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HER2-low status as a distinct breast cancer subtype: myth or truth? Analysis of the WSG trials WSG-ADAPT-HR+/HER2-, WSG-PlanB, and WSG-ADAPT-TN



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Abstract

Background New data show that not only HER2-overexpressing breast cancer (BC) tumors but also HER2-low tumors, classically considered as HER2-negative, respond to HER2-targeting antibody–drug-conjugates. Our objective was to analyze the prevalence of HER2-low BC in a pooled analysis of contemporary early BC trials and to evaluate its role as a prognostic factor in terms of survival in comparison to HER2-zero BC.

Methods We evaluated 5598 patients with locally HR+/HER2- BC from the screening cohort of WSG-ADAPT-HR+/HER2-, 2592 patients with HR+/HER2- or HR-/HER2- from the adjuvant WSG-PlanB trial, and 336 patients from the WSG-ADAPT-TN trial. Central HER2 testing was performed prospectively in WSG-ADAPT and retrospectively in WSG-PlanB. Following ASCO/CAP guidelines, HER2-low status was defined as immunohistochemistry (IHC) 1 + or 2 + and in situ hybridization (ISH)-negative, and HER2-zero was defined as IHC 0. Agreement between HER2 assessments was evaluated with Cohen's kappa coefficient, and effects of HER2 status on pathological complete response (pCR) and on survival were analyzed with logistic regression and Cox proportional hazards models, respectively.

Findings In WSG-ADAPT-HR+/HER2-, 3198 (64.6%) tumors were HER2-low by the central and 3096 (55.6%) by the local histology (agreement for HER2-low status was 61.0%). In HR+/HER2- cases from WSG-PlanB, 601 tumors (28.7%) were HER2-low. In both cohorts, HER2-low status was significantly associated with higher *ERBB2* mRNA expression by Oncotype DX test in comparison to HER2-zero: mean 9.3 vs. 9.1 (p < .001) by local HER2 assessment in WSG-ADAPT and mean 9.2 vs. 8.8 (p < .001) in WSG-PlanB. Furthermore, patients with HER2-low tumors in WSG-ADAPT-HR+/HER2- significantly less often had a pCR compared to the HER2-zero tumors (p = .015). No significant difference

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was observed in (invasive and/or distant) disease-free survival (DFS) between centrally HER2-low and HER2-zero tumors in both HR + /HER2- cohorts (WSG-ADAPT-HR + /HER2- distant DFS: unadjusted HR = 1.06, 95%CI 0.83–1.36, similar results for local assessment; WSG-PlanB DFS: unadjusted HR = 1.28, 95%CI 0.91–1.82). In the HR-/HER2- WSG-PlanB cohort, centrally HER2-low tumors (10.5%) were associated with better DFS (unadjusted HR = 0.21, 95%CI 0.05–0.83), this association was not observed in the WSG-ADAPT-TN.

Conclusion The prevalence of HER2-low status varied between the analyzed trials. Our results show that survival does not differ between HER2-low and HER2-zero tumors in HR + /HER2- cohorts; however, HER2-low status appears to have an inconsistent impact on survival in TNBC. Therefore, our findings do not support the characterization of HER2-low status as a distinct BC subtype.

Introduction

Breast carcinoma is the most common malignant tumor disease in women [1-3]. A worldwide incidence of about 2.26 million cases in 2020, according to the global cancer burden, vividly demonstrates the extent of the disease and its global impact [4]. Different breast cancer subtypes are known, including the most common hormone receptor (HR)-positive, human epidermal growth factor receptor 2 (HER2)-negative type, to less frequent triplenegative breast cancer (HR-negative and HER2-negative) and HER2 – positive breast cancer [5].

HER2 is a prototype oncogene and an established therapeutic target in breast cancer (BC). Determination of the HER2 status in BC is a clinical routine that uses a combination of immunohistochemistry (IHC) to evaluate HER2 protein expression levels and in situ hybridization (ISH) to assess HER2 gene status [6, 7]. In 2007, the American Society of Clinical Oncology/College of American Pathologists (ASCO/CAP) released the first recommendations for HER2 testing for the distinction between HER2-negative and HER2-positive tumors [8]. The subsequent update in 2013 aimed at detecting and eliminating false negative cases by changing the cutoff for IHC score 3+ from 30 to 10% and changing the ISH HER2/CEP17 ratio \geq 2.0 or HER2 absolute gene copy number \geq 6.0 (previously HER2/CEP17 ratio \geq 2.2 or HER2 average gene copy number ≥ 6.0 [9]. The 2018 update focused on the relevance of IHC score 0 versus 1+by providing practical recommendations for testing and reporting [10]. The most recent 2023 update introduced the HER2-low category defined as IHC score 1+or 2+(ISH-negative)[11]. According to the current IHC algorithm, HER2-negative tumors are defined as tumors that are completely negative for HER2 (IHC score 0) as well as tumors with low (IHC score 1+) or moderate HER2 expression (IHC score 2+, ISH-negative). The minority of breast tumors are HER2-positive (15%) and show a HER2 overexpression with an IHC score 3+or 2+and ISH-positive. Until recently, IHC scores 0, 1+, and 2+with ISH-negative within the HER2-negative group have often been combined due to a lack of clinical relevance. However, a recent study (DESTINY-Breast04) showed that not only BC patients with HER2 overexpression but also patients with a low or moderate HER2 expression could benefit from antibody-drug conjugates like trastuzumab deruxtecan (T-DXd) [12]. Therefore, the question arises whether this HER2-low subgroup is a distinct entity different from HER2 IHC 0 BC.

The purpose of this study was to investigate whether patients with tumors with low HER2 expression (IHC 1 + or 2 + with ISH-negative) have a different clinical outcome compared to completely HER2-negative patients (HER2-zero, IHC 0). We also evaluated the agreement between the HER2 score derived from local and central pathologists and the association with the expression of the *ERBB2* gene on the mRNA level.

We used data from cohorts of patients with tumors originally classified as HER2- BC by local measurements, enrolled into three large clinical trials performed by the West Germany Study Group (WSG) in early BC: WSG-ADAPT-HR+/HER2- [13], WSG-ADAPT-TN [14], and adjuvant WSG-PlanB [15].

A standardized evaluation of HER2 expression in pretherapeutic core biopsies has been performed prospectively in the WSG-ADAPT trials and retrospectively in the WSG-PlanB trial as a part of central pathology assessment. Endpoints included disease-free survival (DFS) and overall survival (OS). Additionally, we compared biologically relevant parameters, e.g., hormone receptor status, grade, tumor proliferation (Ki-67), and stromal tumorinfiltrating lymphocytes (sTILs) between the HER2-low and HER2-zero tumors.

Methods

Study design, clinical cohorts, and central pathology.

WSG-ADAPT-HR+/HER2- (NCT01779206) was a multi-center, randomized, open-label, phase II/III neoadjuvant trial in patients with primary unilateral invasive, locally hormone receptor (HR)-positive, and HER2-negative BC. Potential candidates for neoadjuvant chemotherapy (cT2-4 or cN+or G3 or Ki-67 \geq 15%) were enrolled. Patients received 3 weeks of induction endocrine therapy (ET). Ki-67 was analyzed centrally in baseline diagnostic core biopsies and after the induction of ET, and patients with Ki-67_{post-ET}≤10% were considered as ET-responders [16]. Patients with cN0-1 and either Recurrence Score (RS, Oncotype DX) ≤ 11 or RS between 12 and 25 with ET-response were included in the "endocrine subtrial" and received no chemotherapy in either neoadjuvant or adjuvant phase. Patients with cN2-3 or RS>25 or very-high-clinical-risk (cN2-3 OR (G3 AND baseline Ki-67>40% AND \geq cT1c) or with ET-non-response (Ki- $67_{\text{post-ET}} > 10\%$) entered the "chemotherapy sub-trial". These patients were randomized with a 1:1 ratio to 8 weeks of intravenous sb-paclitaxel or nab-paclitaxel. Afterwards, patients received 4 cycles of epirubicin/ cyclophosphamide (EC) q2w within a dose-dense chemotherapy schedule. Results for the primary endpoint, invasive DFS (iDFS), were published in 2022 [13].

WSG-PlanB (NCT01049425) was a multi-center, randomized, open-label, phase III adjuvant trial in patients with unilateral primary invasive BC with adequate surgical treatment and HER2-negative status, pT1-4c, known HR-status, pN+ or pN0 with at least one risk factor (pT2 or greater, G2-3, high uPA/PAI-1, age <35 years, or HRnegative status). Patients were randomized to 6 cycles of docetaxel plus cyclophosphamide q3w or to 4 cycles of EC q3w. Following the amendment, enrolled patients with HR+disease and RS <12 received ET without chemotherapy. Results for the primary endpoint, DFS, were published in 2019 [17].

WSG-ADAPT-TN (NCT01815242) was a multi-center, randomized, open-label, phase II neoadjuvant trial in unilateral primary invasive, cT1c-cT4c or cN+centrally-confirmed triple-negative (TN) BC. Patients were randomized with a 1:1 ratio to nab-paclitaxel plus gemcitabine q3w or to nab-paclitaxel plus carboplatin q3w. Results for the primary endpoint, pathological complete response (pCR, defined as ypT0/is, ypN0, assessed after twelve weeks of treatment), were published in 2017 [14].

HER2 testing was conducted in each trial following the ASCO/CAP guidelines at that time: according to 2007 guidelines in WSG-PlanB [8] and according to 2007 and 2013 guidelines in WSG-ADAPT trials [8, 9]. In the WSG-ADAPT trials, central HER2 testing was performed prospectively before randomization. In the WSG-PlanB trial, central HER2 determination was performed retrospectively using tissue microarrays from surgical samples. In line with the German Gynecological Oncology Group guidelines (Arbeitsgemeinschaft Gynäkologische Onkologie, AGO, [18]), HER2-low status was defined for the purpose of this analysis as IHC 1+or 2+and ISH-negative, and HER2-zero was defined as IHC 0. HER2 IHC status was determined at diagnosis by a local and the first central assessment and after three weeks of treatment by the second central assessment. Additionally, the three IHC assessments were combined and defined as (i) HER2-negative if all three assessments were HER2-negative (IHC scores 0, 1+, and 2+with ISH-negative) or at least one assessment was HER2-negative while others were HER2-negative or had unclear HER2 status; (ii) HER2-positive if at least one assessment was HER2-positive; (iii) had unclear HER2 status if all three assessments were unclear; (iv) or were defined as HER2-low in the remaining cases.

Correlations between the different HER2 status evaluations were defined as follows: as no change if the first (diagnostic) and second (after three weeks of treatment) central assessments were identical; as a decrease if the first central assessment was HER2-low while the second central assessment was HER2-negative; as an increase if the first central assessment was HER2-negative while the second central assessment was HER2-low or HER2-positive or the first central assessment was HER2-low and the second central assessment was HER2-low and the second central assessment was HER2-positive.

Gene expression analysis

Expression of the ERBB2 gene mRNA was evaluated in baseline tumor biopsies. Oncotype DX test (Exact Sciences, [19]) utilizing reverse transcriptase quantitative polymerase chain reaction (RT-qPCR) was used to analyze the expression of 21 genes in the WSG-ADAPT-HR+/HER2- trial and in the HR+cohort within the WSG-PlanB trial, as described previously [13, 20]. Customized nCounter 119-gene expression panels (NanoString Technologies Inc.) utilizing the multiplex nucleic acid hybridization, were used in the WSG-ADAPT-TN trial and in the HR- cohort within the WSG-PlanB trial [21, 22]. The customized 119-gene panel included targets for PAM50 intrinsic subtype predictor, Claudin-low subtype predictor, and VEGF/ hypoxia signature (see https://www.ncbi.nlm.nih.gov/ geo/query/acc.cgi?view=data&acc=GPL17071&id= 42240&db=GeoDb_blob111), and nine immune-related genes (PD1, PDL1, CD8, CD4, AR, FOXP3, and three additional genes for PAM50). In both gene expression assays, the raw ERBB2 gene expression data was normalized using the set of housekeeping genes: ACTB, GAPDH, GUS, RPLPO, and TFRC in the Oncotype DX assay and ACTB, MRPL19, PSMC4, RPLP0, and SF3A1 in the nCounter assay. Additionally, ERBB2 mRNA levels in a few patients in the HR+/HER2- cohort in the WSG-PlanB trial were analyzed by the PAM50 gene expression assay (NanoString Technologies Inc.).

Statistical analysis

Patient's and tumor's characteristics were compared between HER2-negative and HER2-low using a Pearson's

Chi-square test, a Fisher's exact test or a linear-by-linear test (categorical variables), and a t-test or Mann-Whitney-U test (continuous variables). Agreement between the different HER2 assessments was evaluated with a Cohen's kappa coefficient, and the percentage agreement was calculated as a percentage of assessments with the same classification of HER2. Effects of HER2 on pCR occurrence were estimated with separate unadjusted and adjusted logistic regressions, while effects of HER2 on distant disease-free survival (dDFS), iDFS, DFS, and OS were estimated with separate unadjusted and adjusted Cox proportional hazards models. iDFS, dDFS, and OS were defined based on the STEEP criteria [23]. iDFS events (in WSG-ADAPT-TN) included local/ regional invasive recurrence, distant recurrence, invasive ipsilateral recurrence, invasive contralateral BC, second primary invasive non-BC, death from any cause; dDFS events in (WSG-ADAPT-TN and WSG-ADAPT-HR+/ HER2-) included distant recurrence, second primary invasive non-BC, death from any cause; and OS event (in WSG-PlanB and WSG-ADAPT-TN) was death from any cause. DFS in WSG-PlanB was defined with the following events: local, regional, or metastatic relapse, second primary cancer (with the exception of curatively treated non-melanoma skin cancer or in situ carcinoma of the cervix), and death from any cause. The adjusted effects of HER2 were obtained by including the patient's and tumor's characteristics that in univariable models had p-values < 0.10 and were significant in multivariable models. Additionally, interaction terms between HER2 and pCR and HER2 and treatment arm were tested in final multivariable models. Survival curves were presented with the Kaplan-Meier method and compared with a log-rank test. Furthermore, the prognostic effects of the HER2 dynamic on survival outcomes were evaluated using the same methodology. P-values ≤ 0.05 were considered statistically significant. All analyses were performed with STATA version 18.

Results

A total of 5598 patients with HR + /HER2- BC (as per initial local assessment) from the screening cohort of WSG-ADAPT-HR+/HER2-, 2162 patients with HR + /HER2- and 430 patients with HR - /HER2- tumors from the adjuvant WSG-PlanB trial and 336 patients from the WSG-ADAPT-TN trial were evaluated. However, a small number of patients in each trial were missing the HER2 IHC data from local or central assessments.

WSG-ADAPT-HR + /HER2- trial

In the cohort of WSG-ADAPT-HR+/HER2-, a total of 55.6% (n=3096 of 5571) and 64.6% (n=3198 of 4948) of tumors were HER2-low (IHC 1+or 2+and

ISH-negative) by local and the first central histological evaluation, respectively (Fig. 1A). 2.1% (n = 106 of 4948) of samples were identified as HER2-positive by the first central assessment and were thus excluded from further analysis. The agreement between the local and the first central evaluation for HER2-low and/or -zero was 61.0% (kappa 0.21, Table 1).

A second central assessment of HER2 status was carried out after a short (2–4 weeks) induction ET with tamoxifen in premenopausal and mostly aromatase inhibitors (AI) in postmenopausal women. At the second central assessment, 75.2% (n=3262 of 4336) of tumors were classified as HER2-low (Fig. 1A). Thus, this assessment showed the highest frequency of HER2-low tumors in comparison to the baseline local and central assessment. The agreement between the first and the second central assessment was 69.2% (kappa 0.30, Table 1). 60.2% (n=823 of 1368) of HER2-zero cases at baseline changed to HER2-low, but only 13.1% (n=345 of 2628) of the cases changed from HER2-low to HER2-zero after the short ET, by central assessment (Fig. 1B).

Higher expression of HER2, with 35% of the tumors classified as IHC 2+, was observed after ET in comparison to both baseline assessments that classified 24% of the tumors as IHC 2+. The clinicopathological parameters for patients with local assessments of HER2 are shown in Table 2. The HER2-low cases showed slightly, but significantly higher *ERBB2* mRNA expression by Oncotype DX in comparison to HER2-zero cases (mean 9.3 vs. 9.1, p < 0.001, for the first local assessment, Table 2; mean 9.4 vs. 8.8, p < 0.001, for first central assessment; mean 9.3 vs. 8.9, p < 0.001, for second central assessment).

A combination of all three HER2 assessments classified 83.5% (n=4675 of 5598) of the tumors as HER2-low. Importantly, HER2-positive status was found in an additional 66 (1.2%) surgical samples.

Ki-67 levels at baseline or at week 3 did not differ between HER2-zero and HER2-low cases.

Patients with HER2-low tumors treated by neoadjuvant chemotherapy (mostly dose-dense paclitaxel-EC or $8 \times$ nab-paclitaxel, q1w—4xEC, q2w) had significantly less often a pCR compared to the HER2-zero patients (14.2% vs. 20.5%; p=0.015, Table 2), and there was no interaction with the therapy arm.

However, after a median of 61 months follow-up, there was no significant difference in dDFS between HER2-low and HER2-zero tumors by neither local (unadjusted HR=0.95, 95%CI 0.76–1.18) and central first (unadjusted HR=1.06, 95%CI 0.83–1.36) nor central second assessment (unadjusted HR=1.04, 95%CI 0.77–1.41, Supplementary Fig. 1A-C). These results were also true if analyzed separately by ET and chemoendocrine therapy. Moreover, the dynamics of HER2 expression (assessed



Fig. 1 A HER2 assessment by local and the first and second central assessment and B changes in HER2 status between the first and the second central assessment in the WSG-ADAPT-HR+/HER2- trial

centrally at baseline and at week 3 of ET) were not associated with survival outcomes (Supplementary Fig. 1D).

HR+/HER2- cohort in the WSG-PlanB trial

In the HR+/HER2- cohort of WSG-PlanB, 28.7% of tumors (n=601 of 2097) were HER2-low by first central assessment (see Table 2 for the clinicopathological characteristics of patients). 15.6% (n=94 of 601) of the HER2-low cases were classified as IHC 2+, and the HER2-low tumors had significantly higher *ERBB2* mRNA

expression by Oncotype DX in comparison to HER2zero tumors (mean 9.2 vs. 8.8, p < 0.001, Table 2). However, no significant difference was observed in OS and DFS between HER2-low and HER2-zero tumors (OS: unadjusted HR = 1.21, 95%CI 0.73–2.02; DFS: unadjusted HR = 1.28, 95%CI 0.91–1.82, Supplementary Fig. 2A, B).

Moreover, no interaction with the efficacy of anthracycline-containing or anthracycline-free treatment was observed neither with immunohistochemical- (p=0.776) nor with RT-qPCR-based HER2 expression (p=0.918). Table 1 Agreement between the local and the first central and the second central assessments in the WSG-ADAPT-HR+HER2- trial

Assessment		First central assessment (baseline)				
		HER2-zero	HER2-low	HER2-positive	Total	
Local assessment (baseline)	HER2-zero	990 (60.3%)	1166 (36.7%)	24 (23.5%)	2180 (44.3%)	
	HER2-low	651 (39.7%)	2013 (63.3%)	78 (76.5%)	2742 (55.7%)	
	Total	1641 (33.3%)	3179 (64.6%)	102 (2.1%)	4922 (100%)	
	Карра			0.208		
Second central assessment (week 3)	HER2-zero	537 (39.3)	345 (13.1%)	4 (5.7%)	886 (21.8%)	
	HER2-low	823 (60.1%)	2230 (84.9%)	18 (25.7%)	3071 (75.5%)	
	HER2-positive	8 (0.6%)	53 (2.0%)	48 (68.6%)	109 (2.7%)	
	Total	1368 (33.6%)	2628 (64.5%)	70 (1.7%)	4066 (100%)	
	Карра			0.298		

However, significantly worse DFS was observed for higher HER2 expression by RT-qPCR in 317 patients with up to 3 positive lymph nodes and RS 0-11 treated by ET alone (unadjusted HR for continuous levels = 2.35, 95%CI 1.10-5.00).

HR-/HER2- cohort in the WSG-PlanB trial

In the HR-/HER2- cohort of WSG-PlanB, only 10.5% (n=45 of 430) of cases were HER2-low and 1.4% (n=6 of 430) were HER2-positive by central assessment. Of all the baseline characteristics studied, only *ERBB2* mRNA expression by nCounter assay was found to be significantly related to HER2 status; *ERBB2* mRNA expression was higher in HER2-low than in HER2-zero tumors (mean -1.0 vs. -2.0; p<0.001). Patients with HER2-low tumors had significantly better DFS (unadjusted HR=0.21, 95%CI 0.05–0.83) but not OS (unadjusted HR=0.38, 95%CI 0.09–1.59) in comparison to HER2-zero cases (Supplementary Fig. 3A, B).

WSG-ADAPT-TN trial

In the HR-/HER2- cohort of the WSG-ADAPT-TN trial, HER2-low status at baseline was found in 37.9% (n=127 of 335) of cases by local assessment and 41.3% (n=137 of 332) by the first central assessment with overall agreement of 65% (kappa 0.28, Fig. 2, Table 3). Moreover, 2.6% (n=6 of 229) of tumors turned out to be HER2-positive, and 54.6% (n=125 of 229) were found to be HER2-low by the second biopsy after 3 weeks of neoadjuvant chemotherapy. The agreement between central assessment at the first and second samples was 67% (kappa 0.37, Table 3). In general, 67.6% (n=227 of 336) of tumors turned out to be HER2-low by at least one of the three assessments.

Only the first central assessment of HER2 was significantly associated with *ERBB2* mRNA levels by nCounter assay with HER2-low cases having a higher *ERBB2* mRNA expression in comparison to HER2-zero (mean 0.3 vs. -0.2, p = 0.001, Table 4). Regarding association with biological patterns, we found significantly lower Ki-67 levels in HER2-low than HER2-zero cases by all three assessments (see Table 5 for the clinicopathological characteristics of patients). Locally assessed HER2-low status was associated with lower odds of pCR occurrence; however, only when the effect was not adjusted for any patient's or tumor's characteristics (unadjusted OR=0.62, 95%CI 0.38–1.00; adjusted OR=0.79, 95%CI 0.45–1.40). This effect was not observed with central assessments.

After 60 months of median follow-up, neither iDFS nor OS were significantly associated with HER2 status by first local and second central assessment (Supplementary Fig. 4), as well as no significant interaction between study arm and pCR status was found. However, in comparison to HER2-zero cases, HER2-low cases assessed by second central assessment had a significantly lower risk for iDFS event up to first two years of follow-up (at 1 year: adjusted HR = 0.28, 95%CI 0.14-0.58; at 2 years adjusted: HR = 0.52, 95%CI 0.31-0.89) but not after longer follow-up, when the effect was adjusted for pCR, clinical node status and sTILs measured at baseline (Supplementary Fig. 4B). This effect significantly differed between carboplatin-free and carboplatin-containing arms up to 1-year followup (interaction test at 1-year follow-up: p = 0.023). It was highly pronounced (weaker regarding pCR) in the carboplatin-free arm (adjusted HR=0.20, 95%CI 0.08–0.54), but not in the carboplatin-containing arm (adjusted HR = 0.85, 95%CI 0.40-1.82, Supplementary Fig. 5A and B). Similarly, a significant effect on dDFS of HER2 assessed by the second central assessment was observed (adjusted HR = 0.47, 95%CI 0.27-0.80) irrespective of pCR (none of HER2-low pCR cases experienced dDFS event, Supplementary Fig. 5C).

Table 2 Clinicopathological parameters in HR + cohorts and association with HER2-low status by local assessment in the WSG-ADAPT-HR + /HER2- trial and by first central assessment WSG-PlanB trial

Characteristic	WSG-ADAPT-H	HR+/HER2- trial			WSG-PlanB trial			
	HER2-zero, N = 2475	HER2-low, N = 3096	Overall, N=5571*	p-value**	HER2-zero, N = 1468	HER2-low, N=601	Overall, N = 2069***	p-value****
Age (years)								
Mean (SD)	55.3 (10.3)	54.6 (10.4)	54.9 (10.4)	.038	55.8 (10.1)	55.4 (9.7)	55.7 (10.0)	.432
Median (min, max)	54 (23, 84)	54 (20, 85)	54 (20, 85)		55.5 (27, 77)	55 (25, 76)	55 (25, 77)	
Estrogen recepto	or (%)							
Mean (SD)	90.4 (17.3)	90.7 (16.8)	90.6 (17.1)	.557	90.6 (21.1)	92.4 (17.8)	91.1 (20.2)	.042
Median (min, max)	100 (0, 100)	100 (0, 100)	100 (0, 100)		100 (0, 100)	100 (0, 100)	100 (0, 100)	
Missing	295 (11.9%)	344 (11.1%)	639 (11.5%)		7 (0.5%)	0 (0.0%)	7 (0.3%)	
Progesterone rec	ceptor (%)							
Mean (SD)	64.4 (36.6)	62.2 (36.7)	63.2 (36.6)	.031	62.7 (41.7)	62.9 (40.4)	62.8 (41.3)	.948
Median (min, max)	80 (0, 100)	80 (0, 100)	80 (0, 100)		80 (0, 100)	80 (0, 100)	80 (0, 100)	
Missing	288 (11.6%)	348 (11.2%)	636 (11.4%)		11 (0.8)	3 (0.5%)	14 (0.7%)	
Ki-67, baseline (%	%)							
Mean (SD)	22.5 (14.9)	22.2 (14.3)	22.3 (14.6)	.436	15.8 (11.0)	16.9 (11.1)	16.1 (11.0)	.057
Median (min, max)	20 (0, 95)	20 (0, 100)	20 (0, 100)		15 (1, 95)	15 (1, 80)	15 (1, 95)	
Missing	413 (16.7%)	452 (14.6%)	865 (15.5%)		83 (5.7%)	18 (3.0%)	101 (4.9%)	
Ki-67, week 3 (%))							
Mean (SD)	12.5 (13.2)	12.4 (12.2)	12.4 (12.6)	.713	NA	NA	NA	NA
Median (min, max)	10 (0, 95)	10 (0, 90)	10 (0, 95)		NA	NA	NA	
Missing	577 (23.3%)	710 (22.9%)	1287 (23.1%)		NA	NA	NA	
RS								
Mean (SD)	19.2 (10.9)	20.3 (10.8)	19.8 (10.9)	<.001	18.9 (10.1)	20.1 (10.1)	19.2 (10.1)	.016
Median (min, max)	17 (0, 75)	18 (0, 76)	18 (0, 76)		18 (0, 69)	18 (0, 99)	18 (0, 99)	
Missing	282 (11.4%)	324 (10.5%)	606 (10.9%)		47 (3.2%)	13 (2.2%)	60 (2.9%)	
ERBB2 mRNA by	Oncotype DX							
Mean (SD)	9.1 (0.7)	9.3 (0.7)	9.2 (0.7)	<.001	8.8 (0.7)	9.2 (0.7)	8.9 (0.7)	<.001
Median (min, max)	9.1 (6.9, 11.9)	9.4 (7.6, 12.9)	9.2 (6.9, 12.9)		8.9 (5.5, 12.7)	9.2 (6.9, 11.7)	9 (5.5, 12.7)	
Missing	611 (24.7%)	793 (25.6%)	1404 (25.2%)		62 (4.2%)	15 (2.5%)	77 (3.7%)	
ERBB2 mRNA by	PAM50*****							
Mean (SD)	NA	NA	NA	NA	-1.8 (0.8)	-0.3 (1.3)	-1.4 (1.1)	.005
Median (min, max)	NA	NA	NA		-1.9 (-2.8, 0.8)	-0.5 (-1.6, 2.0)	-1.7 (-2.8, 2.0)	
Missing	NA	NA	NA		1448 (98.6%)	595 (99.0%)	2043 (98.7%)	
Histology Type								
NST	1749 (80.6%)	2304 (84.1%)	4053 (82.5%)	.014	1226 (83.7%)	519 (86.4%)	1745 (84.5%)	.304
ILC	367 (16.9%)	383 (14.0%)	750 (15.3%)		201 (13.7%)	70 (11.7%)	271 (13.1%)	
Mucinous	31 (1.4%)	30 (1.1%)	61 (1.2%)		0 (0.0%)	0 (0.0%)	0 (0.0%)	
Other	24 (1.1%)	23 (0.8%)	47 (1.0%)		38 (2.6%)	12 (2.0%)	50 (2.4%)	
Missing	304 (12.3%)	356 (11.5%)	660 (11.9%)		3 (0.2%)	0 (0.0%)	3 (0.1%)	
Histological grad	le							
0-1	146 (8.1%)	171 (7.6%)	317 (7.8%)	.687	83 (5.7%)	33 (5.5%)	116 (5.6%)	.063
2	979 (54.2%)	1260 (55.9%)	2239 (55.2%)		919 (62.6%)	348 (57.9%)	1267 (61.2%)	

Table 2 (continued)

Characteristic	WSG-ADAPT-H	HR+/HER2- trial			WSG-PlanB trial			
	HER2-zero, N = 2475	HER2-low, N = 3096	Overall, N = 5571*	p-value**	HER2-zero, N = 1468	HER2-low, N=601	Overall, N = 2069***	p-value****
3	682 (37.7%)	822 (36.5%)	1504 (37.0%)		466 (31.7%)	220 (36.6%)	686 (33.2%)	
Missing	668 (27.0%)	843 (27.2%)	1511 (27.1%)		0 (0.0%)	0 (0.0%)	0 (0.0%)	
Tumor stage****	***							
0/is	26 (1.3%)	10 (0.4%)	36 (0.8%)	.984	0 (0.0%)	0 (0.0%)	0 (0.0%)	.161
1	1081 (52.5%)	1366 (53.7%)	2447 (53.2%)		785 (53.6%)	339 (56.6%)	1124 (54.5%)	
2	856 (41.6%)	1059 (41.6%)	1915 (41.6%)		611 (41.7%)	238 (39.7%)	849 (41.2%)	
3–4	97 (4.7%)	108 (4.3%)	205 (4.5%)		68 (4.6%)	22 (3.7%)	90 (4.4%)	
Missing	415 (16.8%)	553 (17.9%)	968 (17.4%)		4 (0.3%)	2 (0.3%)	6 (0.3%)	
Nodal stage****	**							
0	1674 (67.6%)	2060 (66.6%)	3734 (67.0%)	.773	817 (55.7%)	354 (58.9%)	1171 (56.6%)	.044
1	612 (24.7%)	817 (26.4%)	1429 (25.7%)		539 (36.7%)	218 (36.3%)	757 (36.6%)	
2-3	189 (7.6%)	218 (7.0%)	407 (7.3%)		112 (7.6%)	29 (4.8%)	141 (6.8%)	
Missing	0 (0.0%)	1 (0.0%)	1 (0.0%)		0 (0.0%)	0 (0.0%)	0 (0.0%)	
pCR (ypT0/is, ypl	N0), neoadjuvant	treated patients o	nly					
No	299 (79.5%)	417 (85.8%)	716 (83.1%)	.015	NA	NA	NA	NA
Yes	77 (20.5%)	69 (14.2%)	146 (16.9%)		NA	NA	NA	
Missing	2099 (84.8%)	2610 (84.3%)	4709 (84.5%)		NA	NA	NA	

Percentages for categories other than missing were obtained among non-missing observations and they sum up to 100%

^{*} 27 patients (0.5%) were classified as having unclear HER2 status and were excluded from the analyses

** P-values were obtained with a t-test (continuous variables) and a Pearson's Chi-square test or a linear-by-linear test (categorical variables)

*** 28 patients (1.3%) were classified as HER2-positive, and 65 (3.0%) patients were classified as having unclear HER2 status and were excluded from the analyses

**** P-values were obtained with a t-test or Mann–Whitney-U test (continuous variables) and a Pearson's Chi-square test or a linear-by-linear test (categorical variables). Missing categories were excluded from statistical testing

***** ERBB2 mRNA levels were analyzed by PAM50 in locally or centrally HR- cases in the WSG-PlanB trial

****** Denotes composite tumor or nodal stage for WSG-ADAPT-HR + /HER2- trial (clinical stage in patients randomized to neoadjuvant treatment, pathological stage in patients after primary surgery); pathological tumor or nodal stage in WSG-PlanB trial

NST, non-special type carcinoma; ILC, invasive lobular carcinoma

Furthermore, the dynamics of HER2 expression had no significant impact on iDFS, dDFS, or OS. However, an increase in HER2 expression during therapy seemed to be associated with better survival outcomes (Fig. 3).

Discussion

For the first time, we have reported results on HER2-low status from three large prospective contemporary trials in HER2-negative early BC according to both central and local assessments and its relationship to *ERBB2* mRNA expression in different genomic signatures.

The first important finding from our study is a clinically meaningful difference in HER2-low status between the older WSG-PlanB trial, which used tissue microarrays performed on surgical samples for central assessment of HER2, and the more current WSG-ADAPT trial, which determined HER2 on complete core biopsies as first assessment. Therefore, different tissue formats used for HER2 assessment in WSG-ADAPT and in the WSG-PlanB trial may well explain the observed differences in HER2-low frequency.

Although a positive association between HER2-low status and ERBB2 mRNA expression was found in all four analyzed data sets, we have seen a significantly lower HER2-low prevalence in the WSG-PlanB trial (28.7% and 10.5% in HR+and HR- samples, respectively) vs. the WSG-ADAPT trial (64.6% and 41.3% in HR+and HR- subsets, respectively). This is despite the fact that the same central lab evaluated the HER2 expression and that HER2 levels by IHC positively correlated with HER2 expression by RT-qPCR from the Oncotype DX test. The HER2-low rate in the WSG-ADAPT, but not in the WSG-PlanB trial, appears comparable to those published by Denkert et al. in the large analysis by the German Breast Group (64% and 36% in the HR + and HR- subsets, respectively) [24] and comparable to the meta-analysis published by Xia et al. (66.5% and 33.5% in the HR+and HR-group)[25]. Similarly, Tarantino et al. reported HER2-low rates of 40% in HR- tumors and 62% in tumors



Fig. 2 A HER2 assessment by local and the first and second central assessment and B changes in HER2 status between the first and the second central assessment in the WSG-ADAPT-TN trial

with ER expression > 95% [26]. Recently, National Cancer Data Base (NCDB) analysis demonstrated a 70% HER2-low rate in HR + /HER2- cases [27].

Interestingly, we have observed a strong increase in HER2 expression with up to 75.2% of HER2-low cases after induction ET with AI or tamoxifen in the WSG-ADAPT-HR+/HER2- trial. These results are in line with the single-center phase II trial published by Chaudhary et al., who reported an increase in HER2 expression in 49% of cases after neoadjuvant ET [28]. A similar effect was observed in the WSG-ADAPT-TN cohort,

with 37.9% of HER2-zero tumors by central assessment becoming HER2-low after 3 weeks of neoadjuvant chemotherapy. A significant dynamics of HER2 expression and a very high rate of HER2-low status, if several biopsies/ analyses were repeated, has also recently been found by Bar et al. [29]. Several pathomechanisms could drive an increased HER2 expression during the ET. For instance, HER2 seems to activate nuclear factor kappa B (NF- κ B), which in turn contributes to resistance against ET [30]. Therefore, it is possible that increased HER2 levels during induction ET may result from the expansion of

Table 3Agreement between the local and the first central andthe second central assessments in the WSG-ADAPT-TN trial

Assessment	First central assessment (baseline)					
	HER2-zero	HER2-low	Total			
Local assessment (b	aseline)					
HER2-zero	142 (73.2%)	63 (46.0%)	205 (61.9%)			
HER2-low	52 (26.8%)	74 (54.0%)	126 (38.1%)			
Total	194 (58.6%)	137 (41.4%)	331 (100.0%)			
Карра		0.275				
Second central asse	ssment (week 3)					
HER2-zero	80 (60.6%)	18 (19.2%)	98 (43.4%)			
HER2-low	50 (37.9%)	72 (76.6%)	122 (54.0%)			
HER2-positive	2 (1.5%)	4 (4.3%)	6 (2.7%)			
Total	132 (58.4%)	94 (41.6%)	226 (100.0%)			
Карра		0.373				

 Table 4
 Comparison of *ERBB2* mRNA expression between HER2zero and HER2-low tumors by the local and central assessments in the WSG-ADAPT-TN trial

HER2 status	ERBB2 mRNA expression by nCounter assay							
	Mean (SD)	Median (min, max)	Missing	p-value				
Local assessm	ent (baseline)							
HER2-zero	- 0.08 (0.94)	- 0.46 (- 0.82, 5.97)	17 of 208 (8%)	.083				
HER2-low	0.13 (1.09)	- 0.26 (- 0.82, 3.78)	13 of 127 (10%)					
First central as	sessment (bas	seline)						
HER2-zero	- 0.17 (0.77)	- 0.46 (- 0.82, 4.00)	16 of 195 (8%)	.001				
HER2-low	0.26 (1.22)	– 0.23 (– 0.82, 5.97)	13 of 137 (9%)					
Second centra	al assessment ((week 3)						
HER2-zero	- 0.05 (0.93)	- 0.39 (- 0.82, 4.00)	4 of 98 (4%)	.134				
HER2-low	0.17 (1.18)	- 0.26 (- 0.81, 5.97)	9 of 125 (7%)					
Any assessme	nt							
HER2-zero	- 0.20 (0.76)	- 0.46 (- 0.82, 4.00)	4 of 103 (4%)	.006				
HER2-low	0.09 (1.08)	- 0.31 (- 0.82, 5.97)	26 of 227 (11%)					

HER2-expressing tumor cells with activated NF-κB signaling. Given the very promising activity of T-DXd and trastuzumab duocarmazine in apparently HER2-negative disease [31, 32], it would interesting to see whether different patterns of transition between HER2-zero and HER2-low status affect the efficacy of these antibody–drug conjugates. Still, several mechanisms of resistance

to T-DXd, including inhibition of ferroptosis, have been described [33] and could limit the activity of antibody–drug conjugates in HER2-low disease.

However, the overall concordance for HER2-low status between local and central assessments was relatively low, despite a similar prevalence of HER2-low status in both assessments and a similar correlation with ERBB2 expression by RT-qPCR. A lack of awareness regarding the importance of distinguishing between the HER2zero and HER2-low subgroups among the HER2-negative cases (at the time when the trials were performed) likely contributed to the observed discordance between local and central results. This problem could be further compounded by the heterogeneous patterns of HER2 expression [34]. Overall, this stresses the need for standardization of HER2 testing in HER2-negative tumors in order to increase the concordance between the assessments. Though criteria for HER2 scoring have been clearly reported by current ASCO/CAP guidelines, it is still debated whether such methods (i.e., IHC and ISH assay) are appropriate for the detection of low levels of HER2 expression. Novel quantitative assays are currently under development and investigation, with the aim of improving the accuracy of HER2 testing [35, 36]. Alternatively, adequate training of pathologists has been recently shown to improve the diagnostic accuracy for identifying HER2-low cases [37]. A recent retrospective study by Viale et al. has shown that about 30% of historically HER2-zero cases would be rescored as HER2-low by a second assessment [38]. Another recent study provides excellent concordance only in tumors with HER2 IHC 3+, but not in cases with lower expression [39].

Type of samples (core biopsy vs. surgery sample) may also play a role in observed different positivity rates, however our data on the second sample in the WSG-ADAPT trial (mostly surgery samples after ET) and also other studies do not support lower HER2 expression on the surgery samples [40] due to methodological issues (e.g. fixation).

Regarding the pCR rate, we have observed lower response rates in HER2-low vs HER2-zero cases in HR+/ HER2- disease, in line with other publications [24, 26, 36] as well as a recently published meta-analysis [41]. However, this effect was pronounced only in tumors with locally assessed HER2-low status, as well as in samples after induction ET. Interestingly, another meta-analysis demonstrated lower pCR rates in HER2-low tumors regardless of the hormone receptor status [25]. A highly preselected patient population in the HR+/HER2cohort in the WSG-ADAPT-HR+/HER2- trial makes any kind of cross-trial comparisons difficult. Impact on pCR was inconsistent in HR-/HER2- disease (found only in the HER2-low by local assessment but not confirmed

Table 5 Clinicopathological parameters and association with HER2-low status by local and by the first and the second centra
assessment in the WSG-ADAPT-TN trial

Baseline Local assessment (baseline)*		e)*	First central	assessment (b	aseline)**	Second central assessment (week 3)***			
	HER2-zero, N=208	HER2-low, N = 127	p-value****	HER2-zero, N = 195	HER2-low, N = 137	p-value ****	HER2-zero, N = 98	HER2-low, N = 125	p-value ****
Age (years)									
Mean (SD)	50.8 (11.7)	52.2 (11.7)	.319	50.6 (11.6)	52.5 (11.8)	.145	51.6 (12.0)	50.5 (10.8)	.462
Median (min, max)	50 (26, 76)	53 (26, 75)		50 (27, 76)	53 (26, 75)		50 (29, 76)	50 (26, 75)	
Ki-67, baseline (%	6)								
Mean (SD)	69.9 (20.3)	60.3 (23.2)	<.001	69.6 (20.4)	62.2 (23.0)	.003	70.4 (19.7)	61.0 (24.7)	.002
Median (min, max)	75 (15, 100)	65 (10, 95)		75 (15, 95)	65 (10, 100)		75 (20, 95)	65 (10, 95)	
Missing	7 (3.4%)	5 (3.9%)		6 (3.1%)	4 (2.9%)		0 (0.0%)	6 (4.8%)	
sTILs, baseline									
Mean (SD)	29.7 (23.7)	28.7 (25.1)	.727	30.7 (24.8)	28.2 (23.8)	.363	30.6 (25.5)	28.1 (23.7)	.452
Median (min, max)	25 (0, 90)	20 (0, 90)		25 (0, 90)	20 (0, 90)		20 (0, 90)	20 (0, 90)	
Missing	7 (3.4%)	6 (4.7%)		5 (2.6%)	7 (5.1%)		1 (1.0%)	4 (3.2%)	
Treatment arm									
Gemcitabine	111 (53.4%)	70 (55.1%)	.755	113 (58.0%)	69 (50.4%)	.172	62 (63.3%)	71 (56.8%)	.329
Carboplatin	97 (46.6%)	57 (44.9%)		82 (42.0%)	68 (49.6%)		36 (36.7%)	54 (43.2%)	
Histological subt	ype								
Invasive ductal carci- noma	202 (98.5%)	124 (97.6%)	.678	191 (98.5%)	134 (97.8%)	.695	95 (96.9%)	121 (98.4%)	.657
Other	3 (1.5%)	3 (2.4%)		3 (1.5%)	3 (2.2%)		3 (3.1%)	2 (1.6%)	
Missing	3 (1.4%)	0 (0.0%)		1 (0.5%)	0 (0.0%)		0 (0.0%)	2 (1.6%)	
Histological grad	ling								
2	11 (5.3%)	11 (8.7%)	.236	10 (5.1%)	11 (8.0%)	.285	3 (3.1%)	13 (10.5%)	.038
3	195 (94.7%)	116 (91.3%)		185 (94.9%)	126 (92.0%)		95 (96.9%)	111 (89.5%)	
Missing	2 (1.0%)	0 (0.0%)		0 (0.0%)	0 (0.0%)		0 (0.0%)	1 (0.8%)	
Clinical tumor sta	age								
1	76 (36.5%)	48 (37.8%)	.817	77 (39.5%)	47 (34.3%)	.337	38 (38.8%)	38 (30.4%)	.190
2–4	132 (63.5%)	79 (62.2%)		118 (60.5%)	90 (65.7%)		60 (61.2%)	87 (69.6%)	
Clinical nodal sta	tus								
0	156 (75.0%)	92 (72.4%)	.604	146 (74.9%)	99 (72.3%)	.595	69 (70.4%)	90 (72.0%)	.794
1–3	52 (25.0%)	35 (27.6%)		49 (25.1%)	38 (27.7%)		29 (29.6%)	35 (28.0%)	
pCR (ypT0/is, ypN	NO)								
No	120 (59.7%)	86 (70.5%)	.05	118 (63.1%)	85 (63.9%)	.882	69 (71.1%)	91 (73.4%)	.71
Yes	81 (40.3%)	36 (29.5%)		69 (36.9%)	48 (36.1%)		28 (28.9%)	33 (26.6%)	
Missing	7 (3.4%)	5 (3.9%)		8 (4.1%)	4 (2.9%)		1 (1.0%)	1 (0.8%)	

Percentages for categories other than missing were obtained among non-missing observations and they sum up to 100%

* 1 patient (0%) was classified as having unclear HER2 status and was excluded from the analyses

** 4 patients (1.2%) were classified as having unclear HER2 status and were excluded from the analyses

*** 6 patients (1.8%) were classified as HER2-positive and 107 (31.9%) patients were classified as having unclear HER2 status and were excluded from the analyses

**** P-values were obtained with a t-test (continuous variables) and a Pearson's Chi-square test or a Fisher's exact test (categorical variables). Missing categories were excluded from statistical testing

by central assessment), which was in line with comparable pCR rates in TNBC published by Denkert et al. and Viale et al. [24, 38].

Moreover, we did not find a significant association between HER2-low status and iDFS in both WSG-ADAPT-HR+/HER2- and WSG-PlanB HR+/



Fig. 3 iDFS according to changes in HER2 status between the first and the second central IHC assessment in patients from the WSG-ADAPT-TN trial

HER2- cohorts by any of the assessments, which is in line with some other studies [24, 36, 42–44]. In both presented analyses from the NCDB, a better outcome in the HER2-low group was pronounced in patients treated by chemotherapy and/or in those with RS>25 [27], or in patients with higher stages of disease [45]. Therefore, patient preselection in both WSG trials may play some role. However, a meta-analysis published currently by Molinelli et al. shows significantly better iDFS and OS in both HR+and HR- cohorts in favor of HER2-low group [41].

In our HR-/HER2- cohorts, a significantly better DFS for HER2-low cases was found in the WSG-PlanB trial (with only 10% HER2-low rate), in line with previously mentioned and other studies [24, 25], but not in the overall analysis of the WSG-ADAPT-TN cohort with clearly higher HER2-low rates. However, a positive association between HER2-low status and pCR, particularly within the carboplatin-free arm, was found within the exploratory time-dependent multivariable analysis, including pCR. Importantly, as HER2-low status appeared highly dynamic between pretreatment and on-treatment biopsy (with up to 68% of all tumors found as HER2-low by one of three assessments), no significant association between change in HER2low status and iDFS was found, but increasing HER2 expression was associated with a non-significant trend towards a better outcome compared to decreasing HER2 levels. To summarize, there was no consistent correlation between HER2-low status and survival outcomes in our data set. These partially divergent results can be explained by different chemotherapy regimens used in the different cohorts as well as by different study designs. However, large differences in HER-low rates in different studies (from 10% up to 100%) as well as dynamics of HER2 expression during treatment indicate that HER2 levels can serve as a marker for different biological processes in the tumor, even in TNBC, rather than as a biological driver of these processes.

Our data, along with others, suggest that ongoing research is essential to improve the accuracy of identifying HER-low cases. Efforts should concentrate on further enhancing reproducibility, reducing interobserver variability, and investigating the role of HER2 expression heterogeneity. Moreover, a better molecular characterization of HER2-low tumors is needed to identify patients most likely to respond to novel antibody–drug conjugates and explore the mechanism of resistance.

Our study has several limitations. For instance, the difference in percentages of HER2-low tumors between the WSG-PlanB and WSG-ADAPT trials could be due to the use of tissue microarrays on surgical specimens in the former and core biopsies in the latter trial. Dissimilar HER2-low frequency between the local and central assessments could result from a former lack of awareness regarding the importance of classifying HER2-negative tumors into HER2-zero and HER2-low cases. Moreover, different methods were used for the analysis of *ERBB2* gene expression levels (Oncotype DX test in HR + cohorts and customized nCounter panel and PAM50 assay in HR- cohorts), which precludes the comparison between the trials.

Conclusions

In conclusion, our analysis of 8526 patients, as the largest data set reported so far to our best knowledge from randomized trials with consistent central and genomic assessment and survival results, does not support the concept that HER2-low is a distinct BC subtype. HER2low BC differs from HER2-zero BC in terms of slightly higher ERBB2/HER2 mRNA expression; however, it does not define a distinct subtype. Moreover, very recently presented results from the DESTINY-06 trial show high efficacy of T-DXd in patients with both HER2-low and even HER2-ultralow expression in metastatic HR+/ HER2- disease compared to first-line chemotherapeutic treatment. This indicates that HER2 expression is a very dynamic and heterogeneous factor, which could possibly explain the efficacy of antibody-drug conjugates also in apparently HER2-negative tumors.

Abbreviations

AI	Aromatase inhibitors
ASCO/CAP	American Society of Clinical Oncology/College of American
	Pathologists
BC	Breast cancer
DFS	Disease-free survival
dDFS	Distant disease-free survival
EC	Epirubicin/cyclophosphamide
ET	Endocrine therapy
HER2	Human epidermal growth factor receptor 2
HR	Hormone receptor
iDFS	Invasive disease-free survival
IHC	Immunohistochemistry
ILC	Invasive lobular carcinoma
ISH	In situ hybridization
NCDB	National Cancer Data Base
NST	Non-special type carcinoma
OS	Overall survival
pCR	Pathological complete response
RS	Recurrence Score
sTILs	Stromal tumor-infiltrating lymphocytes
T-DXd	Trastuzumab deruxtecan
TNBC	Triple-negative breast cancer
WSG	West Germany Study Group

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13058-025-01969-z.

Supplementary Figure 1	
Supplementary Figure 2	
Supplementary Figure 3	
Supplementary Figure 4	
Supplementary Figure 5	

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Author contributions

GS participated in the conception and design of the study, collection and assembly of data, data analysis and interpretation, and manuscript writing.

OG participated in the conception and design of the study, collection and assembly of data, data analysis and interpretation, and manuscript writing. MC participated in the conception and design of the study, collection and assembly of data, data analysis and interpretation, and manuscript writing. MR participated in the collection and assembly of data, and manuscript writing. SK participated in the collection and assembly of data, and manuscript writing. UN participated in the collection and assembly of data, and manuscript writing. MB participated in the collection and assembly of data, and manuscript writing. BA participated in the collection and assembly of data, and manuscript writing. KI H participated in the collection and assembly of data. and manuscript writing. HF participated in the collection and assembly of data, and manuscript writing. EMG participated in the collection and assembly of data, and manuscript writing. CS participated in the collection and assembly of data, and manuscript writing. RM participated in the collection and assembly of data, and manuscript writing. WM participated in the collection and assembly of data, and manuscript writing. TR participated in the collection and assembly of data, and manuscript writing. BN participated in the collection and assembly of data, and manuscript writing. AS participated in the collection and assembly of data, and manuscript writing. RW participated in the collection and assembly of data, and manuscript writing. MG participated in the collection and assembly of data, and manuscript writing. KJ participated in data analysis, visualization, and interpretation, manuscript writing. SB participated in data analysis, visualization, and interpretation, manuscript writing. CzE participated in data analysis, visualization, and interpretation, manuscript writing. ML participated in the collection and assembly of data, and manuscript writing. CKL participated in the collection and assembly of data, and manuscript writing. AP participated in data analysis and interpretation, and manuscript writing. PS participated in data analysis and interpretation, and manuscript writing. RB participated in data analysis and interpretation and manuscript writing. HHK participated in the conception and design of the study, collection and assembly of data, data analysis and interpretation, and manuscript writing. EFS participated in conception and design of the study, data analysis and interpretation, and manuscript writing. NH participated in the conception and design of the study, collection and assembly of data, data analysis and interpretation, and manuscript writing.

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Availability of data and materials

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

WSG-ADAPT und PlanB trials were approved by institutional ethic boards and ethic committees, and national authorities and were conducted according to Declaration of Helsinki. All patients have signed consent to participate.

Consent for publication

Not applicable.

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References

- Board WHOCWHOCTE, Cancer IAfRo: WHO Classification of Breast Tumours: WHO Classification of Tumours, Volume 2: World Health Organization; 2019.
- Kashyap D, Pal D, Sharma R, Garg VK, Goel N, Koundal D, Zaguia A, Koundal S, Belay A. Global increase in breast cancer incidence: risk factors and preventive measures. Biomed Res Int. 2022;2022:9605439.
- Rojas K, Stuckey A. Breast cancer epidemiology and risk factors. Clin Obstet Gynecol. 2016;59(4):651–72.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209–49.
- Howlader N, Altekruse SF, Li Cl, Chen VW, Clarke CA, Ries LA, Cronin KA. US incidence of breast cancer subtypes defined by joint hormone receptor and HER2 status. J Natl Cancer Inst. 2014;106(5):dju055.
- Sapino A, Goia M, Recupero D, Marchio C. Current challenges for HER2 testing in diagnostic pathology: state of the art and controversial issues. Front Oncol. 2013;3:129.
- Annaratone L, Sarotto I, Marchiò C: HER2 in Breast Cancer. In: *Encyclopedia* of *Pathology*. edn. Edited by van Krieken JHJM. Cham: Springer International Publishing; 2019: 1–11.
- Wolff AC, Hammond ME, Schwartz JN, Hagerty KL, Allred DC, Cote RJ, Dowsett M, Fitzgibbons PL, Hanna WM, Langer A, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for human epidermal growth factor receptor 2 testing in breast cancer. Arch Pathol Lab Med. 2007;131(1):18–43.
- Wolff AC, Hammond ME, Hicks DG, Dowsett M, McShane LM, Allison KH, Allred DC, Bartlett JM, Bilous M, Fitzgibbons P, et al. Recommendations for human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline update. J Clin Oncol. 2013;31(31):3997–4013.
- Wolff AC, Hammond MEH, Allison KH, Harvey BE, Mangu PB, Bartlett JMS, Bilous M, Ellis IO, Fitzgibbons P, Hanna W, et al. Human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists Clinical Practice Guideline Focused Update. J Clin Oncol. 2018;36(20):2105–22.
- 11. Wolff AC, Somerfield MR, Dowsett M, Hammond MEH, Hayes DF, McShane LM, Saphner TJ, Spears PA, Allison KH. Human epidermal growth factor receptor 2 testing in breast cancer: ASCO-College of American pathologists guideline update. J Clin Oncol. 2023;41(22):3867–72.

- Modi S, Jacot W, Yamashita T, Sohn J, Vidal M, Tokunaga E, Tsurutani J, Ueno NT, Prat A, Chae YS, et al. Trastuzumab deruxtecan in previously treated HER2-low advanced breast cancer. N Engl J Med. 2022;387(1):9–20.
- Nitz UA, Gluz O, Kümmel S, Christgen M, Braun M, Aktas B, Lüdtke-Heckenkamp K, Forstbauer H, Grischke E-M, Schumacher C, et al. Endocrine therapy response and 21-gene expression assay for therapy guidance in HR+/HER2– early breast cancer. J Clin Oncol. 2022;40(23):2557–67.
- Gluz O, Nitz U, Liedtke C, Christgen M, Grischke EM, Forstbauer H, Braun M, Warm M, Hackmann J, Uleer C, et al. Comparison of neoadjuvant Nab-Paclitaxel+Carboplatin vs Nab-Paclitaxel+Gemcitabine in triple-negative breast cancer: randomized WSG-ADAPT-TN trial results. J Natl Cancer Inst. 2018;110(6):628–37.
- 15. Gluz O, Nitz UA, Christgen M, Kates RE, Shak S, Clemens M, Kraemer S, Aktas B, Kuemmel S, Reimer T *et al.* West German Study Group Phase III PlanB Trial: First Prospective Outcome Data for the 21-Gene Recurrence Score Assay and Concordance of Prognostic Markers by Central and Local Pathology Assessment. J Clin Oncol 2016.
- 16. Nitz U, Gluz O, Kreipe HH, Christgen M, Kuemmel S, Baehner FL, Shak S, Aktas B, Braun M, Ludtke-Heckenkamp K, et al. The run-in phase of the prospective WSG-ADAPT HR+/HER2- trial demonstrates the feasibility of a study design combining static and dynamic biomarker assessments for individualized therapy in early breast cancer. Ther Adv Med Oncol. 2020;12:1758835920973130.
- Nitz U, Gluz O, Clemens M, Malter W, Reimer T, Nuding B, Aktas B, Stefek A, Pollmanns A, Lorenz-Salehi F, et al. West German Study PlanB trial: adjuvant four cycles of Epirubicin and cyclophosphamide plus docetaxel versus six cycles of docetaxel and cyclophosphamide in HER2-negative early breast cancer. J Clin Oncol. 2019;37(10):799–808.
- German Gynecological Oncology Group (Arbeitsgemeinschaft Gynäkologische Onkologie, AGO): Diagnosis and Treatment of Patients with early and advanced Breast Cancer [https://www.ago-online.de/fileadmin/agoonline/downloads/_leitlinien/kommission_mamma/2024/englisch/Einze ldateien/AGO_2024E_04_Pathology.pdf]
- Paik S, Shak S, Tang G, Kim C, Baker J, Cronin M, Baehner FL, Walker MG, Watson D, Park T, et al. A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. N Engl J Med. 2004;351(27):2817–26.
- Harbeck N, Gluz O, Clemens MR, Malter W, Reimer T, Nuding B, Aktas B, Stefek A, Pollmanns A, Lorenz-Salehi F, et al. Prospective WSG phase III PlanB trial: final analysis of adjuvant 4xEC→4x doc vs. 6x docetaxel/ cyclophosphamide in patients with high clinical risk and intermediateto-high genomic risk HER2-negative, early breast cancer. J Clin Oncol. 2017;35(15_suppl):504–504.
- Gluz O, Kolberg-Liedtke C, Prat A, Christgen M, Gebauer D, Kates R, Paré L, Grischke EM, Forstbauer H, Braun M, et al. Efficacy of deescalated chemotherapy according to PAM50 subtypes, immune and proliferation genes in triple-negative early breast cancer: primary translational analysis of the WSG-ADAPT-TN trial. Int J Cancer. 2020;146(1):262–71.
- 22. Gluz O, Liedtke C, Prat A, Christgen M, Gebauer D, Kates R, Pelz E, Clemens M, Warm M, Aktas B, et al. Abstract P2–10–03: genomic markers but not molecular subtypes provide prognostic impact and predict anthracycline efficacy in early triple-negative breast cancer: results from the prospective WSG PlanB trial. Cancer Res. 2018;78(4_supplement):P2-10-03-P12-10-03.
- Hudis CA, Barlow WE, Costantino JP, Gray RJ, Pritchard KI, Chapman JA, Sparano JA, Hunsberger S, Enos RA, Gelber RD, et al. Proposal for standardized definitions for efficacy end points in adjuvant breast cancer trials: the STEEP system. J Clin Oncol. 2007;25(15):2127–32.
- Denkert C, Seither F, Schneeweiss A, Link T, Blohmer J-U, Just M, Wimberger P, Forberger A, Tesch H, Jackisch C, et al. Clinical and molecular characteristics of HER2-low-positive breast cancer: pooled analysis of individual patient data from four prospective, neoadjuvant clinical trials. Lancet Oncol. 2021;22(8):1151–61.
- Xia LY, Cao XC, Yu Y. Survival outcomes in HER2-low versus HER2-zero breast cancer after neoadjuvant chemotherapy: a meta-analysis. World J Surg Oncol. 2024;22(1):106.
- Tarantino P, Jin Q, Tayob N, Jeselsohn RM, Schnitt SJ, Vincuilla J, Parker T, Tyekucheva S, Li T, Lin NU, et al. Prognostic and biologic significance of ERBB2-low expression in early-stage breast cancer. JAMA Oncol. 2022;8(8):1177–83.

- Roy AM, Jiang C, Perimbeti S, Deng L, Shapiro CL, Gandhi S. Oncotype Dx score, HER2 low expression, and clinical outcomes in early-stage breast cancer: a national cancer database analysis. Cancers. 2023;15(17):4264.
- Chaudhary LN, Jorns JM, Sun Y, Cheng YC, Kamaraju S, Burfeind J, Gonyo MB, Kong AL, Patten C, Yen T, et al. Frequent upregulation of HER2 protein in hormone-receptor-positive HER2-negative breast cancer after short-term neoadjuvant endocrine therapy. Breast Cancer Res Treat. 2023;201(3):387–96.
- 29. Bar Y, Dedeoglu AS, Fell GG, Moffett NJ, Boyraz B, Ly A, Bardia A, Moy B, Ellisen LW, Isakoff SJ. Dynamic HER2-low status among patients with triple negative breast cancer (TNBC): the impact of repeat biopsies. J Clin Oncol. 2023;41(16_suppl):1005–1005.
- Diep S, Maddukuri M, Yamauchi S, Geshow G, Delk NA. Interleukin-1 and nuclear factor Kappa B signaling promote breast cancer progression and treatment resistance. Cells. 2022;11(10):1673.
- 31. Curigliano G, Hu X, Dent RA, Yonemori K, Carlos H, Barrios S, Ohaughnessy J, Wildiers H, Zhang Q, Im SA, Saura C, et al. Trastuzumab deruxtecan (T-DXd) vs physician's choice of chemotherapy (TPC) in patients (pts) with hormone receptor-positive (HR+), human epidermal growth factor receptor 2 (HER2)-low or HER2-ultralow metastatic breast cancer (mBC) with prior endocrine therapy (ET): primary results from DESTINY-Breast06 (DB-06). J Clin Oncol. 2024;42(17_suppl):LBA1000.
- Banerji U, van Herpen CML, Saura C, Thistlethwaite F, Lord S, Moreno V, Macpherson IR, Boni V, Rolfo C, de Vries EGE, et al. Trastuzumab duocarmazine in locally advanced and metastatic solid tumours and HER2expressing breast cancer: a phase 1 dose-escalation and dose-expansion study. Lancet Oncol. 2019;20(8):1124–35.
- Zou Y, Yang A, Chen B, Deng X, Xie J, Dai D, Zhang J, Tang H, Wu T, Zhou Z, et al. crVDAC3 alleviates ferroptosis by impeding HSPB1 ubiquitination and confers trastuzumab deruxtecan resistance in HER2-low breast cancer. Drug Resist Updat. 2024;77: 101126.
- Marchiò C, Annaratone L, Marques A, Casorzo L, Berrino E, Sapino A. Evolving concepts in HER2 evaluation in breast cancer: Heterogeneity, HER2-low carcinomas and beyond. Semin Cancer Biol. 2021;72:123–35.
- Venetis K, Crimini E, Sajjadi E, Corti C, Guerini-Rocco E, Viale G, Curigliano G, Criscitiello C, Fusco N. HER2 low, ultra-low, and novel complementary biomarkers: expanding the spectrum of HER2 positivity in breast cancer. Front Mol Biosci. 2022;9: 834651.
- Corti C, Giugliano F, Nicolò E, Tarantino P, Criscitiello C, Curigliano G. HER2-low breast cancer: a new subtype? Curr Treat Options Oncol. 2023;24(5):468–78.
- 37. Rüschoff J, Penner A, Ellis IO, Hammond MEH, Lebeau A, Osamura RY, Penault-Llorca F, Rojo F, Desai C, Moh A *et al.* Global study on the accuracy of human epidermal growth factor receptor 2-low diagnosis in breast cancer. *Arch Pathol Lab Med* 2024.
- Viale G, Basik M, Niikura N, Tokunaga E, Brucker S, Penault-Llorca F, Hayashi N, Sohn J, Teixeira-de-Sousa R, Brufsky AM, et al. Retrospective study to estimate the prevalence and describe the clinicopathological characteristics, treatments received, and outcomes of HER2-low breast cancer. ESMO Open. 2023;8(4):101615.
- 39. Zaakouk M, Quinn C, Provenzano E, Boyd C, Callagy G, Elsheikh S, Flint J, Millican-Slater R, Gunavardhan A, Mir Y, et al. Concordance of HER2-low scoring in breast carcinoma among expert pathologists in the United Kingdom and the republic of Ireland -on behalf of the UK national coordinating committee for breast pathology. Breast. 2023;70:82–91.
- Lu Y, Zhu S, Tong Y, Fei X, Jiang W, Shen K, Chen X. HER2-low status is not accurate in breast cancer core needle biopsy samples: an analysis of 5610 consecutive patients. Cancers (Basel). 2022;14(24):6200.
- Molinelli C, Jacobs F, Agostinetto E, Nader-Marta G, Ceppi M, Bruzzone M, Blondeaux E, Schettini F, Prat A, Viale G, et al. Prognostic value of HER2low status in breast cancer: a systematic review and meta-analysis. ESMO Open. 2023;8(4): 101592.
- 42. Schettini F, Chic N, Brasó-Maristany F, Paré L, Pascual T, Conte B, Martínez-Sáez O, Adamo B, Vidal M, Barnadas E, et al. Clinical, pathological, and PAM50 gene expression features of HER2-low breast cancer. NPJ Breast Cancer. 2021;7(1):1.
- 43. Won HS, Ahn J, Kim Y, Kim JS, Song JY, Kim HK, Lee J, Park HK, Kim YS. Clinical significance of HER2-low expression in early breast cancer: a nationwide study from the Korean Breast Cancer Society. Breast Cancer Res. 2022;24(1):22.

- Page 15 of 15
- Wei T, Kang Y, Wang X, Yue J, Xu B, Yuan P. Prognostic value and relapse pattern of HER2-low in hormone receptor-positive breast cancer. Thorac Cancer. 2024;15(7):550–8.
- Peiffer D, Howard FM, Chen N, Hahn OM, Nanda R, Olopade OI, Huo D. Abstract HER2–11: HER2–11 epidemiology and prognosis of HER2-Low Breast Cancer (BC) in the National Cancer Data Base (NCDB). Cancer Res. 2023;83(5_supplement):HER2-11-HER12-11.

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