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Abstract: In Switzerland, viruses belonging to two different phylogenetic groups of small ruminant lentiviruses (SRLV) are currently circulating: the caprine arthritis-encephalitis virus (CAEV) and visna/maedi virus (VMV). In the past two decades, a mandatory national control program has led to a very low prevalence of seropositivity, while completely eliminating CAE as a clinical manifestation. However, in order to reduce the high costs and effort associated with this program, adjustments based on the most recent epidemiological knowledge are needed. The purpose of this study was to estimate the seroprevalence of CAEV and VMV using the newest diagnostic tools available, and to identify potential risk factors for infection with these viruses in Switzerland. For the prevalence estimation, a census was carried out including 10,696 farms with a total of 85,454 goats. Blood samples were analysed using a 3-step serological testing algorithm consisting of Chekit ELISA, Western Blot and SU5 ELISA. A risk factor analysis was conducted using logistic regression models built with data obtained from a mail questionnaire, and serological results from the census. The apparent herd-level prevalences were 0.38%, 2.77%, and 3.04% for CAEV, VMV and SRLV, respectively. Animal-level prevalences were 0.06% for CAEV, 0.55% for VMV, and 0.61% for SRLV. No statistically significant risk factors associated with CAEV or VMV infection were identified. However, the proportional high number of CAEV seropositive dwarf goats, in relation to their population size, could indicate that these hobby breeds may slip through some of the official controls. For an infection with SRLV, a medium herd size (7-40 goats) was found to be protective, compared with smaller (OR=1.90, p=0.034) and larger herds (OR=1.95, p=0.038). In conclusion, considering that all CAEV positive animals were culled, these results imply that CAEV is no longer actively spreading and has successfully been controlled in Switzerland. However, given the uncertain pathogenic potential of VMV in goats, future surveillance should also be taking into account the not insignificant number of VMV circulating in the Swiss goat population.

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Bern, 13 December 2016

Dear Editorial team,

Enclosed, please find the revised manuscript "A census to determine the prevalence and risk factors for caprine arthritis-encephalitis virus and visna/maedi virus in the Swiss goat population" by B. Thomann, L. C. Falzon, G. Bertoni, H.R. Vogt, G. Schüpbach-Regula, and I. Magouras.

We thank for valuable comments, which have substantially contributed to comprehension and clarity of this manuscript.

We respond below to each point brought up during the reviewing process.

On behalf of the authors,

Beat Thomann

Revision Note:

Dear Reviewer,

Thank you for your valuable comments on the analyses of dwarf goat farms. This helped us to identify a mistake (comparison of farm sizes of dwarf and "non-dwarf goat" farms) that led to a potential misunderstanding of the entire section. We corrected this mistake and also adjusted the methods and results section, which should have improved overall comprehension and clarity.

Reviewer #2: this is much improved although the English is a bit poor in places.

One concern I still have is that the authors are still focusing in on 7 seropositive dwarf goats for which they have done further analysis without presenting initial population data. These results are done as a proportion of positive goats ie not using a population denominator and they then analyse farms with dwarf goats as a proportion of positive farms and report that these farms were smaller than other CAEV positive farms etc but the analysis is not described in the methods and I cannot see these descriptive data or what was tested to develop the final model. These analyses are based on data from 32 farms (line 215), and contrast to their logistic model where other variables were significant or near significant at p 0.05, it raises the question of how this model was built and which tested e.g. purchase of female goats.

Only the descriptive statistics of the CAEV seropositive goats (former Table 5, now Table 4) is based on the responses (n=34) of the additional questions on individual goats (as described in lines 130-133). All analyses for dwarf goat farms were conducted on a farm level with the same, full dataset that was used for the CAEV risk factor logistic modeling (n=163).

Thanks to the reviewer's comments, we identified a mistake in the manuscript: Dwarf goat farms (n=24, independent of CAEV status) were not smaller than other CAEV positive farms (as stated in the previous version of the manuscript: "Dwarf goat farms were smaller (p<0.001) than other CAEV positive farms." [Revision 1, lines 219-220]), but they were smaller than "non-dwarf goat" farms (n= 139, independent of CAEV status). Thank you for pointing this out! We understand that this mistake has led to confusion on the whole sections of dwarf goat analyses and have corrected and rewritten this paragraph:

"Of the 47 CAEV seropositive goats detected during the census, detailed data on 34 goats located on 32 farms were reported through the questionnaire and available for descriptive statistics (Table 4). Seven of the 34 (20.6%) CAEV positive goats were dwarf goats, whose overall population size (Zwergziegen-IG, 2016) is substantially smaller compared with the population of the main goat breeds in Switzerland, such as Chamois colored or Saanen (SZZV, 2016). Dwarf goat farms (n=24) were smaller (median=8, p<0.001) than "non-dwarf goat" farms (n=139, median=16). Furthermore, fewer (66.7%, p=0.002) dwarf goat owners were professional farmers, compared to owners of other goat breeds (90.0%). Only 37.5% (n=9) of the dwarf goat farms had been tested for SRLV before 2012, compared to 56.8% (n=79, p=0.079) of the farms without dwarf goats." [lines 217-226]

Given that variables such as not tested before 2012 is not significant in the full model and is also 0.08 significance in the model of 32 farms, small farms are more likely to have dwarf goats but these are protective for SRLV positivity (Table 3 supplementary material) what does ia proportional risk model mean? after all these are all positive farms. I still believe there is insufficient logic to present this section of results.

As stated above statistics on dwarf goat farms were not conducted with the 32 CAEV positive farms only, but the same dataset as for the CAEV risk factor analysis (n=163). In the "full model" as the reviewer refers to, "tested before 2012" was checked for an association with CAEV seropositive farm status. In the "additional statistical analyses" the association between "dwarf goat farm" and "tested before 2012" was investigated, but with the same dataset. This methodology was not described in the methods section, and has now been added:

"For the additional statistical analyses on dwarf goat farms, the same data set as for the CAEV risk factor analysis was used. Farm size comparisons are based on differences in medians (Wilcoxon rank-sum test), whereas a chi-square test was used to analyse educational level of the farmers and testing for SRLV before 2012." [lines 158-161]

Thank you a lot for pointing this out!

Former Supplementary Table III (now Table 3) does not indicate that small farms are protective for SRLV positivity. (OR: 1.90, p=0.034).

We believe that by correcting the mistake mentioned above and by adjusting the methods [lines 158-161] and results section [lines 216-226], the additional analyses conducted with dwarf goat farms should now be clear, correct and justified.

tables are muddled and need some reflection -

Table 1 is fine but needs number of herds and animals testd in the title Table 2 and 3, why put these in the main text and not supplementary table 3 which is the model for these data Table 4, need number of herds and case control in title under cases and control this is presumably number, given there are many more controls than cases a % is needed to compare the relative proportions Table 5 - is this the data used ot build the model in 3.2.2 why are other variables not used? is the model appropriate etc

- The title of Table 1 has been reworded: "Herd-level (n=10,696) and animal-level (n=85,454) seroprevalences (95% CI) for CAEV, VMV and SRLV."
- Tables 2 and 3 have been moved to the supplementary material (now Supplementary Table I and II). Former Table 3 (now Supp. Table II) shows only descriptive statistics of continuous variables. The data used were from all questionnaire responses (n=341), as described in line 194-196.
- Table 4 (now Table 2) was adjusted according to the suggestions, the title has been updated and proportions (%) were added to the table: "Univariable analysis of variables tested for their association with CAEV seropositivity in case (n=32) and control (n=131) herds."
- Table 5 (now Table 4): This data was not used for any modeling but for characterization purposes only. All logistic regression analyses are based on herd-level data, including the analyses for farms that hold dwarf goats.

supp Table 1 needs number of herds etc, i am not sure this table is needed at all supp Table II again needs numbers time periods etc in title i question if supp III should be in the main text

Thank you for these suggestions. The tables have been adjusted accordingly.

- Supplementary Table 1 has been removed from the manuscript.
- The title of Supplementary Table II (now Supp. Table III) has been reworded and proportions (%) have been added to the table: "Univariable analysis of variables tested for their association with VMV seropositivity in case (n=100) and control (n=131) herds."
- Supplementary Table III has been moved to the main text (now Table 3).

1 2	1 2	A census to determine the prevalence and risk factors for caprine arthritis-encephalitis virus and visna/maedi virus in the Swiss goat population
3 4 5 6 7 8 9 10 11 12 13 14	3 4	B. Thomann ¹ *, L. C. Falzon ¹ , G. Bertoni ² , H.R. Vogt ³ , G. Schüpbach-Regula ¹ , I. Magouras ¹
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1 Abstract

2 In Switzerland, viruses belonging to two different phylogenetic groups of small ruminant 3 lentiviruses (SRLV) are currently circulating: the caprine arthritis-encephalitis virus (CAEV) 4 and visna/maedi virus (VMV). In the past two decades, a mandatory national control program has led to a very low prevalence of seropositivity, while completely eliminating CAE as a 5 clinical manifestation. However, in order to reduce the high costs and effort associated with 6 7 this program, adjustments based on the most recent epidemiological knowledge are needed. The purpose of this study was to estimate the seroprevalence of CAEV and VMV using the 8 9 newest diagnostic tools available, and to identify potential risk factors for infection with these viruses in Switzerland. For the prevalence estimation, a census was carried out including 10 10,696 farms with a total of 85,454 goats. Blood samples were analysed using a 3-step 11 12 serological testing algorithm consisting of Chekit ELISA, Western Blot and SU5 ELISA. A risk factor analysis was conducted using logistic regression models built with data obtained 13 from a mail questionnaire, and serological results from the census. The apparent herd-level 14 15 prevalences were 0.38%, 2.77%, and 3.04% for CAEV, VMV and SRLV, respectively. Animal-level prevalences were 0.06% for CAEV, 0.55% for VMV, and 0.61% for SRLV. No 16 statistically significant risk factors associated with CAEV or VMV infection were identified. 17 However, the proportional high number of CAEV seropositive dwarf goats, in relation to their 18 19 population size, could indicate that these hobby breeds may slip through some of the official controls. For an infection with SRLV, a medium herd size (7-40 goats) was found to be 20 21 protective, compared with smaller (OR=1.90, p=0.034) and larger herds (OR=1.95, p=0.038). In conclusion, considering that all CAEV positive animals were culled, these results imply 22 23 that CAEV is no longer actively spreading and has successfully been controlled in Switzerland. However, given the uncertain pathogenic potential of VMV in goats, future 24

surveillance should also be taking into account the not insignificant number of VMVcirculating in the Swiss goat population.

27 Keywords

- 28 Small ruminant lentivirus; Caprine Arthritis-Encephalitis Virus; Visna/maedi Virus;
- 29 Prevalence; Risk Factor; Census

30 **1. Introduction**

31 Caprine Arthritis-Encephalitis (CAE) is a widespread goat disease caused by the CAE-Virus (CAEV), a Lentivirus within the family of Retroviridae. Another virus of the same 32 33 genus is the Visna/Maedi Virus (VMV), which is responsible for the Visna-Maedi (VM) disease in sheep. Together, CAEV and VMV form the group of Small Ruminant Lentiviruses 34 (SRLV). Small Ruminant Lentiviruses, depending on the genotype, may cause chronic 35 36 inflammatory and degenerative lesions in the joints, mammary glands, lungs and brain of goats and sheep (Dawson, 1987; Narayan and Clements, 1989). Of the different SRLV 37 genotypes, B1 is considered as the prototype for CAEV, whereas genotype A1 is the 38 39 prototype for VMV. While B1 was the principal genotype circulating in Swiss goat herds before the eradication campaign, it seems that, at present, VMV A4 is the dominating SRLV 40 genotype (Cardinaux et al., 2013). From now on, the terms CAEV and VMV will be used to 41 42 refer to either genotypes B or genotypes A, respectively, to allow for better readability of the manuscript. 43

In Switzerland, a voluntary eradication program for CAE in goats was launched in 1984, 44 when seroprevalence in the Swiss goat population was around 60-80% (Peterhans et al., 45 2004). In 1998, eradication of CAE became mandatory, and in the same year the last 46 47 confirmed clinical CAE case in Switzerland was reported. From 2006 until 2011, the Swiss Federal Food Safety and Veterinary Office (FSVO) reported prevalences of 0.77% to 10.28% 48 (FSVO 2006 – 2011), based on an annual sampling of 25% of all goat farms. However, 49 50 previous diagnostic tests could not distinguish between CAEV and VMV (Zanoni, 1998), and 51 estimates published before 2012 therefore refer to infections with all SRLV, including goats 52 seropositive for VMV. Following the introduction of an additional diagnostic test which 53 permitted to distinguish between the two genotypes, in 2012 the Swiss federal authorities 54 decided to conduct a census and restrict the eradication campaign to the CAEV genotypes55 only.

Despite previous achievements and the ongoing eradication program, seroconversions, 56 mainly related to VMV, continue to occur in Switzerland. To reduce further transmission of 57 these viruses, it is important to understand which factors are associated with CAEV and VMV 58 59 infections in goats. The main routes of transmission of SRLV are either vertically, through 60 ingestion of infected colostrum or milk, or horizontally through inhalation of respiratory secretions, though intra-uterine infection can also occur (Blacklaws et al., 2004). In addition, 61 62 natural cross species transmission in mixed flocks from goat to sheep, and vice versa, has also been described (Germain et al., 2008; Leroux et al., 1997; Pisoni et al., 2005; Shah et al., 63 64 2004). Potential risk factors described in the literature include close contact to SRLV 65 seropositive sheep, the import of goats from abroad, the presence of seropositive bucks in a herd, as well as large herds (Brülisauer et al., 2005; Kaba et al., 2013). As mentioned 66 67 previously however, diagnostic tools available at the time when these studies were conducted could not distinguish between CAEV and VMV infections, and their results often refer to 68 69 SRLV in general. Given the advent of new diagnostic tools, a risk factor analysis for the separate genotypes could shed light on specific differences between the viruses. 70

The objectives of this study were to use the census data based on the new diagnostic test to (i) provide for the first time SRLV prevalence estimates in the Swiss goat population, discriminating between CAEV and VMV, and (ii) determine risk factors associated with CAEV and VMV infection in goats. Results from this study will permit the veterinary authorities to make evidence based decisions on future surveillance and control of this disease, thus optimizing resource allocation.

78 2. Materials and methods

79 2.1 Prevalence estimation

80

2.1.1 Census and laboratory analysis

81 As part of the national CAE control program, a census of all goat herds registered in Switzerland was conducted and coordinated by the FSVO. Goat herds were visited and 82 83 sampled by veterinarians between October 2011 and August 2012. The sampling unit was the single animal and blood samples (5-10ml whole blood) were taken from all goats that were 84 85 six months of age or older. The diagnostic procedure consisted of different serological 86 methods forming a 3-step serial testing algorithm (Figure 1). All blood samples were initially screened using Chekit CAEV/VMV Total Antibody Screening ELISA Test (Idexx 87 88 Switzerland AG, Liebefeld, Switzerland) by one of the ten accredited regional laboratories. 89 Positive and indeterminate sera were then sent to the National Reference Laboratory at the Institute of Veterinary Virology, University of Bern, where the same ELISA test was 90 91 repeated. If the ELISA positive result was confirmed, a Western Blot (WB) was performed, 92 and WB positive sera were then analysed using a panel of surface subunit (SU) 5 peptides derived from different SRLV genotypes (Bertoni et al., 2014; Mordasini et al., 2006). This 93 94 SU5 ELISA analysis permitted to distinguish between CAEV and VMV positive sera; 95 undifferentiated sera were classified as "CAEV positive by default".

96

2.1.2 Statistical analysis

97 The estimation of herd- and animal-level prevalences was based on the census data 98 and the confirmed seropositive results from the National Reference Laboratory. A herd was 99 considered CAEV or VMV seropositive if at least one goat had a corresponding SU5 ELISA 100 positive test result. All apparent prevalences (AP) were then converted into true prevalence 101 (TP) estimates by means of the Rogan and Gladen estimator (1978). In order to visualize 102 seropositive cases, a map with the spatial distribution of positive farms was designed using ArcGIS 10 (ESRI, 2010). Case farms were located based on their postal code. Information
about the density of small ruminants at the municipality level (FSVO) was mapped as a
background layer.

106 *2.2 Risk factor analysis*

107 *2.2.1 Study design and study population*

108 A case-control study was conducted to determine risk factors associated with a CAEV and 109 VMV positive farm status. For the cases, all farms that tested positive in the census were selected and three groups of cases were defined: (i) CAEV, (ii) VMV and (iii) SRLV. CAEV 110 111 cases were defined as farms that tested CAEV seropositive in the SU5 ELISA. All farms that 112 tested VMV positive, but without CAEV and "CAEV positive by default" results, were 113 treated as VMV cases. Finally, SRLV cases were all those farms that had at least either one 114 CAEV or VMV but had no "CAEV positive by default" test result. Controls were selected randomly from farms that tested negative in the census and had no history of SRLV 115 seropositivity in the previous five years (2006 until 2011). Since only 41 farms were tested 116 117 positive for CAEV and 284 farms positive for VMV, all these farms were included in our study. The number of control farms required was estimated using a logistic regression power 118 119 analysis conducted in PASS12 (Hintze, 2013), to detect an odds ratio (OR) for risk factors 120 greater than 2, with a power of 80% ($\alpha = 5\%$). Based on these calculations, a minimum of 120 121 controls was necessary.

122 2.2.2 Questionnaire

Epidemiological data from each case and control farm were collected through a standardized mail questionnaire. As the questionnaire was distributed to farmers in December 2013 none of the farms that tested positive in the 2012 CAE census were still under animal movement restrictions at the time the data were collected. The questionnaire consisted of 33 questions grouped into the following 5 categories: (i) farm type, breeds and number of animals; (ii)

128 farming know-how and education; (iii) contact with goats from other farms through trade, 129 breeding, shows or common pasture; (iv) contact with own or foreign sheep through common 130 pasture; (v) farm status for CAEV infection in the previous five years. Farmers with CAEV 131 positive goats were also asked to provide specific information on each CAEV positive goat. These additional seven questions, which aimed to gather information on the age, sex, breed, 132 133 origin, place of purchase and contact with sheep, were used for descriptive purposes only. The 134 questionnaire was pre-tested on goat owners to ensure clarity of the questions. To reduce bias 135 due to possible changes in management practices or increased disease awareness following 136 the census results, and to account for the time lag between when the census and the survey 137 were conducted, questions on animal movement or contact were phrased as "mean over the past two years", "on average", "normally" or "temporarily". The questionnaire was 138 139 distributed in German, French or Italian language and is available from the corresponding 140 author upon request.

141 *2.2.3 Statistical analyses*

The questionnaire data were transferred into an Excel spreadsheet (Microsoft Excel 2010) and analysed using STATA 13 (StataCorp., 2013). Predictor variables were checked for missing values and possible errors in data transcription, and continuous variables were also checked for linearity and restructured if needed.

To identify possible risk factors, the three case groups, (i) CAEV, (ii) VMV, and (iii) SRLV, were defined as binary outcomes (positive or negative) in the model-building process. For each of the three independent models, the corresponding data set was used and univariable associations between the putative risk factors as independent variables and the serological status were tested using logistic regression. Univariable ORs and p values were calculated and ranked. Independent variables that were significantly associated with outcomes at a liberal p value of <0.20 were assessed for co-linearity (Pearson correlation coefficient 153 >0.8) and defined as candidate risk factors. The multivariable logistic regression models were built using a stepwise backward elimination procedure, initially including all candidate risk 154 155 factor as independent variables. Based on likelihood-ratio tests, candidate risk factors that 156 were significant (p < 0.05) were retained in the final model and were tested for two-way 157 interactions (p < 0.05). The goodness-of-fit of the final models were tested using the Hosmer-158 Lemeshow test. For the additional statistical analyses on dwarf goat farms, the same data set 159 as for the CAEV risk factor analysis was used. Farm size comparisons are based on 160 differences in medians (Wilcoxon rank-sum test), whereas a chi-square test was used to 161 analyse educational level of the farmers and testing for SRLV before 2012.

162 **3. Results**

163

3.1 Seroprevalence and spatial distribution

164 Within the framework of the census, 85,454 individual goats located on 10,696 different farms were tested. A total of 729 animals from 452 farms were classified as WB 165 166 positive, of which 47 (6.4%) animals were further classified as CAEV infected and 473 167 (64.9%) as VMV infected by means of the SU5 ELISA. The remaining 209 (28.7%) WB 168 positive sera were classified as "CAEV positive by default" as they could not be further described using this panel of SU5 peptides; these were therefore excluded from further 169 170 prevalence estimation. However, for the purposes of the national eradication program, these animals were considered as potentially CAEV infected and were therefore culled. 171

172 The 47 CAEV positive goats were located on 41 farms. Of these farms, 36 (87.8%) 173 had 1 positive goat each, 4 farms had 2 positive goats, and 1 farm had 3 positive goats; the median within-herd prevalence was 5.9% (min=0.5%, max=66.7%). The 473 VMV infected 174 goats were located on 296 farms. The majority of these farms (78.7%) had only one VMV 175 176 seropositive goat. Twelve farms (4%) had both CAEV and VMV positive goats, resulting in a total of 325 SRLV positive farms. Consequently, the AP at a herd level were 0.38%, 2.77%, 177 178 and 3.04% for CAEV, VMV and SRLV, respectively, while the TP estimates were 0.40%, 179 32.21% and 32.27%. The animal-level AP were 0.06%, 0.55% and 0.61%, while TP estimates were 0.06%, 6.40% and 6.46% for CAEV, VMV and SRLV, respectively. All prevalence 180 181 estimates are listed in Table 1.

The 41 CAEV positive farms were evenly distributed over the whole territory of Switzerland and no regional patterns could be observed (Figure 2). Similarly, the VMV positive farms were detected all over Switzerland; however, regional patterns in accordance with the regional differences in animal density and topography, were noticeable for these farms.

187 *3.2 Risk factor analysis*

188

3.2.1 Study population and response rate

189 Based on the diagnostic test results, questionnaires were administered to all 452 farms 190 that tested positive with the WB, including the 127 "CAEV positive by default" farms. 191 However, the latter were only considered for descriptive purposes and later excluded from the 192 risk factor analysis. Overall, 752 questionnaires (including 300 for control farms) were sent 193 out, of which 13 could not be delivered. Of the 739 delivered questionnaires, 396 were 194 completed and returned, resulting in an overall response rate of 54%. Of these, 55 farms had 195 to be omitted either because the farm identification was missing (n=37) or because they no 196 longer kept goats (n=18). The response rate for CAEV positive farms was 78% (32 of 41). 197 Finally, a full data set was available for 341 farms. Descriptive statistics of the categorical and continuous variables of these farms are presented in Supplementary Table I and 198 199 Supplementary Table II, respectively.

200

3.2.2 Logistic regression

201 Based on the case definition, the risk factor analysis was conducted on 32 CAEV, 100 202 VMV and 125 SRLV case farms. A total of 131 controls were used for the risk factor analysis. The univariable analysis identified "purchase of female goats" (p=0.093, OR=2.17) 203 204 as the only predictor variable marginally associated (p < 0.10) with a positive CAEV status, 205 while the association between dwarf goat ownership and CAEV positive status was just above 206 the threshold value (p=0.208, OR=1.88). Thus, no multivariable model for CAEV could be 207 built. Results of the univariable analysis for other variables considered relevant are presented 208 in Table 2. The final multivariable model for VMV did not identify any significant risk 209 factors either. Nevertheless, four variables were marginally significant in the univariable 210 analysis: "herd size" (small vs. medium herds, OR=1.82; large vs. medium herds, OR=1.89) and "Appenzell breed" (OR=3.21) as candidate risk factors, while "know-how" (OR=0.53) 211

and "breeding done with own buck" (OR=0.62) as putative protective factors for VMV in goats (Supplementary Table III). For an infection with SRLV, the final model included the following two variables: "Appenzell breed" (OR=4.10) and "herd size" (small *vs.* medium herds, OR=1.90; large *vs* medium herds, OR=1.95) (Table 3).

216

3.3 Additional statistical analyses

217 Of the 47 CAEV seropositive goats detected during the census, detailed data on 34 goats 218 located on 32 farms were reported through the questionnaire and available for descriptive 219 statistics (Table 4). Seven of the 34 (20.6%) CAEV positive goats were dwarf goats, whose overall population size (Zwergziegen-IG, 2016) is substantially smaller compared with the 220 population of the main goat breeds in Switzerland, such as Chamois colored or Saanen 221 222 (SZZV, 2016). Dwarf goat farms (n=24) were smaller (median=8, p<0.001) than "non-dwarf" 223 goat" farms (n=139, median=16). Furthermore, fewer (66.7%, p=0.002) dwarf goat owners 224 were professional farmers, compared to owners of other goat breeds (90.0%). Only 37.5% 225 (n=9) of the dwarf goat farms had been tested for SRLV before 2012, compared to 56.8% (n=79, p=0.079) of the farms without dwarf goats. 226

227 4. Discussion

This is the first study in Switzerland where, by means of newly applied diagnostic tools, SRLV infected goats could be further classified as CAEV (genotypes B) and VMV (genotypes A) infected. This, in turn, allowed for genotype specific prevalence estimations and risk factor analyses. In addition, the prevalence estimations published in this study are based on a census, compared to earlier Swiss publications.

233 Only 47 CAEV positive goats (TP animal-level 0.06%), located on 41 farms (TP herd-234 level 0.40%) were detected, indicating that the eradication campaign is progressing 235 successfully. A comparison with previous publications should be done with caution as, due to 236 the lack of discriminatory tests for the different genotypes until recently, as they refer to a 237 global SRLV seroprevalence. Furthermore, different studies reported either AP or TP, and 238 should therefore not be compared directly. The herd-level AP for SRLV described in this 239 study (3.04%) differs from previous estimates for Switzerland (0.77% - 10.28%, FSVO 2006 240 - 2011); it is higher compared to that reported for 2006 and 2007, but lower than that reported 241 between 2008 and 2011. However, the estimates in 2008 and 2009 could have increased due 242 to the ongoing bluetongue vaccination campaign (FSVO, 2009, 2010), which might have 243 generated false positive results in the Screening ELISA Test (Valas et al., 2011). For the years 244 2010 and 2011, the reported estimates are considered as not representative as either the 245 sample sizes were too small or the samples were not taken randomly (FSVO, 2011, 2012). 246 The large difference between SRLV AP and TP estimates in this study is driven by the poor 247 sensitivity of the Screening ELISA for VMV genotypes, which comprise the vast majority 248 (91%) of SRLV seropositive goats detected during the census, and resulted a combined 249 overall sensitivity of 8.6% (Cardinaux et al., 2013). Since the SRLV positive results could not 250 be subdivided into CAEV or VMV prior to the 2012 census, it remains unclear how much of 251 the previous SRLV estimates were due to VMV cases. VMV seropositive goats are currently

excluded from mandatory culling as VMV infection in goats is still considered as nonpathogenic (despite recent histological evidence by Deubelbeiss et al., (2014) that VMV could cause mastitis) and VMV prevalence may thus rise in the coming years. Considering the fact that all goats that tested CAEV positive were culled, it might be assumed that VMV prevalence (TP herd-level: 32.21%, TP animal-level: 6.40%) currently equals SRLV prevalence.

A considerable number of WB positive animals escaping SU5 ELISA classification (28.7%) were reported by the National Reference Laboratory as "CAEV positive by default" and therefore culled. This decision was taken to reduce the risk that infected animals with false negative SU5 ELISA results may jeopardize the success of the eradication campaign. Previous work has shown that the sensitivity of the SU5 ELISA test is excellent with well characterized viruses but may fail with variant strains, leading to possible false negative cases (Bertoni et al., 2014).

During the 2012 census, numerous farms were tested for SRLV for the first time, and several of these were hobby farms that were not previously registered. It is plausible that dwarf goats are held on such farms, with potentially less professional management practices. Even though, before the 2012 census, the dwarf goat farms were not tested significantly less frequently compared with other farms (p=0.079), a trend is identifiable. This might be a possible explanation for the proportionally high number of CAEV seropositive dwarf goats detected.

Since only few publications distinguish between CAEV and VMV, knowledge on the transmission routes for each genotype is limited. Vertical transmission through colostrum and milk appears to be the main infection route for CAEV-infected goats. The situation in sheep appears to be more complex, with horizontal transmission of SRLV playing a more important role (Alvarez et al., 2005; Bertoni et al., 2014; Peterhans et al., 2004). This indicates that

277 alternative control measures might be necessary when aiming to control SRLV in sheep 278 populations. In CAEV and VMV co-infected goats, there is evidence that VMV is more 279 efficiently transmitted to the suckling kids, suggesting a rapid spread of VMV in goat 280 populations (Pisoni et al., 2010). Further research is needed to understand how efficiently CAEV is transmitted horizontally, both within goat herds, as well as between goats and sheep. 281 282 A low horizontal transmission rate of CAEV could explain the presence of only a single 283 seropositive goat in each CAEV case farm, as well as the even and almost random distribution 284 of CAEV cases over the country. Even if some CAEV positive goats have escaped detection due to imperfect diagnostic tests, we expect a very slow spread of the virus in the future 285 286 (unpublished data). In contrast, the regional patterns observed with VMV cases are suggestive 287 of a more efficient horizontal transmission of these viruses.

288 No significant risk factors for an infection with CAEV could be determined in this 289 study, which differs from other publications (Brülisauer et al., 2005; Kaba et al., 2013), 290 though the univariable regressions identified an association between CAEV positive status 291 and "purchase of female goats", similar to previous findings by Brülisauer et al. (2005). 292 However, as mentioned before, past studies refer to risk factors for an infection with SRLV 293 and not only with CAEV. Additionally, a major limitation in our risk factor analysis was the 294 small number of CAEV case farms (n=32) included (despite the high response rate). 295 Ironically, this fact can be seen as a negative "side effect" of the success of the eradication 296 campaign over the last two decades, which has led to a low prevalence. Furthermore, the 297 retrospective study design might have introduced bias due to potential improved disease 298 awareness and consequent changes in management practices following the census results.

Although no risk factors for an infection with VMV were determined, the univariable analysis identified seven variables below the threshold point (p<0.20). This difference in number of putative risk factors, compared to the CAEV analysis, may be attributed to the 302 higher number of VMV case farms (n=100) included in the analysis. SRLV risk factors 303 obtained from the present case-control study, similar to the SRLV prevalence, were strongly 304 influenced by the large proportion of VMV case farms. The observed higher odds (OR=1.90) 305 of SRLV seropositivity in small herds, compared to medium sized herds, could be explained 306 by less professional management practices often observed on such smaller farms. On the other 307 hand, large herds could be at a higher odds (OR=1.95) of infection due to greater animal 308 movements which may favor disease transmission (Kaba et al., 2013). Goats from the 309 Appenzell breed were found to be a significant risk factor for SRLV (OR=4.10), and this 310 tendency was also observed with VMV positive status. However, only a small number of 311 participating farms (n=12) held goats from the Appenzell breed.

312 Conclusion

313 While the disappearance of clinical cases of CAEV-induced arthritis in Switzerland 314 suggested that the eradication campaign had succeeded in eliminating virulent CAEV strains 315 from the goat population, this study, based on a detailed epidemiological analysis of a 316 serological survey involving the entire goat population, further confirms and expands this 317 observation. Therefore we believe that classical CAEV genotypes are no longer circulating 318 between goats in Switzerland, permitting us to conclude that these viruses may be considered 319 as eradicated. In contrast, it seems that VMV genotypes may be actively transmitted in goat 320 herds, pointing to the importance of regularly monitoring these infections, particularly since 321 their pathogenic potential is still uncertain. In this respect, we are currently developing 322 epidemiological models that will permit to perform this monitoring in a targeted and 323 economically efficient way.

325 **Conflicts of interest**

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332 **References**

- Alvarez, V., Arranz, J., Daltabuit-Test, M., Leginagoikoa, I., Juste, R.A., Amorena, B., de
 Andrés, D., Luján, L.L., Badiola, J.J., Berriatua, E., 2005. Relative contribution of
 colostrum from Maedi-Visna virus (MVV) infected ewes to MVV-seroprevalence in
 lambs. Res. Vet. Sci. 78, 237–43. doi:10.1016/j.rvsc.2004.09.006
- Bertoni, G., Cardinaux, L., Deubelbeiss, M., Zahno, M.-L., Vogt, H.-R., 2014. SU5 serology
 as a novel tool to support a challenging caprine arthritis encephalitis (CAEV) eradication
 campaign., in: Rackwitz, R., Pees, M., Aschenbach, J.R., Gäbel, G. (Eds.), 7. Leipziger
- 340Tierärztekongress. University of Leipzig, Leipzig. pp. 229–232.
- 341 Blacklaws, B.A., Berriatua, E., Torsteinsdottir, S., Watt, N.J., de Andres, D., Klein, D.,
- Harkiss, G.D., 2004. Transmission of small ruminant lentiviruses. Vet. Microbiol. 101,
 199–208. doi:10.1016/j.vetmic.2004.04.006
- Brülisauer, F., Vogt, H.-R., Perler, L., Rüfenacht, J., 2005. Risk factors for the infection of
 Swiss goat herds with small ruminant lentivirus: a case-control study. Vet. Rec. 157,
 229–233. doi:10.1136/vr.157.8.229
- Cardinaux, L., Zahno, M.-L., Deubelbeiss, M., Zanoni, R., Vogt, H.-R., Bertoni, G., 2013.
 Virological and phylogenetic characterization of attenuated small ruminant lentivirus
 isolates eluding efficient serological detection. Vet. Microbiol. 162, 572–81.
 doi:10.1016/j.vetmic.2012.11.017
- 351 Dawson, M., 1987. Pathogenesis of maedi-visna. Vet. Rec. 120, 451–4.
- Deubelbeiss, M., Blatti-Cardinaux, L., Zahno, M.-L., Zanoni, R., Vogt, H.-R., Posthaus, H.,
 Bertoni, G., 2014. Characterization of small ruminant lentivirus A4 subtype isolates and
 assessment of their pathogenic potential in naturally infected goats. Virol. J. 11, 65.

doi:10.1186/1743-422X-11-65

356 ESRI, 2010. ArcGIS, Version 10.1.

- 357 FSVO, 2012. Jahresbericht Seuchenfreiheit 2011 [WWW Document]. URL
 358 http://www.blv.admin.ch/dokumentation/04506/04518/index.html?lang=de (accessed
 359 4.26.16).
- 360 FSVO, 2011. Jahresbericht Seuchenfreiheit 2010 [WWW Document]. URL
 361 http://www.blv.admin.ch/dokumentation/04506/04518/index.html?lang=de (accessed
 362 4.26.16).
- 363 FSVO, 2010. Jahresbericht Seuchenfreiheit 2009 [WWW Document]. URL
 364 http://www.blv.admin.ch/dokumentation/04506/04518/index.html?lang=de (accessed
 365 4.26.16).
- 366 FSVO, 2009. Jahresbericht Seuchenfreiheit 2008 [WWW Document]. URL
 367 http://www.blv.admin.ch/dokumentation/04506/04518/index.html?lang=de (accessed
 368 4.26.16).
- Germain, K., Croise, B., Valas, S., 2008. Field evaluation of a gag/env heteroduplex mobility
 assay for genetic subtyping of small-ruminant lentiviruses. J. Gen. Virol. 89, 2020–8.
 doi:10.1099/vir.0.2008/000851-0
- 372 Hintze, J., 2013. PASS 12. NCSS, LLC.
- Kaba, J., Czopowicz, M., Ganter, M., Nowicki, M., Witkowski, L., Nowicka, D., SzaluśJordanow, O., 2013. Risk factors associated with seropositivity to small ruminant
 lentiviruses in goat herds. Res. Vet. Sci. 94, 225–7. doi:10.1016/j.rvsc.2012.09.018
- 276 Leroux, C., Chastang, J., Greenland, T., Mornex, J.F., 1997. Genomic heterogeneity of small

- 377 ruminant lentiviruses: existence of heterogeneous populations in sheep and of the same
 378 lentiviral genotypes in sheep and goats. Arch. Virol. 142, 1125–37.
- 379 Mordasini, F., Vogt, H.-R., Zahno, M.-L., Maeschli, A., Nenci, C., Zanoni, R., Peterhans, E.,

Bertoni, G., 2006. Analysis of the antibody response to an immunodominant epitope of

- the envelope glycoprotein of a lentivirus and its diagnostic potential. J. Clin. Microbiol.
- 382 44, 981–91. doi:10.1128/JCM.44.3.981-991.2006
- 383 Narayan, O., Clements, J.E., 1989. Biology and Pathogenesis of Lentiviruses. J. Gen. Virol.
 384 70, 1617–1639. doi:10.1099/0022-1317-70-7-1617
- 385 Peterhans, E., Greenland, T., Badiola, J., Harkiss, G., Bertoni, G., Amorena, B., Eliaszewicz,
- 386 M., Juste, R.A., Krassnig, R., Lafont, J.-P., Lenihan, P., Pétursson, G., Pritchard, G.,
- Thorley, J., Vitu, C., Mornex, J.-F., Pépin, M., 2004. Routes of transmission and
 consequences of small ruminant lentiviruses (SRLVs) infection and eradication schemes.

389Vet. Res. 35, 257–74. doi:10.1051/vetres:2004014

- Pisoni, G., Bertoni, G., Manarolla, G., Vogt, H.-R., Scaccabarozzi, L., Locatelli, C., Moroni,
 P., 2010. Genetic analysis of small ruminant lentiviruses following lactogenic
- **392** transmission. Virology 407, 91–9. doi:10.1016/j.virol.2010.08.004
- Pisoni, G., Quasso, A., Moroni, P., 2005. Phylogenetic analysis of small-ruminant lentivirus
 subtype B1 in mixed flocks: evidence for natural transmission from goats to sheep.
 Virology 339, 147–52. doi:10.1016/j.virol.2005.06.013
- Rogan, W.J., Gladen, B., 1978. Estimating Prevalence from the Results of a Screening Test.
 Am. J. Epidemiol. 107, 71–76.
- Shah, C., Huder, J.B., Böni, J., Schönmann, M., Mühlherr, J., Lutz, H., Schüpbach, J., 2004.
- 399 Direct evidence for natural transmission of small-ruminant lentiviruses of subtype A4

400 from goats to sheep and vice versa. J. Virol. 78, 7518–22. doi:10.1128/JVI.78.14.7518401 7522.2004

402 StataCorp., 2013. Stata Statistical Software: Release 13.

403 SZZV, 2016. Herdebuchbestand 2016 [WWW Document]. URL
404 http://szzv.caprovis.ch/files/Herdebuch/Herdebuchbestand 2016.pdf (accessed 10.17.16).

Valas, S., Le Ven, A., Croise, B., Maquigneau, M., Perrin, C., 2011. Interference of
vaccination against bluetongue virus serotypes 1 and 8 with serological diagnosis of
small-ruminant lentivirus infection. Clin. Vaccine Immunol. 18, 513–7.
doi:10.1128/CVI.00343-10

- Zanoni, R.G., 1998. Phylogenetic analysis of small ruminant lentiviruses. J. Gen. Virol. 79,
 1951–1961.
- 411 Zwergziegen-IG, 2016. Protokoll Generalversammlung 2016 [WWW Document]. URL
 412 http://www.zwergziegen-ig.ch/files/Protokoll-GV-2016.pdf (accessed 10.17.16).

Figure 1

- 3-step serological testing algorithm used to determine SRLV status with corresponding test
- sensitivities/specificities (%) for CAEV and VMV.
 - ¹CAEV: 97.9/98.1; VMV: 43.0/100
 - ²CAEV: 98.0/95.0; VMV: 35.0/100
 - ³CAEV: 99.0/99.0; VMV: 57.0/50.0

Figure 2

Spatial distribution of CAEV (n=41) and VMV (n=284) seropositive farms.

2 Herd-level (n=10,696) and animal-level (n=85,454) seroprevalences (95% CI) for CAEV,

3 VMV and SRLV.

	CAEV (genotypes B)	VMV (genotypes A)	SRLV (genotypes A and B)
Herd-level			
Number of seropositive herds	41	296	325 ^a
Apparent seroprevalence	0.38%	2.77%	3.04%
True seroprevalence	0.40%	32.21%	32.27%
-	(0.34-0.46)	(31.92-32.50)	(31.97-32.57)
Animal-level			
Number of seropositive animals	47	473	520
Apparent seroprevalence	0.06%	0.55%	0.61%
True animal seroprevalence	0.06%	6.40%	6.46%
	(0.06-0.07)	(6.35-6.44)	(6.41-6.51)

4 ^a 12 farms with both CAEV and VMV positive animals.

2 Univariable analysis of variables tested for their association with CAEV seropositivity in case (n=32) and control (n=131) herds.

X7 · 11	Description	Cases		Controls				Odds Ratio
Variable	Description	n	%	n	%	<i>p</i> value	Odds Ratio	(95% CI)
Animal show	Goats taken to breed shows or goat markets at least every other year	10	31.3	41	31.3	0.996	1.00	0.43-2.30
Breeding	Breeding of animals done on farm	27	84.4	116	88.5	0.520	0.70	0.23-2.09
Breed Appenzell	Appenzell breed present in herd	2	6.3	3	2.3	0.264	2.84	0.45-17.78
Breed dwarf goat	Dwarf goat breed present in herd	7	21.9	17	13.0	0.208	1.88	0.70-5.01
Contact alpine	Common alpine pasture with foreign goats/sheep	10	31.3	37	28.2	0.737	1.15	0.50-2.67
Contact sheep	Keep goats and sheep together with direct contact	7	21.9	21	16.0	0.434	1.47	0.56-3.83
Herd size						0.642		
	Medium herd (7-40 goats)	15	46.9	71	54.2	(ref)	1.00	-
	Small herd (1-6 goats)	8	25.0	33	25.2	0.777	1.15	0.44-2.97
	Large herd (>40 goats)	9	28.1	27	20.6	0.341	1.58	0.62-4.03
Know-how	Know-how based on agricultural education and/or membership in a goat association	27	84.4	114	87.0	0.695	0.81	0.27-2.38
Own buck	Breeding done with own buck(s)	22	68.8	87	66.4	0.801	1.11	0.48-2.55
Purchase	Purchase of at least one goat per year	15	46.9	59	45.0	0.852	1.08	0.50-2.34
Purchase female	Purchase of at least one female goat per year	9	28.1	20	15.3	0.093	2.17	0.88-5.37
Tested before 2012	Farm tested for SRLV before the 2012 census (2006-2011)	16	50.0	72	55.0	0.614	0.82	0.38-1.78
Wildlife						0.403		
	No observed contact with wildlife	24	75.0	98	74.8	(ref)	1.00	-
	Observed contact with ibex	3	9.4	5	3.8	0.241	2.45	0.55-10.97
	Observed contact with chamois and other Swiss wildlife	5	15.6	28	21.4	0.556	0.73	0.25-2.09

- Final multivariable logistic regression model of risk factors associated with small ruminant lentiviruses (SRLV) infection in goats. 2
- 3

Variable	Description	se	p value	Odds Ratio	Odds Ratio (95% CI)
Breed Appenzell	Appenzell breed present in herd	2.83	0.041	4.10	1.06-15.84
Herd size			0.039		
	Medium herd (7-40 goats)		(ref)	1.00	-
	Small herd (1-6 goats)	0.57	0.034	1.90	1.05-3.43
	Large herd (>40 goats)	0.63	0.038	1.95	1.04-3.66

2 Characterization of individual goats (n=34) that tested CAEV seropositive during the 2012

3 census.

Variable	Categories	n	%
Sex	Female	28	82
	Male	6	18
Origin	Own breeding	20	59
	Bought in	14	41
Place of purchase (n=14)	Private	14	100
	Market	0	0
Contact with sheep (n=33)	Yes	8	24
	No	20	61
	Not known	5	15
Breed	Dwarf goat	7	20
	Mixed breed	7	20
	Saanen	6	18
	Chamois colored	4	12
	Grisons striped	3	9
	Peacock goat	3	9
	Buren	2	6
	Valais Blackneck	1	3
	Nera Verzasca	1	3

- **Supplementary Table I** Descriptive statistics of categorical variables in a study to determine risk factors for SRLV seropositivity (n=341).

Variable	Description	Categories	n	%
Alpine pasture	Goats taken on alpine pasture in the summer	Yes No	134 207	39 61
Animal show	Goats taken to breed shows or goat markets at least every other year	Yes No	112 229	33 67
Breeding	Breeding of animals done on farm	Yes No	290 51	85 15
Contact alpine	Common alpine pasture with foreign goats/sheep	Yes No	106 235	31 69
Contact sheep	Keep goats and sheep together with direct contact	Yes No	64 277	19 81
Contact wildlife	Observed contact (<5meters) with wildlife animals	Yes No	89 252	26 74
Farm type	Goat farm production type	Milk Other	131 210	38 62
Know-how	Know-how based on agriculture education and/or goat association membership	Yes No	281 60	82 18
Purchase	Purchase of at least one goat per year	Yes No	142 199	42 58
Tested before 2012	Farm tested for SRLV before the 2012 census (2006-2011)	Yes No	191 150	56 44

1 Supplementary Table II

- 2 Characterization of continuous variables in a study to determine risk factors for SRLV
- **3** seropositivity (n=341).

Variable	Mean	Median	Min	Max
Number of goats per farm				
Overall (n=341)*	32.2	14	1	300
CAEV (n=32)	34.5	11	3	165
VMV (n=100)	36.7	12.5	2	300
SRLV (n=125)	32.2 34.5 36.7 35.4 30.0 1.9	12	2	300
Controls (n=131)	30.0	13	1	270
Number of bucks (n=341)	1.9	1	0	26
Number of purchased goats / year (n=142)†	2.7	1.5	0	60
Number of sheep (n=80)††	32.6	11.5	2	600

4 * including CAE positive by default tested farms

5 † only for farms purchasing goats

 $\dot{\dagger}$ $\dot{\dagger}$ only for those farms where sheep were also present on the farm

1

Supplementary Table III Univariable analysis of variables tested for their association with VMV seropositivity in case (n=100) and control (n=131) herds. 2

X7 · 11	Description	C	ases	Controls		1		Odds Ratio
Variable	Description	n	%	n	%	<i>p</i> value	Odds Ratio	(95% CI)
Animal show	Goats taken to breed shows or goat markets at least every other year	30	30.0	41	31.3	0.832	0.94	0.53-1.66
Breeding	Breeding of animals done on farm	82	82.0	116	88.5	0.162	0.59	0.28-1.24
Breed Appenzell	Appenzell breed present in herd	7	7.0	3	2.3	0.097	3.21	0.81-12.75
Breed dwarf goat	Dwarf goat breed present in herd	18	18.0	17	13.0	0.293	1.47	0.72-3.03
Contact alpine	Common alpine pasture with foreign goats/sheep	34	34.0	37	28.2	0.348	1.31	0.75-2.30
Contact sheep	Keep goats and sheep together with direct contact	21	21.0	21	16.0	0.333	1.39	0.71-2.72
Herd size						0.071		
	Medium herd (7-40 goats)	39	39.0	71	54.2	(ref)	1.00	-
	Small herd (1-6 goats)	33	33.0	33	25.2	0.059	1.82	0.98-3.39
	Large herd (>40 goats)	28	28.0	27	20.6	0.058	1.89	0.98-3.64
Know-how	Know-how based on agricultural education and/or membership in a goat association	78	78.0	114	87.0	0.072	0.53	0.26-1.06
Own buck	Breeding done with own buck(s)	55	55.0	87	66.4	0.078	0.62	0.36-1.06
Purchase	Purchase of at least one goat per year	36	36.0	59	45.0	0.167	0.69	0.40-1.17
Purchase female	Purchase of at least one female goat per year	18	18.0	20	15.3	0.579	1.22	0.61-2.45
Tested before 2012	Farm tested for SRLV before the 2012 census (2006-2011)	63	63.0	72	55.0	0.219	1.40	0.82-2.38
Wildlife						0.169		
	No observed contact with wildlife	70	70.0	98	74.8	(ref)	1.00	-
	Observed contact with ibex	10	10.0	5	3.8	0.071	2.80	0.92-8.55
	Observed contact with chamois and other Swiss wildlife	20	20.0	28	21.4	1.000	1.00	0.52-1.92



