

# Development and Validation of Amoxicillin by RP-HPLC Method in Bulk drug and Pharmaceutical dosage forms

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**Abstract:** A new, simple, specific, sensitive, rapid, accurate and precise RP-HPLC method was developed for the estimation of Amoxicillin in bulk and pharmaceutical formulations. Amoxicillin was chromatographed on a hypersil C18 column (250x4.6mm I.D., particle size 5 µm) in a mobile phase consisting of potassium dihydrogen phosphate and methanol in the ratio 95:05 v/v. The mobile phase was pumped at a flow rate of 1.0 ml/min with detection at 283 nm. The detector response was linear in the concentration of 20-100 µg/ml. The intra and inter day variation was found to be less than 2%. The mean recovery of the drug from the solution was 99.39%. The proposed method is simple, fast, accurate, precise and reproducible hence, it can be applied for routine quality control analysis of Amoxicillin in bulk and pharmaceutical formulations.

**Keywords:** RP-HPLC, Amoxicillin, precision, accuracy.

## INTRODUCTION

Amoxicillin<sup>1</sup> is chemically (2*S*,5*R*,6*R*)-6-[[*(2R)*-2-amino-2-(4-hydroxyphenyl)-acetyl] amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo [3.2.0] heptane-2-carboxylic acid. It is freely soluble in water. The molecular formula is C<sub>16</sub>H<sub>19</sub>N<sub>3</sub>O<sub>5</sub>S and Molecular weight is 419.45<sup>1</sup>. It is official drug in Indian pharmacopoeia<sup>2</sup>, British Pharmacopoeia<sup>3</sup>. Amoxicillin is a moderate-spectrum bacteriolytic β-lactum antibiotic used to treat bacterial infections caused by susceptible micro organisms. It is usually the drug of choice within the class because it is better absorbed following oral administration. Amoxicillin acts by inhibiting the synthesis of bacterial cell wall. It is soluble in water, methanol, slightly soluble in ethanol and partially insoluble in toluene<sup>4</sup>. Literature survey reveals that, only few spectrophotometric methods and few analytical methods have been reported for the

quantitative estimation of Amoxicillin in bulk drug and pharmaceutical formulation. Hence an attempt has been made to develop new HPLC methods for its estimation in bulk and pharmaceutical formulation with good precision, accuracy, linearity and reproducibility.

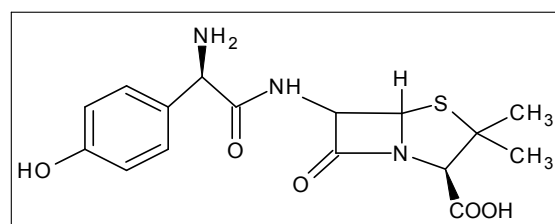


Figure 1. Chemical structure of Amoxicillin

## MATERIALS AND METHODS

An isocratic high pressure liquid chromatography (shimadzu HPLC winchrome - ES Series) with one LC-10 AT VP pumps, with UV/VIS detector SPD-10A VP, CTS-10 AS VP column oven (shimadzu), and a hypersil C-18 Column 250 mm x 4.6 mm i.d. particle size 5  $\mu\text{m}$ ) was used. The HPLC system was equipped with the software class class-vp (Shimadzu).

## REAGENTS AND CHEMICALS

All the chemicals used were of HPLC grade and A.R. grade. Distilled water was used for making the solutions. The commercially available Amoxicillin tablets were procured from the local market.

## CHROMATOGRAPHIC CONDITIONS

The content of the mobile phase was potassium dihydrogen phosphate and methanol in the ratio(95:05v/v). The mobile phase was filtered through 0.45  $\mu\text{m}$  membrane filter and sonicated for 15 min. The flow rate of the mobile phase was maintained at 1.0 ml/min. The column temperature was set ambient and the detection was carried out by UV-detector wavelength at 254 nm. The run time was set at 10 min and the volume of the injection loop was 20  $\mu\text{L}$ . Prior to injection of the drug solution, the column was equilibrated for atleast 30 min with the mobile phase flowing through the system.

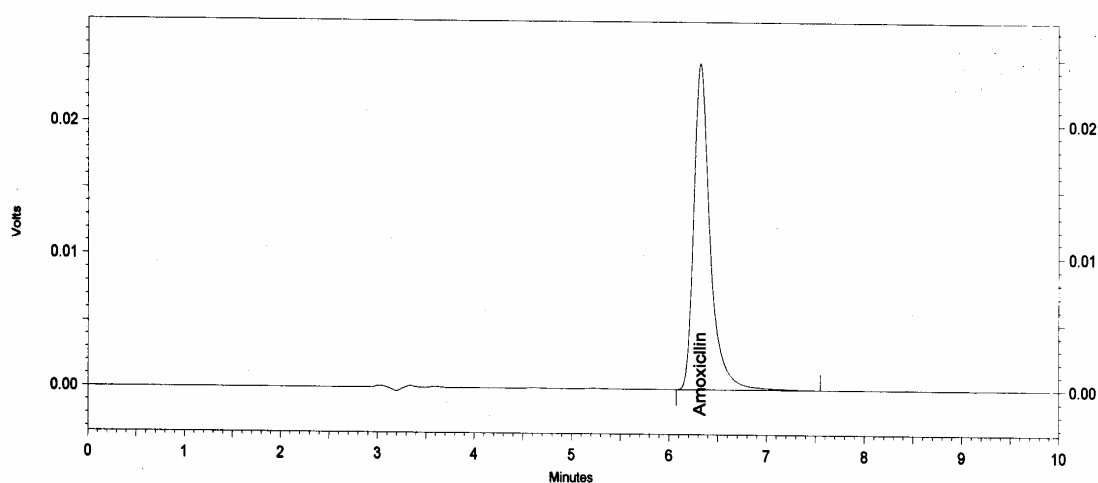
## PROCEDURE

Stock solution of Amoxicillin was prepared by dissolving 100 mg of Amoxicillin in 100 ml standard volumetric flask containing approximately 50 ml of mobile phase and the solution was sonicated for 20 min and then the volume was made upto the mark with mobile phase to obtain a concentration of 1000  $\mu\text{g/ml}$ . Subsequent dilutions of this solution were made with mobile phase to obtain the concentration range of 20-100  $\mu\text{g/ml}$ . The standard solutions prepared as above were injected into the 20  $\mu\text{L}$  loop and the chromatogram was recorded and shown in **Figure 2**.

The retention time of Amoxicillin was found to be 6.30 min. The calibration curve was constructed by plotting concentration against peak area ratio. The calibration curve is found to be linear and shown in Figure 3. The amount of Amoxicillin present in sample was calculated through the standard calibration curve. The linearity experiment was carried out in triplicate to ascertain accuracy and precision of the method. The peak area ratios of the drug against concentration are found to be linear and the results are expressed in **Table 1**.

**Table 1. Calibration data of the method**

| Concentration ( $\mu\text{g/ml}$ ) | Peak area |
|------------------------------------|-----------|
| 0                                  | 0         |
| 20                                 | 58544     |
| 40                                 | 117011.7  |
| 60                                 | 175632.3  |
| 80                                 | 234154.7  |
| 100                                | 286059.7  |



**Figure 2. Typical chromatogram of Amoxicillin**

**ASSAY**

25 tablets each containing 250 mg of Amoxicillin weighed accurately and powdered. A quantity equivalent to 100 mg of Amoxicillin was weighed accurately and transferred to 100 ml volumetric flask containing approximately 50 ml of mobile phase. The contents were sonicated for 20 min and volume was made upto the mark with the mobile phase. The resulting solution was filtered through a membrane filter. The solution obtained was then diluted with the mobile phase so as to obtain a concentration of 1000 µg/ml. Sample solution was injected under the same chromatographic conditions and the chromatogram was recorded in triplicate. The amount of Amoxicillin present in tablet formulation was determined by comparing the peak area from the standard. The results are furnished in **Table 2**.

**VALIDATION OF PROPOSED METHOD<sup>5-9</sup>**

Selectivity of the method was assessed on the basis of elution of Amoxicillin using the above mentioned chromatographic conditions. This study was validated for linearity, precision, accuracy, limit of detection, limit of quantitation and robustness. The results are furnished in **Table 3**.

**LINEARITY**

The standard curve was obtained in the concentration range of 20-100 µg/mL. The linearity was evaluated by

linear regression analysis using the least square method. It was found that correlation coefficient and regression analysis are within the limits.

**PRECISION**

The precision was assessed in terms of intra-day and inter-day variation. The intra-day and inter-day variation in the peak area of drug solution was calculated in terms of coefficient of variation (C.V.). The results are furnished in **Table 4**.

**LIMIT OF DETECTION (LOD) AND LIMIT OF QUANTITATION (LOQ)**

The LOD and LOQ for Amoxicillin were predicted basing on the parameters of standard error of estimate and slope, calculated from linearity of the response data of Amoxicillin.

**ROBUSTNESS**

The robustness was checked by changing the flow rate to 0.9 and 1.1 ml/min.

**ACCURACY**

The accuracy of the HPLC method was assessed by adding known amount of standard drug solution to a pre-analyzed tablet formulation. The recovery studies were carried out in triplicate. The accuracy was expressed in terms of recovery at three levels 80%, 100% and 120%. The results are furnished in **Table 5**.

**Table 2. Assay of Amoxicillin**

| Components | Mean* | Standard Deviation* | Co-efficient of Variation* | Standard Error* |
|------------|-------|---------------------|----------------------------|-----------------|
| AMOX       | 99.74 | 0.110574            | 0.110855                   | 0.0.045143      |

\*n = 6

**Table 3. System suitability parameters**

| Parameters                               | RP-HPLC method |
|--|----------------|
| Linearity range (µg/ml)                  | 20-100 µg/ml   |
| Slope                                    | 2879           |
| Intercept                                | 1251           |
| Regression coefficient (r <sup>2</sup> ) | 0.9996         |
| Limit of Detection (µg/ml)               | 0.4139         |
| Limit of Quantification (µg/ml)          | 1.2545         |
| Retention time (min)                     | 6.30           |
| Tailing factor                           | 1.46           |
| Theoretical plate                        | 7695           |

**Table 4. Precision of the proposed HPLC method**

| Concentration<br>(µg/ml) | Mean*        |              | Standard<br>Deviation* |              | Co-efficient of<br>Variation* |              | Standard<br>Error* |              |
|--------------------------|--------------|--------------|------------------------|--------------|-------------------------------|--------------|--------------------|--------------|
|                          | Intra<br>day | Inter<br>day | Intra<br>day           | Inter<br>day | Intra<br>day                  | Inter<br>day | Intra<br>day       | Inter<br>day |
| 40                       | 39.55        | 39.59        | 0.1216                 | 0.0484       | 0.3075                        | 0.1224       | 0.0702             | 0.0279       |
| 60                       | 59.91        | 59.74        | 0.1422                 | 0.0499       | 0.2374                        | 0.0836       | 0.0821             | 0.0288       |
| 80                       | 79.90        | 79.78        | 0.0611                 | 0.0244       | 0.0764                        | 0.0306       | 0.0352             | 0.0141       |

\*n = 6

**Table 5. Recovery studies of the proposed HPLC method**

| Sr No | List of<br>% Recovery | Mean*   | Standard<br>Deviation* | Co-efficient<br>of variation |
|-------|-----------------------|---------|------------------------|------------------------------|
| 1     | 80                    | 99.2600 | 1.0452                 | 1.0530                       |
| 2     | 100                   | 99.5367 | 0.1102                 | 0.1106                       |
| 3     | 120                   | 99.3733 | 0.3761                 | 0.3784                       |

n=3

## RESULTS AND DISCUSSION

Optimization of the chromatographic conditions were carried out with various combinations of buffer a methanol and by observing the peak parameters, the run time of the method was set at 10 min, Amoxicillin appeared on the typical chromatogram at 6.30 min, which indicates a good base line. When the same drug solution was injected 3 times, the retention time of the drug was same. Linearity range was observed in the concentration range of 20-100 µg/ml. The regression equation of Amoxicillin concentration over its peak area ratio was found to be  $Y = 2859X + 1751$  ( $r=0.9996$ ) where Y is the peak area ratio and X is the concentration of Amoxicillin (Fig. 3). The proposed HPLC method was also validated for intra-day and inter-day variation. The coefficient of variation in the peak area of the drug for 3 replicate injections was found to be less than 2%. The tailing factor was found to be 1.4, which indicates good shape of peak. The number of theoretical plates was found to be 7695,

which indicates efficient performance of the column. The limit of detection and limit of quantitation was found to be 0.4139µg/ml and 1.2545 µg/ml which indicates the sensitivity of the method. The use of buffer and methanol in the ratio of 95:05 v/v resulted in peak with good shape and resolution. The high percentage of recovery of Amoxicillin ranging from 99.26-99.53 indicates that the proposed method is highly accurate. No interfering peaks were found in the chromatogram indicating that excipients used in tablet formulation did not interfere with the estimation of the drug by proposed HPLC method.

## CONCLUSION

The proposed HPLC method was found to be simple, rapid, sensitive, precise and accurate for the estimation of Amoxicillin in pharmaceutical formulations. Hence, this method can easily and conveniently be adopted for routine quality control analysis of Amoxicillin in bulk and pharmaceutical formulations.

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