

RESEARCH ARTICLE

Open Access



Genome-wide investigation of WRKY gene family in pineapple: evolution and expression profiles during development and stress

Tao Xie^{1,2}, Chengjie Chen^{1,2}, Chuhao Li^{1,2}, Jiarou Liu^{1,2}, Chaoyang Liu^{1,2*} and Yehua He^{1,2*}

Abstract

Background: WRKY proteins comprise a large family of transcription factors that play important roles in many aspects of physiological processes and adaptation to environment. However, little information was available about the WRKY genes in pineapple (*Ananas comosus*), an important tropical fruit. The recent release of the whole-genome sequence of pineapple allowed us to perform a genome-wide investigation into the organization and expression profiling of pineapple WRKY genes.

Results: In the present study, 54 pineapple WRKY (AcWRKY) genes were identified and renamed on the basis of their respective chromosome distribution. According to their structural and phylogenetic features, the 54 AcWRKYs were further classified into three main groups with several subgroups. The segmental duplication events played a major role in the expansion of pineapple WRKY gene family. Synteny analysis and phylogenetic comparison of group III WRKY genes provided deep insight into the evolutionary characteristics of pineapple WRKY genes. Expression profiles derived from transcriptome data and real-time quantitative PCR analysis exhibited distinct expression patterns of AcWRKY genes in various tissues and in response to different abiotic stress and hormonal treatments.

Conclusions: Fifty four WRKY genes were identified in pineapple and the structure of their encoded proteins, their evolutionary characteristics and expression patterns were examined in this study. This systematic analysis provided a foundation for further functional characterization of WRKY genes with an aim of pineapple crop improvement.

Keywords: Pineapple, WRKY, Genome-wide, Expression patterns, Abiotic stress

Background

The WRKY transcription factors (TFs) are one of the largest families in higher plants and are found throughout the green lineage [1]. The most prominent feature of the WRKY proteins is the 60 amino acid long WRKY domain, which comprises the highly conserved signature WRKYGQK followed by a C2H2- or C2HC-type of zinc-finger motif. Both heptapeptide sequence and zinc-finger motif are required for the high binding affinity of WRKY TFs to the consensus W-box cis-elements [2, 3]. Based on the number of WRKY domains and the

structure of their zinc-finger motifs, the WRKY proteins can be classified into three main groups (I–III), those with two WRKY domains belong to Group I, while those with one WRKY domain belong to Group II or III [4].

Since the first WRKY gene, SPF1, was cloned from sweet potato [5], a large number of WRKY proteins have been experimentally identified from various plant species. Substantial evidence indicates that the WRKY transcription factors are involved in plant defense regulatory networks, including response to various biotic and abiotic stresses [6, 7]. The WRKY TFs are one of the best-characterized classes of plant defense transcription factors and at the forefront of research on plant defense responses. The WRKY proteins play a key role in plant defense against biotic stresses including bacterial, fungal

* Correspondence: liuchaoyang@scau.edu.cn; heyehua@hotmail.com

¹Key Laboratory of Biology and Germplasm Enhancement of Horticultural Crops in South China, Ministry of Agriculture, South China Agricultural University, Guangzhou 510642, China

Full list of author information is available at the end of the article



and viral pathogens [8–10]. A large number of WRKY TFs are induced by abiotic stresses and take part in the regulation of plant tolerance to abiotic stress [11]. For example, VvWRKY11 from grapevine is involved in the response to dehydration stress [12]. AtWRKY75 is induced in the plant during phosphate deficiency while suppression of WRKY75 expression leads to the increased susceptibility to phosphate stress [13]. The WRKY proteins play central roles in various aspects of physiological processes, including senescence [14], seed dormancy and germination [15], embryogenesis [16], trichome development, modulation of flowering time [17, 18], fruit flavor [19], biosynthesis of secondary metabolites [20–22]. They are also key components in some signal transduction processes mediated by plant hormones such as abscisic acid (ABA) and salicylic acid (SA) [6]. The Arabidopsis WRKY39 positively regulates the cooperation between the SA- and JA-activated signaling pathways that mediate response to heat stress [23]. OsWRKY31 mediates crosstalk between auxin and the defense signal transductions [24].

The WRKY proteins are conserved in evolutionary history throughout the plant kingdom, and the expansions of this family seem to be related to the evolution and diversity of the plants [25]. Although WRKY TFs are considered as plant-specific, they were also reported in non-plant species, e.g. protist, slime mold, and unicellular algae [4]. The evolution of the WRKY gene family provide insights into how biotic and abiotic stress responses and signaling evolved as plant went from simpler, unicellular to more complex, multicellular flowering plants. The availability of increasing numbers of sequenced genomes has facilitated the evolutionary studies of the WRKY genes family, and large-scale genome-wide analysis of WRKY genes have been described in many different species, which would be helpful to understanding their evolutionary origin and biological functions.

Pineapple, *Ananas comosus* (L.) Merr., is the only species of the Bromeliaceae family grown commercially for its fruit [26]. After banana and citrus, it is the third most important tropical fruit in world production, and is found in almost all the tropical and subtropical regions of the world [27]. The environmental stresses significantly affect the growth and development of the pineapple plant. The pineapple fruit can be injured by sunburn under high temperature, the low temperature can result in diminished growth and the plant is usually severely damaged by frost [26]. To a certain extent the pineapple can survive under drought conditions owing to the plant morphological features and its crassulacean acid metabolism. Nevertheless, prolonged extreme droughts affect growth and yield dramatically. The biotic stresses like pests, diseases, and weeds can also cause

significant yield loss in plantation pineapple production. Additionally, pineapple is also an important species in the research of the monocot evolution owing to its pivotal phylogenetic position at the base of the order Poales [28]. The pineapple is cultivated worldwide and has great economic and research value and so there is considerable interest in identifying important functional genes.

The WRKY gene family has been extensively studied in many plant species. However, current basic knowledge of WRKY proteins in pineapple is still limited. Due to the importance of the WRKY genes in various physiological programs, it would be of interest to make a systematic investigation of the WRKY family in pineapple. Recent completion of the pineapple genome sequencing provided an opportunity to reveal the organization, expression and evolutionary traits of pineapple WRKY gene family at the genome-wide level [29]. In the present study, we identified 54 pineapple *WRKY* genes and classified them into three main groups. The comprehensive analysis including the exon-intron organization, motif compositions, gene duplications, chromosome distribution, phylogenetic and synteny analysis were further investigated. Global expression analysis was performed to identify involvement of specific WRKY gene family members in different biological processes in pineapple. This study provided valuable clues for functional characterization of WRKY gene family members in pineapple.

Results

Identification of the WRKY proteins in pineapple

A total of 56 candidate gene models corresponding to the Pfam WRKY family were originally obtained. The annotation of these gene models were further checked using available pineapple transcriptome data. Ten erroneously predicted WRKY gene models were manually curated and two redundant sequences (*Aco030791.1* and *Aco025780.1*) were then removed. Finally, 54 gene models were selected and annotated as being pineapple *WRKY* genes based on the presence of apparently complete WRKY domains. The validated *AcWRKY* gene sequences were available in Additional file 1. A total of 51 *WRKY* genes could be mapped on the linkage groups and were renamed from AcWRKY1 to AcWRKY51 based on their order on the linkage groups. Three *WRKY* genes (*Aco027950.1*, *Aco028684.1*, and *Aco031477.1*) that could not be conclusively mapped to any linkage groups were renamed AcWRKY52- AcWRKY54 respectively.

Gene characteristics, including the length of the CDS (Coding Sequence), the length of the protein sequence, the protein molecular weight (MW), isoelectric point (pI), and the subcellular localization were analyzed (Additional file 1). Among the 54 *AcWRKY* proteins,

AcWRKY14 was identified to be the smallest protein with 122 amino acid (aa), whereas the largest one was AcWRKY23 (1320 aa). The MW of the proteins ranged from 13.7 to 144.5 kDa, and the pI ranged from 5.11 (AcWRKY10) to 10.08 (AcWRKY54). The predicted sub-cellular localization results showed that 51 AcWRKY proteins were located in the nuclear region, whereas three proteins were located in the chloroplast.

Multiple sequence alignment, phylogenetic analysis, and classification of *AcWRKY* genes

The phylogenetic relationship of the AcWRKY proteins was examined by multiple sequence alignment of their WRKY domains, which span approximately 60 amino acids. The WRKY domain of seven different Arabidopsis WRKY proteins (ATWRKY58, 40, 61, 50, 74, 65, 54) from each of the groups or subgroups, were randomly selected as representatives for the further comparison. As shown in Fig. 1, the sequences in the WRKY domain were highly conserved. A total of 50 AcWRKY proteins were found to have the highly conserved sequence WRKYGQK, while the others (AcWRKY14, AcWRKY23, AcWRKY27 and AcWRKY43) vary by a single amino acid.

The phylogenetic analysis (Fig. 2) indicated that the pineapple WRKY domains could be divided into three large groups corresponding to group I, II and III in Arabidopsis as defined by Eulgem et al. [2]. Among the 54 AcWRKY proteins, 12 belong to group I, 34 to group II, and 8 to group III. The 14 members from group I all contained two WRKY domains and C2H2-type zinc-finger motifs (C-X₄-C-X₂₂₋₂₃-H-X-H), without the domain loss events which usually happened in some monocotyledonous species [30]. The N-terminal and C-terminal WRKY domains were clustered in different clades, which may reflect the parallel evolution of the two domains. The WRKY members in group II can be further clustered into five subgroups (IIa-IIe), two WRKY proteins belong to IIa, 7 to IIb, 13 to IIc, 7 to IId, and 5 to IIE. Seven of the eight members in group III contain the C2HC-type zinc fingers (C-X₇-C-X₂₃-H-X-C), whereas the remaining AcWRKY14 possess a C2H2 type of zinc finger motif. The extended WRKY domains, which were found in several group III WRKY proteins in some monocot species like rice and Brachypodium [30], were not observed in pineapple.

The 'leucine-rich repeat' (LRR) motif, typical domain of resistance (R) proteins, was detected in AcWRKY23 belonging to group IIc. AcWRKY23 could be further characterized as an R protein-WRKY gene in pineapple. The existence of such chimeric proteins was one of the unusual features of the WRKY gene family in flowering plants like Arabidopsis and rice [25]. Further phylogenetic and the protein architecture analysis for AcWRKY23

and several R protein-WRKY proteins in other species (Additional file 2) indicated that AcWRKY23 was not belong to any group that have been characterized in Rinerson et al. [25].

Gene structure and motif composition of pineapple WRKY gene family

The exon-intron organizations of all the identified *AcWRKY* genes were examined to gain more insight into the evolution of the WRKY family in pineapple. As shown in Fig. 3b, all *AcWRKY* genes possessed two to six exons (four with two exons, 27 with three exons, six with four exons, 11 with five exons, and six with six exons). Genes with only one exon were not observed. Genes within the same group usually have a similar structure, for example, all group IIe members contained three exons and two introns. Further analyses indicated that all *AcWRKY* genes contained an intron in their respective WRKY domains. The distribution of introns and the intron phase were coincident with the alignment clusters of *AcWRKY* genes. The V-type intron, a phase-0 intron, was only observed in group IIa and IIb. The R-type intron (a phase-1 intron) was widely distributed in all the other groups (group I, IIc, IId, IIE and III), similar with that in rice and Arabidopsis [25]. No introns were found in the N-terminal WRKY domains of genes belonging to group I.

A schematic representing the structure of all AcWRKY proteins was constructed from the MEME motif analysis results. As exhibited in Fig. 3c, other than motifs 1 and 2 which are the WRKY domains widely distributed, AcWRKY members within the same groups were usually found to share a similar motif composition (Additional file 3). For example, motif 9 is unique to group I, whereas motif 10 is specific to group IIa and IIb. The clustered AcWRKY pairs, i.e. AcWRKY47/35, AcWRKY51/53, showed highly similar motif distribution. The similar motif arrangements among AcWRKY proteins within subgroups indicated that the protein architecture is conserved within a specific subfamily. The functions of most of these conserved motifs remain to be elucidated. Overall, the conserved motif compositions and similar gene structures of the WRKY members in the same group, together with the phylogenetic analysis results, could strongly support the reliability of the group classifications.

Evolutionary analysis of group III WRKY genes in pineapple and several different species

The group III WRKY genes are thought to have originated after the divergence of the monocots and dicots, and seem to have played a key role in plant adaption and evolution [31]. Here, we further investigated the

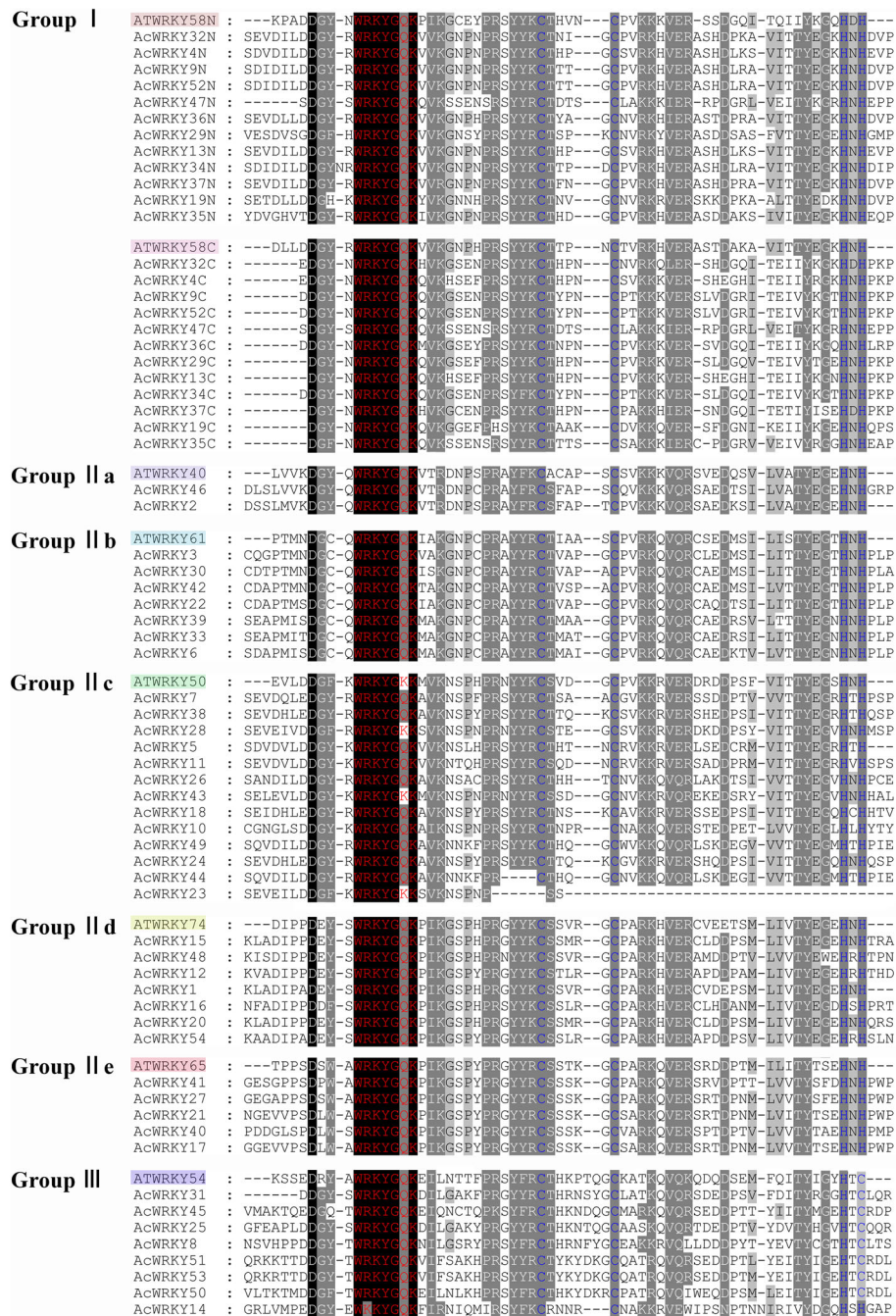


Fig. 1 Alignment of multiple AcWRKY and selected AtWRKY domain amino acid sequences. 'N' and 'C' indicate the N-terminal and C-terminal WRKY domain of a specific WRKY protein

duplication and diversification of group III genes during evolution, basing on the available pineapple WRKY III genes. A phylogenetic tree of WRKY III complete protein sequences from ten representative species, including six monocots (pineapple, rice, maize, banana, Brachypodium and millet) and four dicots (Arabidopsis, grape, tomato and poplar), was constructed using MEGA 5.0 [30–35].

As indicated in Fig. 4, the WRKY III proteins were divided in to seven clades by the phylogenetic tree. Each of the ten species contributed at least one WRKY III gene to Clade 1 and Clade 2, and these two clades were further divided into several different subclades. WRKY members from the phylogenetically closer species were clustered together within the two clades. For example, Clade 1a contained member from dicots, Clade 1b

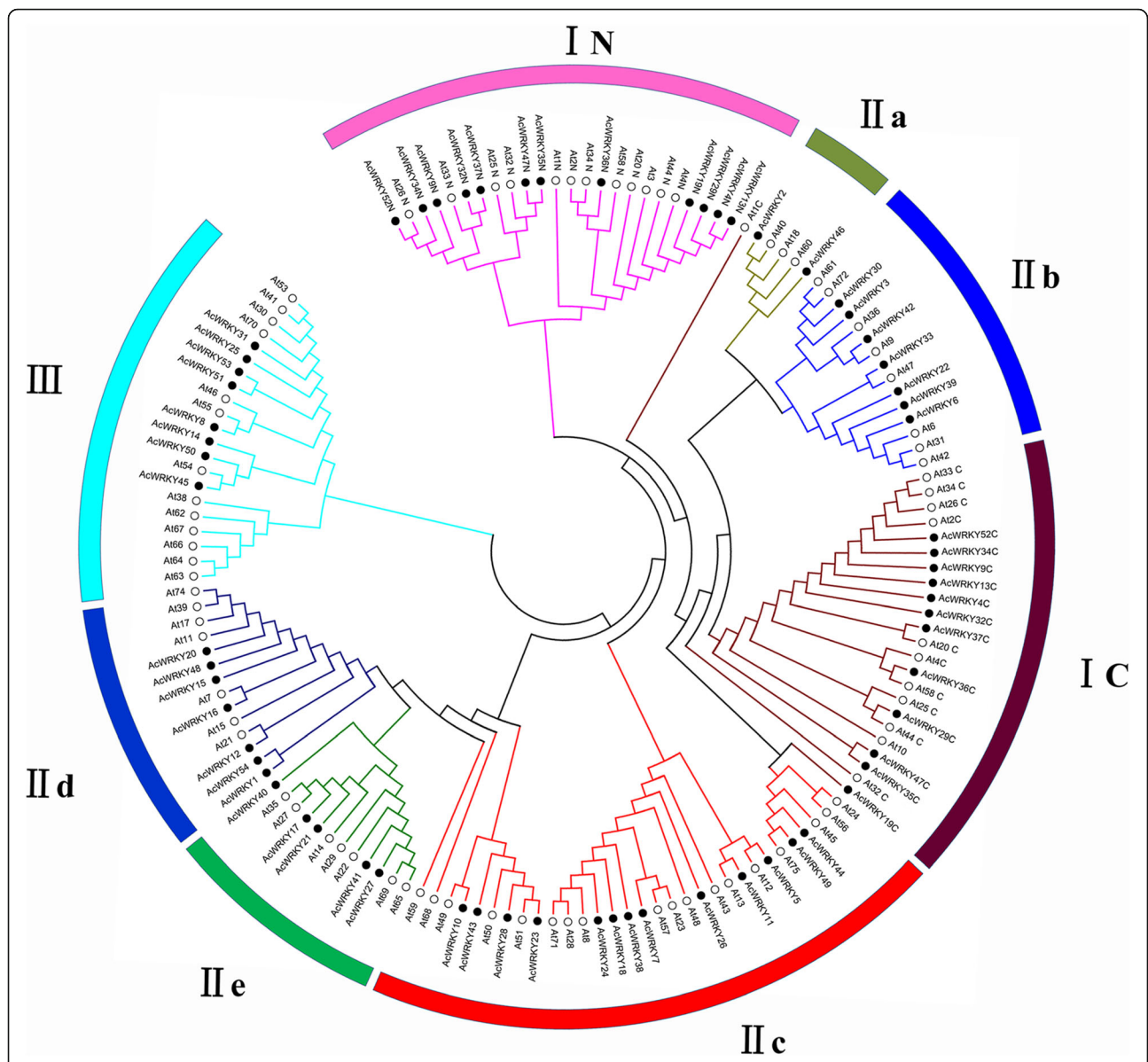


Fig. 2 Unrooted phylogenetic tree representing relationships among WRKY domains of pineapple and Arabidopsis. The different-colored arcs indicate different groups (or subgroups) of WRKY domains. Group I proteins with the suffix 'N' or 'C' indicates the N-terminal or the C-terminal WRKY domains. The black solid circles and hollow circles represent WRKY domain from pineapple and Arabidopsis, respectively. WRKY proteins from Arabidopsis with the prefix 'At' indicate 'AtWRKY'

possessed member from pineapple and banana, the non-grass monocots, whereas Clade 1c contained proteins only from the grass species. Additionally, Clade 3 contained the grass-species specific WRKY members. Nine WRKY genes from eight different species were included in Clade 4, indicating that they might be orthologues that originated from a single ancestral gene. Interestingly, AcWRKY14 was clustered together with a series of rice WRKY III proteins (in Clade 7), implying that the different evolutionary patterns of group III WRKY in rice and pineapple may occur after their divergence.

We also use the MEME web server to search the conserved motifs which were shared with the WRKY III proteins. A total of 20 distinct conserved motifs were found, motif 1, motif 2, motif 3 and motif 6 were found to encode the WRKY domain. As illustrated in Fig. 4, most WRKY members within the same clade, especially the most closely related members, usually shared common motif compositions (e.g. AcWRKY8 and MusaWRKY101), indicating potential functional similarities among WRKY proteins. Motif 15 was unique to the members in Clade 1, which may be important to the functions of unique WRKY III protein. Interestingly,

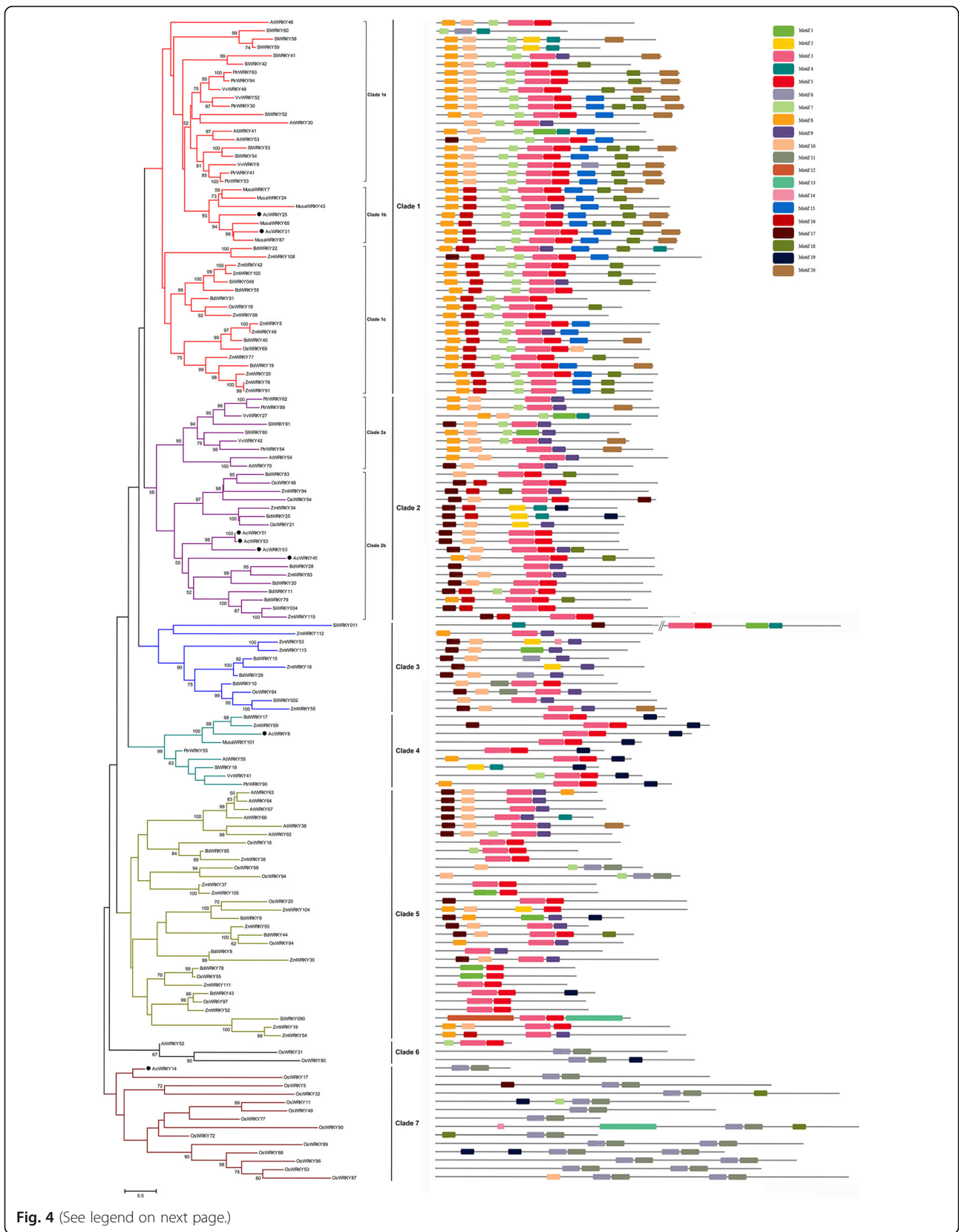


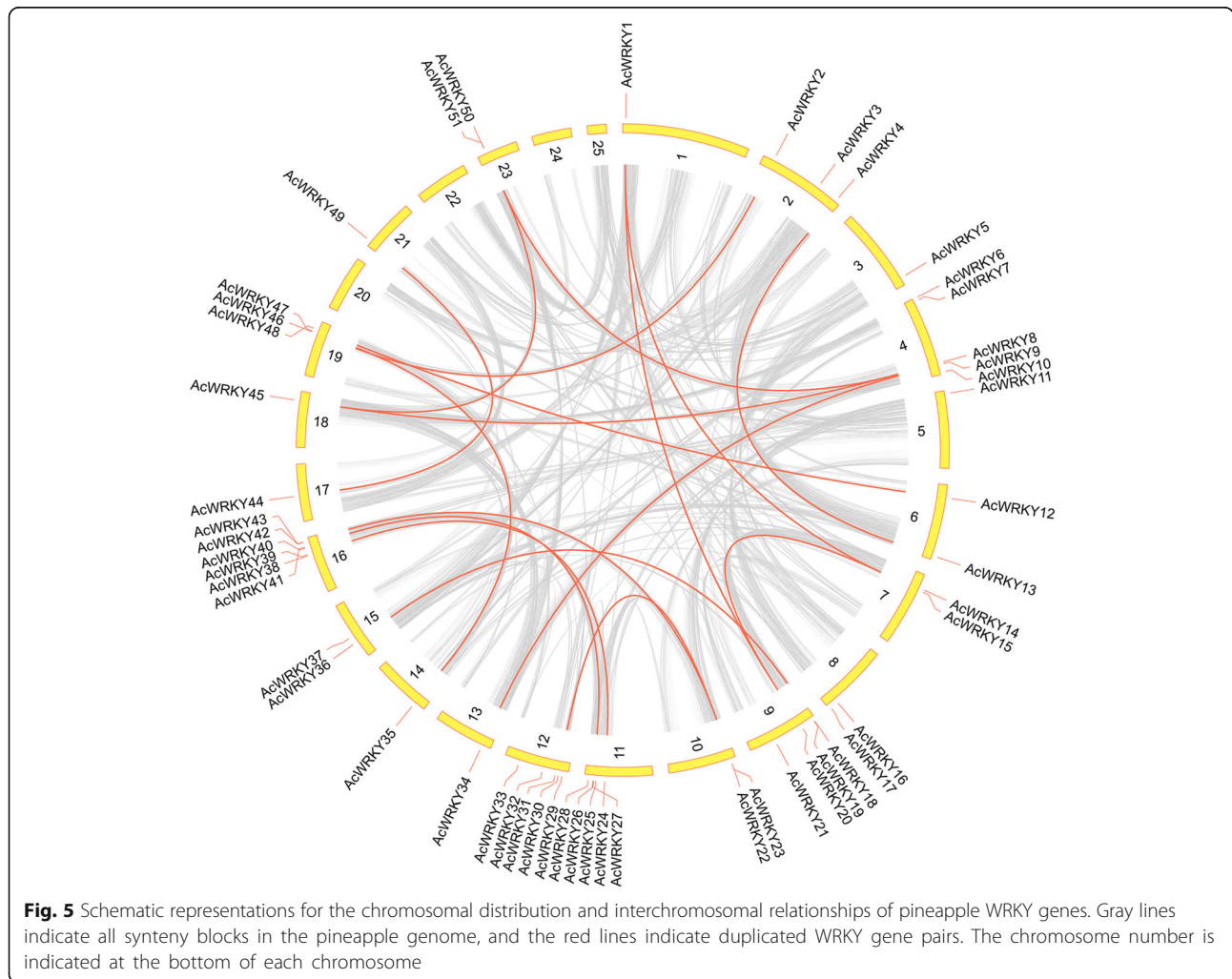
Fig. 4 (See legend on next page.)

(See figure on previous page.)

Fig. 4 Phylogenetic relationships and motif compositions of group III WRKY proteins from ten different plant species. Left panel: an unrooted phylogenetic tree constructed using MEGA 5.0 by the Neighbor-Joining method. The proteins are clustered into seven main clades with several subclades. Subtrees branch lines are colored indicating different clades. The black solid circles indicate group III WRKY proteins from pineapple. Right panel: distribution of conserved motifs in the group III WRKY proteins. The different-colored boxes represent different motifs and their position in each WRKY protein sequence

AcWRKY22/23, *AcWRKY26/27*, *AcWRKY38/39*, *AcWRKY42/43*, *AcWRKY46/47*, and *AcWRKY50/51*) were clustered into seven tandem duplication event regions on pineapple linkage group 09, 10, 11, 16, 19, and 23. LG16 had two clusters, indicating a hot spot of WRKY gene distribution. Besides the tandem duplication events, 17 segmental duplication events with 27 WRKY genes were also identified with BLASTP and MCScanX methods (Additional file 4). These results indicated that some *AcWRKY* genes were possibly generated by gene duplication and the segmental duplication events played a major driving force for *AcWRKY* evolution.

To further infer the phylogenetic mechanisms of pineapple WRKY family, we constructed five comparative syntentic maps of pineapple associated with five representative species, including two dicots (Arabidopsis and grape) and three monocots (banana, rice and maize) (Fig. 6). A total of 44 *AcWRKY* genes showed syntentic relationship with those in banana, followed by maize (39), rice (37), grape (33) and Arabidopsis (18) (Additional file 5). The numbers of orthologous pairs between the other five species (bananas, maize, rice, grape and Arabidopsis) were 145, 89, 56, 48 and 25. Some *AcWRKY* genes were found to be associated with at least three syntentic gene pairs (particularly between pineapple and bananas WRKY genes), such



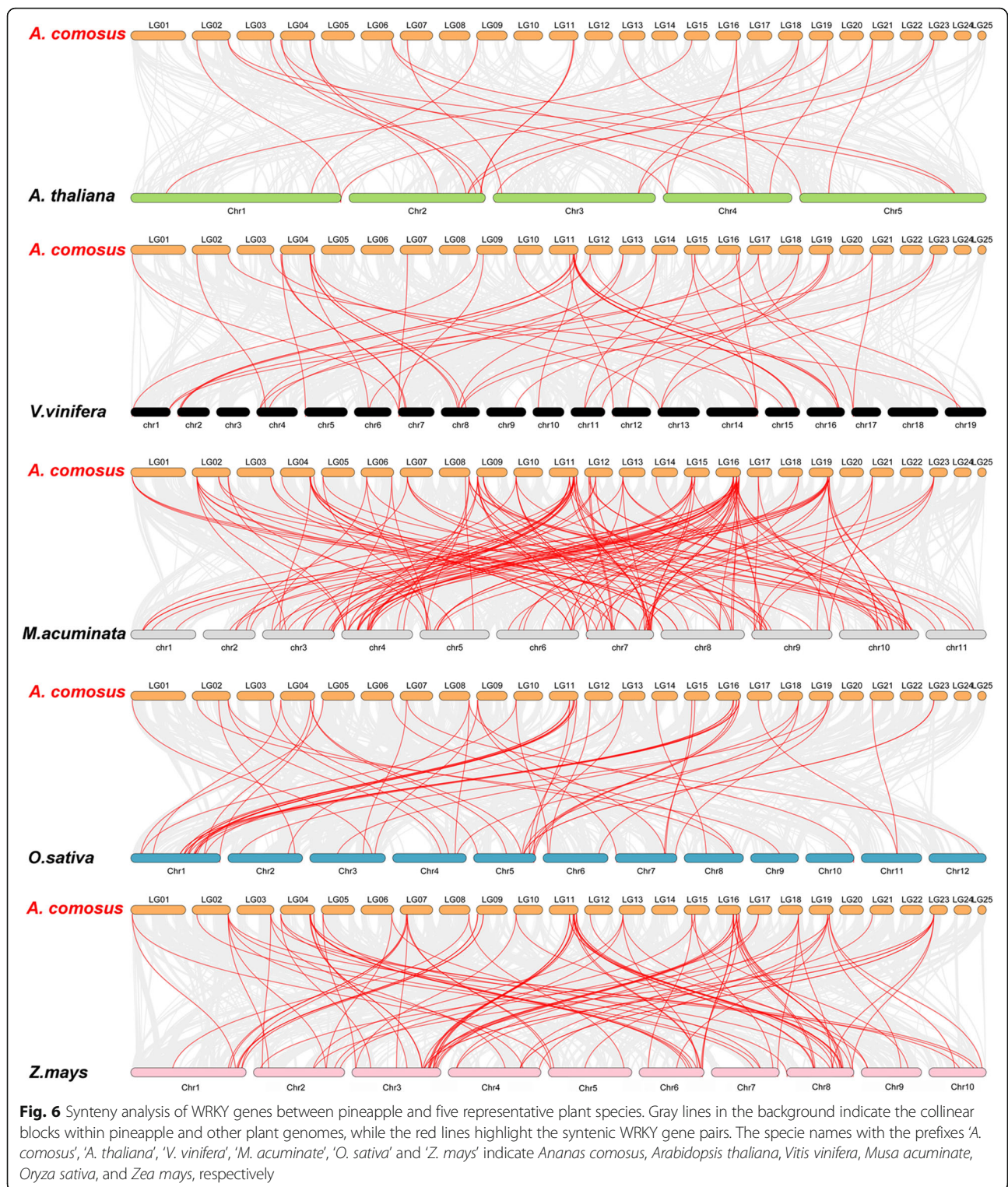


Fig. 6 Synteny analysis of WRKY genes between pineapple and five representative plant species. Gray lines in the background indicate the collinear blocks within pineapple and other plant genomes, while the red lines highlight the syntenic WRKY gene pairs. The species names with the prefixes 'A. comosus', 'A. thaliana', 'V. vinifera', 'M. acuminata', 'O. sativa' and 'Z. mays' indicate *Ananas comosus*, *Arabidopsis thaliana*, *Vitis vinifera*, *Musa acuminata*, *Oryza sativa*, and *Zea mays*, respectively

as *AcWRKY36* and *AcWRKY40*, guessed that these genes may have played an important role of WRKY gene family during evolution. Significantly, some WRKY collinear gene pairs identified between pineapple and rice were anchored to the highly conserved syntenic blocks, which spanning

more than 100 genes. In contrast, those between pineapple and Arabidopsis were all located in syntenic blocks that possessed less than 30 orthologous gene pairs. Similar patterns were also observed between pineapple and bananas/maize versus pineapple and grape, which may be

related to the phylogenetic relationship between pineapple and other five plant species.

Interestingly, some collinear gene pairs (with seven *AcWRKY* genes) identified between pineapple and rice/maize/bananas were not found between pineapple and Arabidopsis/grape, such as *AcWRKY51/MusaWRKY76*, *AcWRKY51/OsWRKY66*, which may indicate that these orthologous pairs formed after the divergence of dicotyledonous and monocotyledonous plants. Additionally, some collinear pairs (with 14 *AcWRKY* genes) were identified between pineapple and all of the other five species, indicating that these orthologous pairs may already exist before the ancestral divergence.

To better understand the evolutionary constraints acting on WRKY gene family, the Ka/Ks ratios of the WRKY gene pairs were calculated. All segmental and tandem duplicated *AcWRKY* gene pairs, and the majority of orthologous WRKY gene pairs had Ka/Ks < 1, suggesting that the pineapple WRKY gene family might have experienced strong purifying selective pressure during evolution.

Expression profiling of pineapple WRKY genes with RNA-seq

The expression patterns of all 54 *AcWRKY* genes in the transcriptome data, which was derived from different developmental stages of pineapple organs/tissues, were investigated in this study (Fig. 7a and Additional file 6). The reliability of the transcriptome data was further validated by quantitative real-time PCR (qRT-PCR) experiments which were carried out on eight representative samples for 14 selected WRKY genes (Fig. 7b). Among the 54 *AcWRKY* genes, *AcWRKY19* was not expressed in all detected samples, which may be pseudogenes or had special temporal and spatial expression patterns not examined in our libraries. Thirty-seven WRKY genes were expressed in all 30 samples tested (FPKM > 0) and 28 genes showed constitutive expression (FPKM > 1 in all samples). Some genes exhibited preferential expression across the detected tissues. Eleven genes in root, one gene in stem (*AcWRKY11*), three genes in leaf (*AcWRKY18/25/28*) and two genes in stamen (*AcWRKY4/22*) showed the highest transcript abundances. The expression of some genes exhibited significant trends in different development stages. For example, the expression levels of *AcWRKY4* and *AcWRKY25* were gradually reduced along with the fruit core development. The transcripts of *AcWRKY25/31/49* were gradually increased during different developmental stages of ovules.

The transcriptional levels of all 54 *AcWRKY* genes in different whole-fruit developmental stages were also investigated (transcriptome data from Ming et al. [29]), and the results showed that the expression of several WRKY genes were associated with the fruit development

(Additional file 7), such as *AcWRKY4/13* that with gradually decreased expression patterns. The diel expression analysis of the *AcWRKY* genes in pineapple photosynthetic (green tip) leaf tissues exhibited that three genes (*AcWRKY50/2/53*) had higher expression in dark compared to that in light period (Additional file 7). In case of expression profiles in cold stress library [37], significantly higher expression (Fold change > 2) of 11 *AcWRKY* genes (*AcWRKY28/50/40/48/46/2/24/45/52/23/9*) was observed in cold-stressed samples as compared to control (Fig. 7c).

Expression patterns of pineapple WRKY genes in response to different treatments

To further confirm whether the expression of *AcWRKY* genes was influenced by different abiotic stresses and hormonal treatments, 16 *AcWRKY* members, whose mRNA levels were relatively high across different tissues, were carefully selected from 54 pineapple WRKY genes. QRT-PCR experiments were further performed to analyze their expression patterns in response to different treatments (Figs. 8 and 9). Overall, some *AcWRKY* genes were significantly induced/repressed by multiple treatments. For instance, *AcWRKY18* significantly responded to SA (Salicylic acid), 2, 4-D (2, 4-Dichlorophenoxyacetic acid), cold and PEG (Polyethylene Glycol) treatments. *AcWRKY35* was induced by all tested treatments except cold stress. In contrast, multiple *AcWRKY* genes were simultaneously induced by one treatment. For example, seven *AcWRKY* genes (*AcWRKY9/18/25/32/45/51/52*) were induced by cold treatment, and five genes (*AcWRKY9/18/35/36/52*) were induced by 2, 4-D treatment. Interestingly, the transcript levels of many *AcWRKY* genes, such as *AcWRKY35*, *AcWRKY36* and *AcWRKY51*, were down-regulated by heat stress treatment. Several genes showed opposing expression patterns under different treatments. For instance, *AcWRKY13* was significantly induced by ABA (Abscisic Acid) and MeJA (Methyl Jasmonate), whereas was repressed by SA treatment.

Discussion

WRKY genes comprise a large family of transcription factors that are ubiquitous to all plant species. The genome-wide analysis of WRKY gene families have been widely carried out in many species whose genomes have been sequenced [35, 38–40]. In the current study, a search for WRKY genes in the pineapple genome resulted in the identification of 54 members, which were designated *AcWRKY1* through *AcWRKY54* on the basis of their chromosomal location.

The conserved structural domains of the pineapple WRKY proteins were assessed in this study. Multiple sequence alignments revealed that three *AcWRKY* proteins (*AcWRKY43*, *AcWRKY28*, and *AcWRKY23*) in

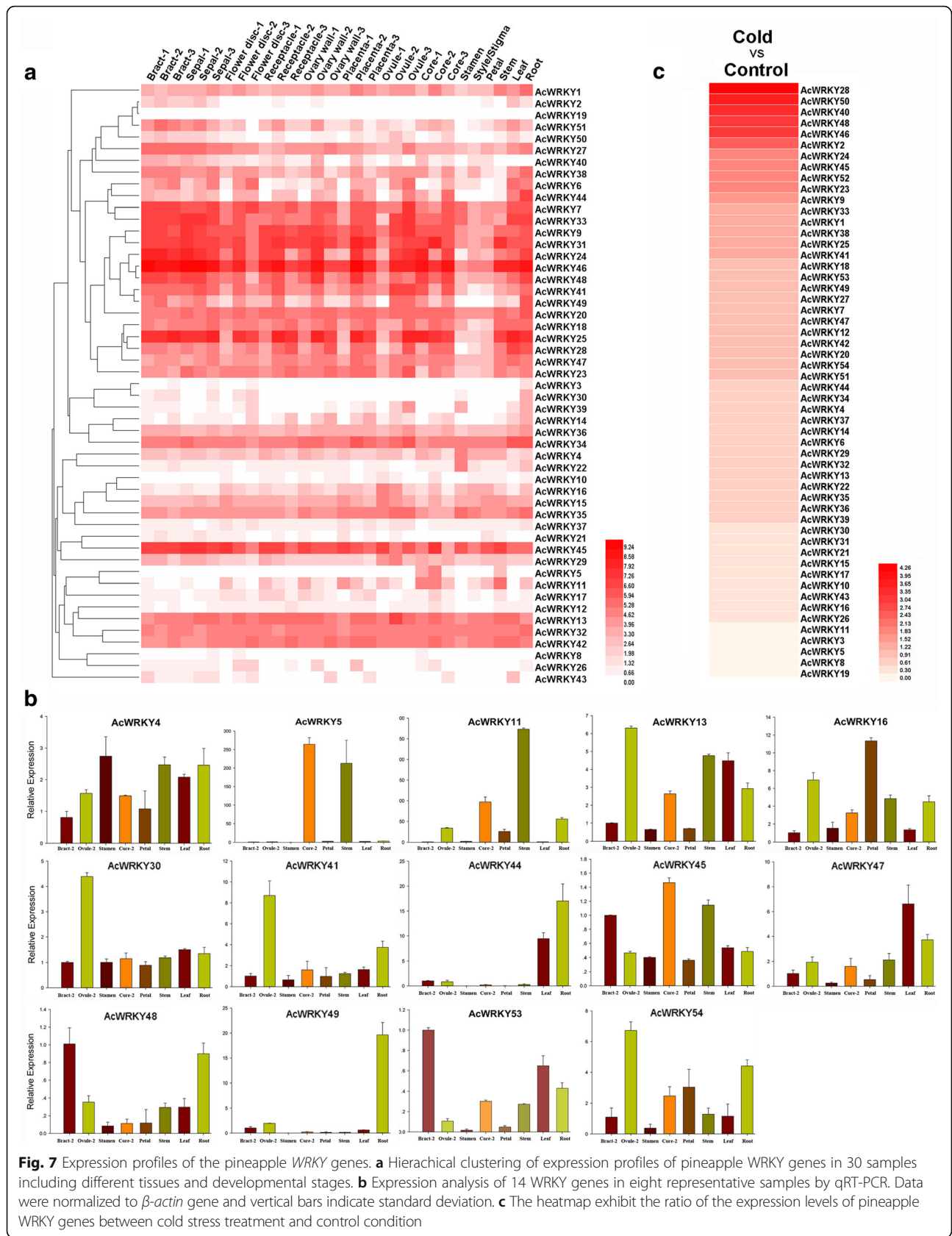


Fig. 7 Expression profiles of the pineapple *WRKY* genes. **a** Hierarchical clustering of expression profiles of pineapple *WRKY* genes in 30 samples including different tissues and developmental stages. **b** Expression analysis of 14 *WRKY* genes in eight representative samples by qRT-PCR. Data were normalized to β -actin gene and vertical bars indicate standard deviation. **c** The heatmap exhibit the ratio of the expression levels of pineapple *WRKY* genes between cold stress treatment and control condition

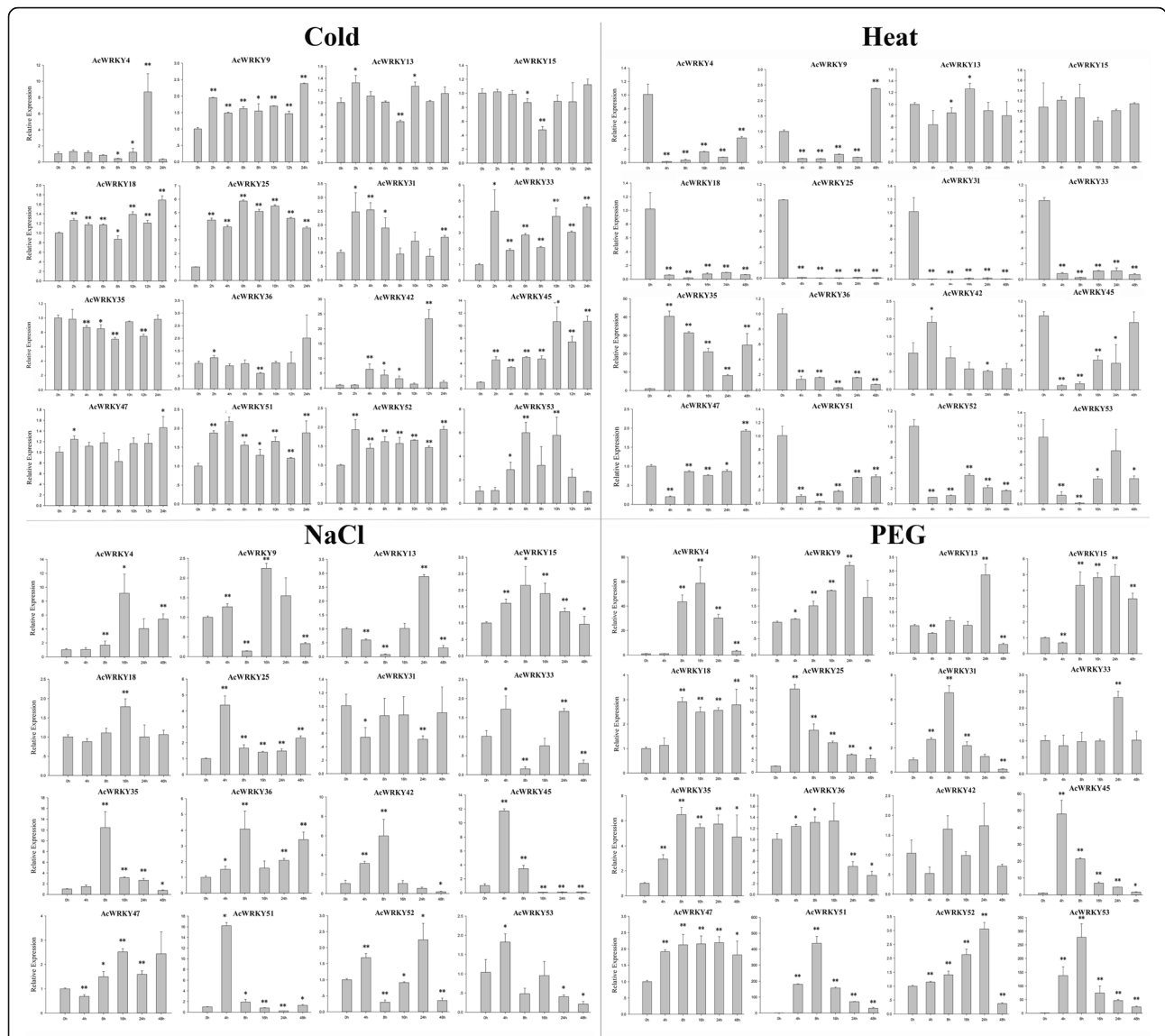


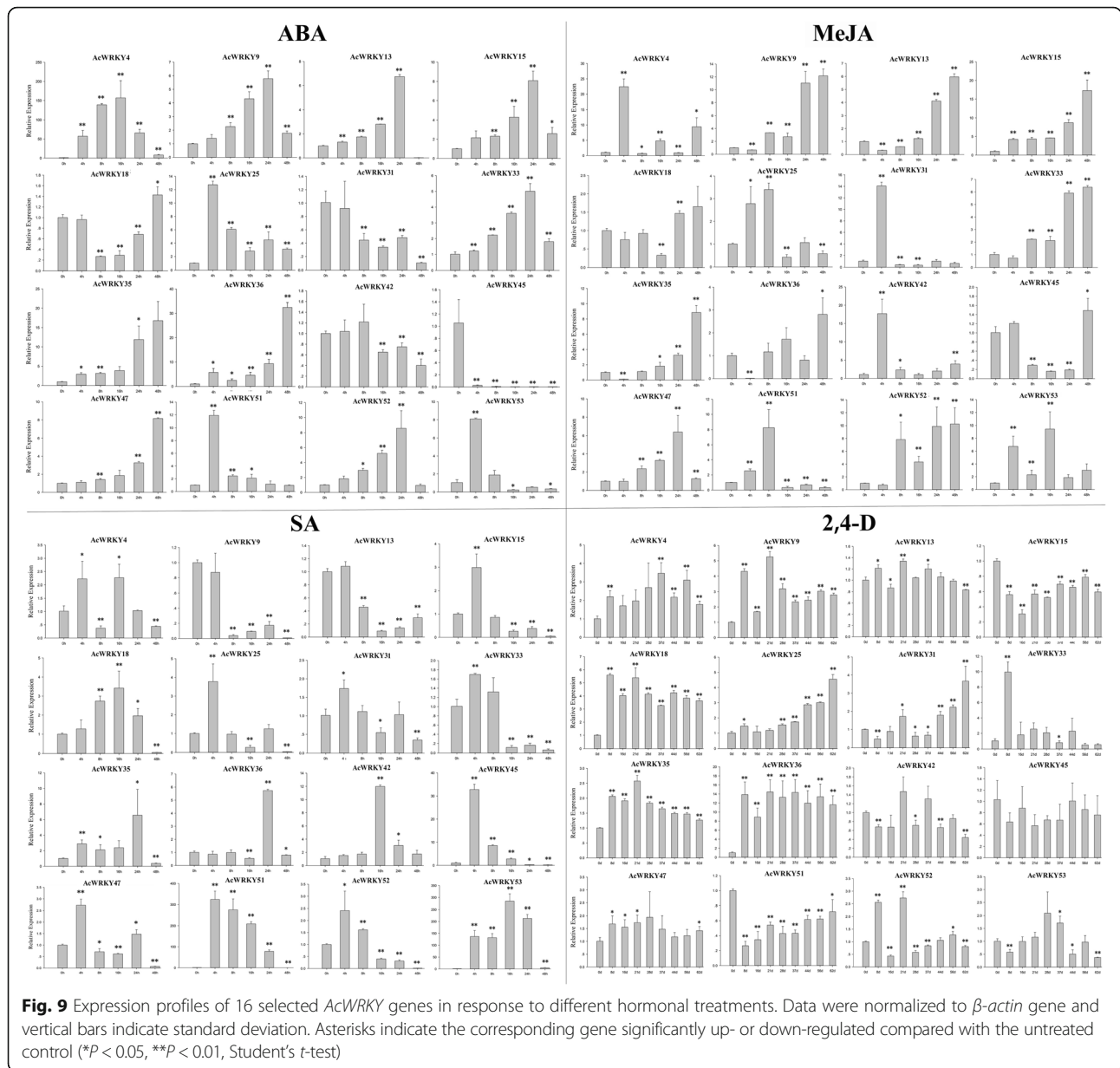
Fig. 8 Expression profiles of 16 selected *AcWRKY* genes in response to various abiotic stress treatments. Data were normalized to β -actin gene and vertical bars indicate standard deviation. Asterisks indicate the corresponding gene significantly up- or down-regulated compared with the untreated control (* $P < 0.05$, ** $P < 0.01$, Student's *t*-test)

group IIc had sequence variation in their WRKY domain. Most characterized WRKY proteins exhibited binding preference to their cognate *cis*-acting *W*-box element, with the help of WRKY domain. According to previous studies, variations in the WRKYGQK motif in WRKY domain might influence normal interactions of WRKY genes with downstream target genes, and therefore these three WRKY proteins might be worthy to further investigate their functions and binding specificities [41, 42].

The domain gain and loss is a divergent force for expansion of the WRKY gene family. The loss of WRKY domain seems to be common in many monocotyledons such as rice and maize [3, 32, 43]. However, WRKY

proteins from group I in pineapple all have two WRKY domains, and no domain loss events were found, suggesting the different characteristics of this group during pineapple evolution. It is reported that the N-terminal WRKY domains showed weak DNA-binding activity and were more variable during evolution. Consisted with previous studies, the pineapple WRKY trees clustered the N-terminal WRKY domains as a monophyletic subtree (Fig. 2), suggesting that the single WRKY domains of group II and III family members are more closely related to the C-terminal domains of group I than to the N-terminal domains [2, 32].

Comparison of the number of *WRKY* genes in pineapple with other sequenced monocot genomes has shown that



pineapple possesses comparatively lesser number of genes [34, 43–45]. Tandem and segmental duplication events have played a critical role in the expansion of WRKY gene family [4]. The whole-genome duplication events are common during angiosperm evolution and usually lead to the expansion of gene families [46]. The σ whole-genome duplication (WGD) event was shared with all Poales, while ρ WGD event is inferred to have occurred after divergence of lineages leading to the grasses and pineapple within the Poales [29]. Therefore, lacking the pan-grass ρ WGD event during pineapple evolution might be a possible reason for the smaller amount of *AcWRKY* genes.

Moreover, the variation in the number of group III WRKY genes was also a potential cause of the diversity

of WRKY gene family size [31]. The previous studies described group III WRKY genes as being the most dynamic group with respect to gene family evolution. In this study, eight group III *AcWRKY* genes were fewer than that in most other monocots, consistent with the smaller amount of pineapple WRKY family members. Different from the group III WRKY genes in many other monocots, that in pineapple have not undergone a lineage-specific radiation, which may be caused by different pattern of duplication events [30].

Among the WRKY gene family, group III genes were considered as the most advanced in terms of evolution, and seem to have played an important role in plant adaptation and evolution [31]. The phylogenetic analysis

of the WRKY III proteins could provide more clues about the evolution history of the WRKY gene family [31]. In our study, the plant WRKY III members from the closer related species were tended to be clustered together. Consistent with several previous studies, both monocots and dicots members were present in many clades, indicating that WRKY III genes diversified before the divergence of monocots and dicots [4, 31, 33]. WRKY III genes formed monocot- and dicot-specific subclades depicting that the WRKY genes have evolved independently after the monocot-dicot split. The pineapple lineage diverged from the lineage leading to grasses early in the history of Poales, and become an outgroup and evolutionary reference for the study of the lineage-specific gene family mobility in grasses [47]. The presence of the grasses-specific clade (Clade 3) indicated that the WRKY III genes may have been expanded independently in grasses after the divergence of pineapple and grasses during the Poales evolution. The lineage-specific expansions of WRKY III members in several grass species such as rice and *Brachypodium* further support this opinion [30, 43]. Similarly, the different evolutionary characteristics were also observed after the divergence of the eurosid group I and II [48].

There is considerable evidence that WRKY genes play significant roles in regulating plant growth and development, and in conferring tolerance to abiotic stresses including salinity, drought, heat, cold and wounding [6, 7, 49]. The Crassulacean acid metabolism plants possess high water-use efficiency and the CAM photosynthesis also confers tolerance to plants against abiotic stress, particularly to drought stress. The genome-sequenced CAM species, pineapple, which was shared conserved syntenic relationships with several important cereal species (like rice and sorghum), has been regarded as an important crop for studying CAM photosynthesis and abiotic stress tolerance [29]. In view of the importance of pineapple in abiotic stress biology and the key roles of WRKY genes in physiological processes and stress responses, the expression patterns of *AcWRKY* genes were investigated in this study, basing on the available transcriptome data and qRT-PCR analysis in response to different treatments. By combining gene expression, phylogenetic and synteny analysis, new clues to the biological function of pineapple WRKY genes could be inferred through comparison with those function-known WRKY genes from model plants.

Some valuable clues about the functional role of *AcWRKY* genes that involved in specific pineapple physiological process were obtained. For example, *AcWRKY7* exhibited the highest expression in mature ovule tissues, while its orthologs in Arabidopsis, *AtWRKY23*, could mediate the embryo development [16], indicating that

AcWRKY7 may share similar functions in pineapple. *AcWRKY5* was specially expressed in stem and fruit core tissues, and with extremely low expression levels in other detected pineapple tissues. Interestingly, its orthologs in Arabidopsis, *AtWRKY12*, was also expressed in stem pith and cortex, where it regulates the secondary cell wall formation [50]. Accordingly, we inferred that *AcWRKY5* may also participate in secondary cell wall formation in corresponding tissues. The pineapple fruit core was developed from inflorescence axis and its firmness was gradually decreased along with the fruit development [26], thus *AcWRKY5* might affect the edible taste of pineapple fruit. *AcWRKY4* was highly expressed in stamen, and have orthologous relationship with two pollen-specific regulators in Arabidopsis, *AtWRKY2* and *AtWRKY34*, indicating that *AcWRKY4* may have similar roles in the pollen developmental modulation [51]. In pineapple photosynthetic leaf tissues, *AcWRKY2* have relative higher expression in dark compared to that in light period (Additional file 7), while its orthologs in Arabidopsis, *AtWRKY40*, was found to be a repressor of high-light-induced signaling and involved in chloroplast dysfunction [52], indicating that *AcWRKY2* may be also associated with the regulation of the high-light stress.

The functional roles of some *AcWRKY* genes which were associated with the various abiotic stresses were also inferred. Low temperature is one of the major environmental stresses that affect pineapple growth and development, and the cold injury usually lead to decreases in crop yield and quality [26, 53]. According to the cold stress transcriptome data [37], the expressions of 11 *AcWRKY* genes were significantly induced. The expression patterns of *AcWRKY9* and *AcWRKY52* were further verified by qRT-PCR analysis (Fig. 8), and they also have phylogenetically closest relationship with *AtWRKY33*, a typical cold-responsive WRKY gene in Arabidopsis, implying their possible roles in pineapple cold stress regulation [54]. The expression of *OsWRKY76* was induced by low temperature, overexpression of *OsWRKY76* in rice plants improved tolerance to cold stress [55], and its orthologs in pineapple, *AcWRKY2*, was also induced by cold stress, suggesting the potential value of *AcWRKY2* in pineapple cold-resistance improvement. Conserved cold-responsive expression features were also observed in orthologous gene pairs like *AcWRKY46-AtWRKY40/18* and *AcWRKY4-AtWRKY34*, suggesting the functional conservation of WRKY genes during evolution [11, 56]. The outstanding drought tolerance was widely found in many CAM species represented by pineapple, and WRKY transcription factors were important components of the drought-stress regulatory network [7]. The drought stress responsive gene *AcWRKY18* was orthologous to *AtWRKY57*, which conferred drought tolerance in Arabidopsis [57], indicating similar function of

AcWRKY18 in drought stress regulation. Accumulating evidence indicated that WRKY genes were regulated by hormones like SA and ABA, and play crucial roles in the hormone signaling network [6, 11]. *AcWRKY9* and its orthologs in Arabidopsis, *AtWRKY25*, were both induced by salt and ABA treatments, implying their similar roles in conferring salt tolerance and ABA signaling network [58]. Consistent with previous studies in other species [6], our current studies showed that some *AcWRKY* genes were differentially expressed following various abiotic stresses and hormone treatments, highlighting the extensive involvement of WRKY genes in environmental adaptation.

It was noteworthy that the expression patterns were not completely consistent between *AcWRKY* genes and their counterparts. For example, *AcWRKY15* and its orthologs in Arabidopsis, *AtWRKY39*, exhibited opposite expression features in response to heat stress [23]. Divergence of gene expression plays an important role in the preservation of duplicated genes. Several pairs of paralogs have different expression patterns, suggesting that they may play diverse roles in pineapple development. For instance, *AcWRKY4* was preferentially expressed in stamen and cold-responsive, while its paralogous gene *AcWRKY13* was highly expressed across different tissues and cold-insensitive. However, some WRKY genes and their paralogues, like *AcWRKY35* and *AcWRKY47*, shared similar transcript abundance profiles, suggesting that they may have redundant functions.

Overall, these above findings provide insight into the potential functional roles of pineapple WRKY genes. The comprehensive analyses were helpful in selecting candidate WRKY genes for further functional characterization, and for the genetic improvement in the agronomic characters and environmental resistance of pineapple.

Conclusions

A comprehensive analysis of WRKY gene family in pineapple was carried out in the present study. Fifty four full-length WRKY genes were characterized and further classified into three main groups, with high similar exon-intron structures and motif compositions within the same groups and subgroups. Synteny analysis and phylogenetic comparison of WRKY genes from several different plant species provided valuable clues about the evolutionary characteristics of pineapple WRKY genes. *AcWRKY* genes played important roles in pineapple growth and development as indicated by their expression patterns in different tissues and in response to various treatments. The phylogenetic and gene expression analysis will shed light on the functional analysis of *AcWRKY* genes. These results provide a valuable resource for better understanding the biological roles of individual WRKY genes in pineapple.

Methods

Gene identification

The hidden Markov model (HMM) file corresponding to the WRKY domain (PF03106) was downloaded from Pfam protein family database (<http://pfam.sanger.ac.uk/>). HMMER 3.0 was used to search the WRKY genes from pineapple genome database. The default parameters were adopted and the cutoff value was set to 0.01. All candidate genes that may contain WRKY domain based on HMMER results were further examined by confirming the existence of the WRKY core sequences using the PFAM and SMART program. Each potential gene was then manually examined to ensure the conserved heptapeptide sequence at the N-terminal region of the predicted WRKY domain. To further check the annotation of the predicted WRKY gene models, available pineapple RNA-seq reads were mapped back to the pineapple genome assembly and gene models, using Tophat2 and Cufflinks [59, 60]. The incorrectly predicted genes were then manually curated and some of them were further validated by PCR amplification and sequencing. The redundant sequences were manually discarded. Fifty-four WRKY gene models were finally identified in the pineapple genome after comprehensive curation. Length of sequences, molecular weights, isoelectric points and subcellular location prediction of identified WRKY proteins were obtained by using tools from ExPasy website (<http://web.expasy.org/protparam/>).

Sequence analysis

The WRKY domain sequences of the characterized WRKY proteins were used to create multiple protein sequence alignments using ClustalW with default parameters. The deduced amino acid sequences in WRKY domains were then adjusted manually using GeneDoc software. The exon-intron organization of pineapple WRKY genes was determined by comparing predicted coding sequences with their corresponding full-length sequences using the online program Gene Structure Display Server (GSDS: <http://gsds.cbi.pku.edu.cn>) [61]. The MEME online program (<http://meme.nbcr.net/meme/intro.html>) for protein sequence analysis was used to identify conserved motifs in the identified pineapple WRKY proteins [62]. The optimized parameters were employed as the following: the number of repetitions, any; the maximum number of motifs, 20; and the optimum width of each motif, between 6 and 100 residues.

Chromosomal distribution and gene duplication

All *AcWRKY* genes were mapped to pineapple chromosomes based on physical location information from the database of pineapple genome using Circos [63]. Multiple Collinearity Scan toolkit (MCScanX) was adopted

to analyze the gene duplication events, with the default parameters [64]. To exhibit the synteny relationship of the orthologous *WRKY* genes obtained from pineapple and other selected species, the syntenic analysis maps were constructed using the Dual Synteny Plotter software (<https://github.com/CJ-Chen/TBtools>) written by ourselves [65]. Non-synonymous (ka) and synonymous (ks) substitution of each duplicated *WRKY* genes were calculated using KaKs_Calculator 2.0 [66].

Phylogenetic analysis and classification of pineapple *WRKY* gene family

All identified pineapple *WRKY* genes were divided into different group according to the At*WRKY* classification scheme and the alignment *WRKY* domains of Ac*WRKY* and At*WRKY* proteins. The phylogenetic trees were inferred using Neighbor-Joining (NJ) method of MEGA 5.0, with the following parameters: Poisson model, pairwise deletion, and 1000 bootstrap replications. Sequence of *WRKY* proteins from Arabidopsis, maize [32], rice [43, 67], grape [39], banana [33], and Brachypodium [44] were obtained based on the description in corresponding literatures and downloaded from the phytozome database (<https://phytozome.jgi.doe.gov/>).

Plant materials and treatments

Ananas comosus cv. Shenwan, a typical cultivated variety, was used throughout the study. The bract, sepal, flower disc, receptacle, ovary walls, placenta, ovule, fruit core from three different developmental stages of pineapple fruit, the stamen, style, petal, stem, leaf and root of mature pineapple plants, were collected separately for RNA extraction and used for further RNA-seq and qRT-PCR analysis. To investigate the expression pattern in response to various stress and hormonal treatments, several Ac*WRKY* genes were selected for further qRT-PCR analysis. For salinity and drought treatments, the callus tissues were subjected to a 150 mM NaCl and 15% PEG6000 solution, respectively, for 4, 8, 12, 24 and 48 h. For phytohormone analysis, the calluses were respectively cultured in MS liquid medium supplied with 100 μ M ABA, SA and MeJA for 4, 8, 12, 24 and 48 h. For 2, 4-D treatments, the calluses were transferred into the MS solid medium with 4 mg/L 2, 4-D. Samples were collected at 8, 16, 21, 28, 37, 44, 56 and 62 days after treatments. For heat and cold stress treatments, the pineapple plantlets were subjected to 40 and 4 $^{\circ}$ C, respectively. The leaves were collected at 2, 4, 6, 8, 10, 12, 24 h in cold treatment and 4, 8, 12, 24, 48 h in heat treatment. All treated tissue samples were immediately frozen in liquid nitrogen and stored at -80° C for subsequent analysis.

RNA extraction and gene expression analysis

Total RNA was extracted using Trizol method as described by Ma et al. [68]. All RNA was analyzed by

agrose gel electrophoresis and then quantified with a Nanodrop ND-1000 spectrophotometer. DNA-free RNA was used for synthesis of first strand of cDNA by using HiScript[®] II 1st Strand cDNA Synthesis Kit (Vazyme) as per manufacturer's recommendations. The quantitative RT-PCR was carried out with the Roche Lightcycler[®] 480 instrument using SYBR Green chemistry. The house-keeping pineapple β -actin gene was used as an internal control. The reaction was carried out as follows: 95 $^{\circ}$ C for 30s, followed by 40 cycles of 95 $^{\circ}$ C, /10 s, 60 $^{\circ}$ C, /30 s. Each reaction was performed in biological triplicates and the data from real-time PCR amplification was analyzed using $2^{-\Delta\Delta CT}$ method. Sequences of the primers used in this study were shown in detail in Additional file 8. Details about the transcriptome data derived from various pineapple tissues were described in Liu et al. [64], and the transcriptome analysis of pineapple under cold stress have been carried out in Chen et al. [37]. The transcript abundance of pineapple *WRKY* genes was calculated as fragments per kilobase of exon model per million mapped reads (FPKM). The heatmaps were created by Heml1.0 based on the transformed data of \log_2 (FPKM+1) values [69]. The transcriptome data used in this study could also be obtained on the website constructed by ourselves (http://118.24.17.128/html/Pineapple_Expression_DB_By_HeLab_AT_SCAU).

Additional files

Additional file 1: List of the 54 Ac*WRKY* genes identified in this study. (XLSX 60 kb)

Additional file 2: Phylogenetic and HMMER analyses of the R protein-*WRKY* families. Left panel: Neighbor-Joining phylogenetic tree derived from the alignment of full length R protein-*WRKY* family members. Numbers indicate bootstrap values from 1000 replicates. Right panel: the HMMER-derived overview of protein architecture with predicted protein domains. The length of protein can be estimated using the scale at the bottom. (TIF 279 kb)

Additional file 3: Analysis and distribution of conserved motifs in pineapple *WRKY* proteins. (XLSX 11 kb)

Additional file 4: Segmentally and tandemly duplicated Ac*WRKY* gene pairs. (XLSX 10 kb)

Additional file 5: One-to-one orthologous relationships between pineapple and other five plant species. (XLSX 46 kb)

Additional file 6: RNA-seq data of 54 Ac*WRKY* genes that were used in this study. (XLSX 90 kb)

Additional file 7: Expression profiles of pineapple *WRKY* genes in different samples. (A) Expression profiles of pineapple *WRKY* genes in the RNA-seq data derived from different whole-fruit developmental stages. (B) Expression profiles of pineapple *WRKY* genes in the RNA-seq data derived from the pineapple green tip leaf tissues at 2-h intervals over a 24-h period. (TIF 2603 kb)

Additional file 8: Sequences of the primers used in this study. (XLSX 11 kb)

Abbreviations

2, 4-D: 2, 4-Dichlorophenoxyacetic acid; ABA: Abscisic Acid; Ac*WRKY*: Pineapple (*Ananas comosus*) *WRKY*; At*WRKY*: *Arabidopsis thaliana* *WRKY*; MeJA: Methyl Jasmonate; PEG: Polyethylene Glycol; SA: Salicylic acid

Acknowledgments

The authors thank to lab members for assistance.

Funding

This study was supported by the Technology Commission of Guangdong Province (2013B020304002), Foundation of Young Creative Talents in Higher Education of Guangdong Province (2017KQNCX020), National Natural Science Foundation of China (31572089), and Modern Agricultural Industry Technology System of Guangdong Province (2016LM1128). The funders had no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials

All data analysed during this study are included in this article and its Additional files.

Authors' contributions

TX and JRL performed the experiments. CJC, CHL analyzed the data. TX and CYL wrote the manuscript. CYL and YHH designed the study. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

Ananas comosus cv. Shenwan is widely cultivated in Guangdong province, China. *Ananas comosus* is not listed in the appendices I, II and III of the Convention on the Trade in Endangered Species of Wild Fauna and Flora, that has been valid from 4 October 2017 (<https://cites.org/eng/app/appendices.php>). The pineapple samples used in this study were collected from horticultural germplasm conservation center of South China Agricultural University (SCAU). Collection of plant materials complied with the institutional, national and international guidelines. No specific permits were required.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Key Laboratory of Biology and Germplasm Enhancement of Horticultural Crops in South China, Ministry of Agriculture, South China Agricultural University, Guangzhou 510642, China. ²College of Horticulture, South China Agricultural University, Guangzhou 510642, China.

Received: 28 August 2017 Accepted: 18 June 2018

Published online: 25 June 2018

References

- Ulker B, Somssich IE. WRKY transcription factors: from DNA binding towards biological function. *Curr Opin Plant Biol.* 2004;7(5):491–8.
- Eulgem T, Rushton PJ, Robatzek S, Somssich IE. The WRKY superfamily of plant transcription factors. *Trends Plant Sci.* 2000;5(5):199–206.
- Brand LH, Fischer NM, Harter K, Kohlbacher O, Wanke D. Elucidating the evolutionary conserved DNA-binding specificities of WRKY transcription factors by molecular dynamics and in vitro binding assays. *Nucleic Acids Res.* 2013;41(21):9764–78.
- Zhang Y, Wang L. The WRKY transcription factor superfamily: its origin in eukaryotes and expansion in plants. *BMC Evol Biol.* 2005;5:1.
- Ishiguro S, Nakamura K. Characterization of a cDNA encoding a novel DNA-binding protein, SPF1, that recognizes SP8 sequences in the 5' upstream regions of genes coding for sporamin and β -amylase from sweet potato. *Mol Gen Genet.* 1994;244(6):563–71.
- Phukan UJ, Jeena GS, Shukla RK. WRKY transcription factors: molecular regulation and stress responses in plants. *Front Plant Sci.* 2016;7:760.
- Jiang J, Ma S, Ye N, Jiang M, Cao J, Zhang J. WRKY transcription factors in plant responses to stresses. *J Integr Plant Biol.* 2017;59(2):86–101.
- Levee V, Major I, Levasseur C, Tremblay L, MacKay J, Seguin A. Expression profiling and functional analysis of *Populus* WRKY23 reveals a regulatory role in defense. *New Phytol.* 2009;184(1):48–70.
- Kloth KJ, Wieggers GL, Busscher-Lange J, van Haarst JC, Kruijer W, Bouwmeester HJ, Dicke M, Jongsma MA. AtWRKY22 promotes susceptibility to aphids and modulates salicylic acid and jasmonic acid signalling. *J Exp Bot.* 2016;67(11):3383–96.
- Pandey SP, Somssich IE. The role of WRKY transcription factors in plant immunity. *Plant Physiol.* 2009;150(4):1648–55.
- Banerjee A, Roychoudhury A. WRKY proteins: signaling and regulation of expression during abiotic stress responses. *Sci World J.* 2015;2015:807560.
- Liu H, Yang W, Liu D, Han Y, Zhang A, Li S. Ectopic expression of a grapevine transcription factor VvWRKY11 contributes to osmotic stress tolerance in *Arabidopsis*. *Mol Biol Rep.* 2011;38(1):417–27.
- Devaiah BN, Karthikeyan AS, Raghothama KG. WRKY75 transcription factor is a modulator of phosphate acquisition and root development in *Arabidopsis*. *Plant Physiol.* 2007;143(4):1789–99.
- Jiang Y, Liang G, Yang S, Yu D. *Arabidopsis* WRKY57 functions as a node of convergence for jasmonic acid- and auxin-mediated signaling in jasmonic acid-induced leaf senescence. *Plant Cell.* 2014;26(1):230–45.
- Huang Y, Feng CZ, Ye Q, Wu WH, Chen YF. *Arabidopsis* WRKY6 transcription factor acts as a positive regulator of abscisic acid signaling during seed germination and early seedling development. *PLoS Genet.* 2016;12(2):e1005833.
- Grunewald W, De Smet I, De Rybel B, Robert HS, van de Cotte B, Willemsen V, Gheyssen G, Weijers D, Friml J, Beeckman T. Tightly controlled WRKY23 expression mediates *Arabidopsis* embryo development. *EMBO Rep.* 2013;14(12):1136–42.
- Yu Y, Hu R, Wang H, Cao Y, He G, Fu C, Zhou G. MiWRKY12, a novel Miscanthus transcription factor, participates in pith secondary cell wall formation and promotes flowering. *Plant Sci.* 2013;212:1–9.
- Li W, Wang H, Yu D. *Arabidopsis* WRKY transcription factors WRKY12 and WRKY13 oppositely regulate flowering under short-day conditions. *Mol Plant.* 2016;9(11):1492–503.
- Ye J, Wang X, Hu T, Zhang F, Wang B, Li C, Yang T, Li H, Lu Y, Giovannoni JJ, et al. An InDel in the promoter of Al-activated malate transporter 9 selected during tomato domestication determines fruit malate contents and aluminum tolerance. *Plant Cell.* 2017; <https://doi.org/10.1105/tpc.17.00211>.
- Mirabella R, Rauwerda H, Allmann S, Scala A, Spyropoulou EA, de Vries M, Boersma MR, Breit TM, Haring MA, Schuurink RC. WRKY40 and WRKY6 act downstream of the green leaf volatile E-2-hexenal in *Arabidopsis*. *Plant J.* 2015;83(6):1082–96.
- Amato A, Cavallini E, Zenoni S, Finezzo L, Begheldo M, Ruperti B, Tornielli GB. A grapevine TTG2-like WRKY transcription factor is involved in regulating vacuolar transport and flavonoid biosynthesis. *Front Plant Sci.* 2016;7:1979.
- Gonzalez A, Brown M, Hatlestad G, Akhavan N, Smith T, Hembd A, Moore J, Montes D, Mosley T, Resendez J. TTG2 controls the developmental regulation of seed coat tannins in *Arabidopsis* by regulating vacuolar transport steps in the proanthocyanidin pathway. *Dev Biol.* 2016;419(1):54–63.
- Li S, Zhou X, Chen L, Huang W, Yu D. Functional characterization of *Arabidopsis thaliana* WRKY39 in heat stress. *Mol Cells.* 2010;29(5):475–83.
- Zhang J, Peng Y, Guo Z. Constitutive expression of pathogen-inducible OsWRKY31 enhances disease resistance and affects root growth and auxin response in transgenic rice plants. *Cell Res.* 2008;18(4):508–21.
- Rinerson CI, Rabara RC, Tripathi P, Shen QJ, Rushton PJ. The evolution of WRKY transcription factors. *BMC Plant Biol.* 2015;15:66.
- Lobo MG, Paull RE. Handbook of pineapple technology: postharvest science, processing and nutrition. Chichester: John Wiley & Sons; 2017.
- Bartholomew DP, Paull RE, Rohrbach KG. The pineapple: botany, production, and uses. Wallingford: CABI; 2002.
- Ming R, Wai CM, Guyot R. Pineapple genome: a reference for monocots and CAM photosynthesis. *Trends Genet.* 2016;32(11):690–6.
- Ming R, VanBuren R, Wai CM, Tang H, Schatz MC, Bowers JE, Lyons E, Wang ML, Chen J, Biggers E, et al. The pineapple genome and the evolution of CAM photosynthesis. *Nat Genet.* 2015;47(12):1435–42.
- Tripathi P, Rabara RC, Langum TJ, Boken AK, Rushton DL, Boomsma DD, Rinerson CI, Rabara J, Reese RN, Chen X. The WRKY transcription factor family in *Brachypodium distachyon*. *BMC Genomics.* 2012;13(1):270.
- Wang Y, Feng L, Zhu Y, Li Y, Yan H, Xiang Y. Comparative genomic analysis of the WRKY III gene family in populus, grape, *Arabidopsis* and rice. *Biol Direct.* 2015;10:48.

32. Wei KF, Chen J, Chen YF, Wu LJ, Xie DX. Molecular phylogenetic and expression analysis of the complete WRKY transcription factor family in maize. *DNA Res.* 2012;19(2):153–64.
33. Goel R, Pandey A, Trivedi PK, Asif MH. Genome-wide analysis of the Musa WRKY gene family: evolution and differential expression during development and stress. *Front Plant Sci.* 2016;7:299.
34. Muthamilarasan M, Bonthala VS, Khandelwal R, Jaishankar J, Shweta S, Nawaz K, Prasad M. Global analysis of WRKY transcription factor superfamily in *Setaria* identifies potential candidates involved in abiotic stress signaling. *Front Plant Sci.* 2015;6:910.
35. Huang S, Gao Y, Liu J, Peng X, Niu X, Fei Z, Cao S, Liu Y. Genome-wide analysis of WRKY transcription factors in *Solanum lycopersicum*. *Mol Gen Genomics.* 2012;287(6):495–513.
36. Holub EB. The arms race is ancient history in *Arabidopsis*, the wildflower. *Nat Rev Genet.* 2001;2(7):516.
37. Chen C, Zhang Y, Xu Z, Luan A, Mao Q, Feng J, Xie T, Gong X, Wang X, Chen H. Transcriptome profiling of the pineapple under low temperature to facilitate its breeding for cold tolerance. *PLoS One.* 2016;11(9):e0163315.
38. Huang X, Li K, Xu X, Yao Z, Jin C, Zhang S. Genome-wide analysis of WRKY transcription factors in white pear (*Pyrus bretschneideri*) reveals evolution and patterns under drought stress. *BMC Genomics.* 2015;16:1104.
39. Guo C, Guo R, Xu X, Gao M, Li X, Song J, Zheng Y, Wang X. Evolution and expression analysis of the grape (*Vitis vinifera* L.) WRKY gene family. *J Exp Bot.* 2014;65(6):1513–28.
40. He H, Dong Q, Shao Y, Jiang H, Zhu S, Cheng B, Xiang Y. Genome-wide survey and characterization of the WRKY gene family in *Populus trichocarpa*. *Plant Cell Rep.* 2012;31(7):1199–217.
41. van Verk MC, Pappaioannou D, Neeleman L, Bol JF, Linthorst HJ. A novel WRKY transcription factor is required for induction of *PR-1a* gene expression by salicylic acid and bacterial elicitors. *Plant Physiol.* 2008;146(4):1983–95.
42. Zhou QY, Tian AG, Zou HF, Xie ZM, Lei G, Huang J, Wang CM, Wang HW, Zhang JS, Chen SY. Soybean WRKY-type transcription factor genes, GmWRKY13, GmWRKY21, and GmWRKY54, confer differential tolerance to abiotic stresses in transgenic *Arabidopsis* plants. *Plant Biotechnol J.* 2008; 6(5):486–503.
43. Ross CA, Liu Y, Shen QJ. The WRKY gene family in rice (*Oryza sativa*). *J Integr Plant Biol.* 2007;49(6):827–42.
44. Wen F, Zhu H, Li P, Jiang M, Mao W, Ong C, Chu Z. Genome-wide evolutionary characterization and expression analyses of WRKY family genes in *Brachypodium distachyon*. *DNA Res.* 2014;21(3):327–39.
45. Kaliyappan R, Viswanathan S, Suthanthiram B, Subbaraya U, Marimuthu Somasundram S, Muthu M. Evolutionary expansion of WRKY gene family in banana and its expression profile during the infection of root lesion nematode, *Pratylenchus coffeae*. *PLoS One.* 2016;11(9):e0162013.
46. Cannon SB, Mitra A, Baumgarten A, Young ND, May G. The roles of segmental and tandem gene duplication in the evolution of large gene families in *Arabidopsis thaliana*. *BMC Plant Biol.* 2004;4(1):10.
47. Xu Q, Liu ZJ. A taste of pineapple evolution through genome sequencing. *Nat Genet.* 2015;47(12):1374–6.
48. Ling J, Jiang W, Zhang Y, Yu H, Mao Z, Gu X, Huang S, Xie B. Genome-wide analysis of WRKY gene family in *Cucumis sativus*. *BMC Genomics.* 2011;12(1):471.
49. Rushton PJ, Somssich IE, Ringler P, Shen QJ. WRKY transcription factors. *Trends Plant Sci.* 2010;15(5):247–58.
50. Wang H, Avci U, Nakashima J, Hahn MG, Chen F, Dixon RA. Mutation of WRKY transcription factors initiates pith secondary wall formation and increases stem biomass in dicotyledonous plants. *P Natl Acad Sci USA.* 2010; 107(51):22338–43.
51. Lei R, Li X, Ma Z, Lv Y, Hu Y, Yu D. *Arabidopsis* WRKY2 and WRKY34 transcription factors interact with VQ20 protein to modulate pollen development and function. *Plant J.* 2017;91(6):962–76.
52. Van Aken O, Zhang B, Law S, Narsai R, Whelan J. AtWRKY40 and AtWRKY63 modulate the expression of stress-responsive nuclear genes encoding mitochondrial and chloroplast proteins. *Plant Physiol.* 2013;162(1):254–71.
53. Py C, Lacoëuilhe JJ, Teisson C. The pineapple. Cultivation and uses: G.-P. Maisonneuve et Larose; 1987.
54. Fu Q, Yu D. Expression profiles of AtWRKY25, AtWRKY26 and AtWRKY33 under abiotic stresses. *Yi Chuan.* 2010;32(8):848–56.
55. Yokotani N, Sato Y, Tanabe S, Chujo T, Shimizu T, Okada K, Yamane H, Shimono M, Sugano S, Takatsuji H, et al. WRKY76 is a rice transcriptional repressor playing opposite roles in blast disease resistance and cold stress tolerance. *J Exp Bot.* 2013;64(16):5085–97.
56. Bakshi M, Oelmüller R. WRKY transcription factors. *Plant Signal Behav.* 2014; 9(2):e27700.
57. Jiang Y, Liang G, Yu D. Activated expression of WRKY57 confers drought tolerance in *Arabidopsis*. *Mol Plant.* 2012;5(6):1375–88.
58. Jiang Y, Deyholos MK. Functional characterization of *Arabidopsis* NaCl-inducible WRKY25 and WRKY33 transcription factors in abiotic stresses. *Plant Mol Biol.* 2009;69(1–2):91–105.
59. Kim D, Perrea G, Trapnell C, Pimentel H, Kelley R, Salzberg SL. TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. *Genome Biol.* 2013;14(4):R36.
60. Trapnell C, Roberts A, Goff L, Pertea G, Kim D, Kelley DR, et al. Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and cufflinks. *Nat Protoc.* 2012;7(3):562–78.
61. Guo AY, Zhu QH, Chen X, Luo JC. GSDS: a gene structure display server. *Yi Chuan.* 2007;29(8):1023.
62. Bailey TL, Boden M, Buske FA, Frith M, Grant CE, Clementi L, Ren J, Li WW, Noble WS. MEME SUITE: tools for motif discovery and searching. *Nucleic Acids Res.* 2009; <https://doi.org/10.1093/nar/gkp335>.
63. Krzywinski M, Schein J, Birol I, Connors J, Gascoyne R, Horsman D, Jones SJ, Marra MA. Circos: an information aesthetic for comparative genomics. *Genome Res.* 2009;19(9):1639–45.
64. Wang Y, Tang H, DeBarry JD, Tan X, Li J, Wang X, Lee T-h, Jin H, Marler B, Guo H. MScanX: a toolkit for detection and evolutionary analysis of gene synteny and collinearity. *Nucleic Acids Res.* 2012;40(7):e49.
65. Liu C, Xie T, Chen C, Luan A, Long J, Li C, Ding Y, He Y. Genome-wide organization and expression profiling of the R2R3-MYB transcription factor family in pineapple (*Ananas comosus*). *BMC Genomics.* 2017;18(1):503.
66. Wang D, Zhang Y, Zhang Z, Zhu J, Yu J. KaKs_Calculator 2.0: a toolkit incorporating gamma-series methods and sliding window strategies. *Genomics Proteomics Bioinformatics.* 2010;8(1):77–80.
67. Group RWW. Nomenclature report on rice WRKY's-conflict regarding gene names and its solution. *Rice.* 2012;5(1):3.
68. Ma J, He Y, Wu C, Liu H, Hu Z, Sun G. Cloning and molecular characterization of a *SERK* gene transcriptionally induced during somatic embryogenesis in *Ananas comosus* cv. Shenwan. *Plant Mol Biol Rep.* 2011; 30(1):195–203.
69. Deng W, Wang Y, Liu Z, Cheng H, Xue Y. Heml: a toolkit for illustrating heatmaps. *PLoS One.* 2014;9(11):e111988.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

