat. Genet.* 45:220–22
- 144. Young CJ. 2006. Molecular relationships of the Australian Ennominae (Lepidoptera: Geometridae) and implications for the phylogeny of the Geometridae from molecular and morphological data. Zootaxa 1264:1–147
- Zahiri R, Holloway JD, Kitching IJ, Lafontaine D, Mutanen M, Wahlberg N. 2012. Molecular phylogenetics of Erebidae (Lepidoptera, Noctuoidea). Syst. Entomol. 37:102–24

- 146. Zahiri R, Kitching IJ, Lafontaine JD, Mutanen M, Kaila L. 2011. A new molecular phylogeny offers hope for a stable family level classification of the Noctuoidea (Lepidoptera). Zool. Scr. 40:158–73
- 147. Zahiri R, Lafontaine JD, Holloway JD, Kitching IJ, Schmidt BC, et al. 2013. Major lineages of Nolidae (Lepidoptera, Noctuoidea) elucidated by molecular phylogenetics. *Cladistics* 29:337–59
- 148. Zahiri R, Lafontaine JD, Schmidt BC, Holloway JD, Kitching IJ, et al. 2013. Relationships among the basal lineages of Noctuidae (Lepidoptera, Noctuoidea) based on eight gene regions. Zool. Scr. 42:488–507
- Zaspel JM, Weller SJ, Wardwell CT, Zahiri R, Wahlberg N. 2014. Phylogeny and evolution of pharmacophagy in tiger moths (Lepidoptera: Erebidae: Arctiinae). PLOS ONE 9:e101975
- Zwick A. 2008. Molecular phylogeny of Anthelidae and other bombycoid taxa (Lepidoptera: Bombycoidea). Syst. Entomol. 33:190–209
- Zwick A, Regier JC, Mitter C, Cummings MP. 2011. Increased gene sampling yields robust support for higher-level clades within Bombycoidea (Lepidoptera). Syst. Entomol. 36:31–43



Annual Review of Entomology

Volume 62, 2017

Contents

| Following the Yellow Brick Road Charles H. Calisher |
|--|
| Behavioral Sabotage of Plant Defenses by Insect Folivores David E. Dussourd |
| Neuropeptides as Regulators of Behavior in Insects Liliane Schoofs, Arnold De Loof, and Matthias Boris Van Hiel |
| Learning in Insect Pollinators and Herbivores *Patricia L. Jones and Anurag A. Agrawal |
| Insect Pathogenic Fungi: Genomics, Molecular Interactions, and Genetic Improvements Chengshu Wang and Sibao Wang |
| Habitat Management to Suppress Pest Populations: Progress and Prospects Geoff M. Gurr, Steve D. Wratten, Douglas A. Landis, and Minsheng You |
| MicroRNAs and the Evolution of Insect Metamorphosis **Xavier Belles** 111 |
| The Impact of Trap Type and Design Features on Survey and Detection of Bark and Woodboring Beetles and Their Associates: A Review and Meta-Analysis *Jeremy D. Allison and Richard A. Redak** |
| Tephritid Integrative Taxonomy: Where We Are Now, with a Focus on the Resolution of Three Tropical Fruit Fly Species Complexes Mark K. Schutze, Massimiliano Virgilio, Allen Norrbom, and Anthony R. Clarke 147 |
| Emerging Themes in Our Understanding of Species Displacements Yulin Gao and Stuart R. Reitz |
| Diversity of Cuticular Micro- and Nanostructures on Insects: Properties, Functions, and Potential Applications Gregory S. Watson, Jolanta A. Watson, and Bronwen W. Cribb |
| Impacts of Insect Herbivores on Plant Populations **Judith H. Myers and Rana M. Sarfraz** |

| Past, Present, and Future of Integrated Control of Apple Pests: The New Zealand Experience | |
|--|-----|
| James T.S. Walker, David Maxwell Suckling, and C. Howard Wearing | 231 |
| Beekeeping from Antiquity Through the Middle Ages Gene Kritsky | 249 |
| Phylogeny and Evolution of Lepidoptera Charles Mitter, Donald R. Davis, and Michael P. Cummings | 265 |
| The Ambrosia Symbiosis: From Evolutionary Ecology to Practical Management Jiri Hulcr and Lukasz L. Stelinski | 285 |
| Social Life in Arid Environments: The Case Study of Cataglyphis Ants Raphaël Boulay, Serge Aron, Xim Cerdá, Claudie Doums, Paul Graham, Abraham Hefetz, and Thibaud Monnin | |
| Processionary Moths and Associated Urtication Risk: Global Change–Driven Effects Andrea Battisti, Stig Larsson, and Alain Roques | 323 |
| African Horse Sickness Virus: History, Transmission, and Current Status Simon Carpenter, Philip S. Mellor, Assane G. Fall, Claire Garros, and Gert J. Venter | 343 |
| Spatial Self-Organization of Ecosystems: Integrating Multiple Mechanisms of Regular-Pattern Formation Robert M. Pringle and Corina E. Tarnita | 359 |
| Evolution of Stored-Product Entomology: Protecting the World Food Supply David W. Hagstrum and Thomas W. Phillips | 379 |
| Ecoinformatics (Big Data) for Agricultural Entomology: Pitfalls, Progress, and Promise Jay A. Rosenheim and Claudio Gratton | 399 |
| Molecular Evolution of Insect Sociality: An Eco-Evo-Devo Synthesis Amy L. Toth and Sandra M. Rehan | 419 |
| Physicochemical Property Variation in Spider Silk: Ecology, Evolution, and Synthetic Production Sean J. Blamires, Todd A. Blackledge, and I-Min Tso | 443 |
| Indexes | |
| Cumulative Index of Contributing Authors, Volumes 53–62 | 461 |
| Cumulative Index of Article Titles, Volumes 53–62 | 467 |